

US011206843B1

(12) **United States Patent**
Strickland

(10) **Patent No.:** **US 11,206,843 B1**
(45) **Date of Patent:** ***Dec. 28, 2021**

(54) **MILK PRODUCT COMPOSITIONS**

(71) Applicant: **BIOMILQ, Inc.**, Durham, NC (US)

(72) Inventor: **Leila Strickland**, Hillsborough, NC (US)

(73) Assignee: **BIOMILQ, Inc.**, Durham, NC (US)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

This patent is subject to a terminal disclaimer.

(21) Appl. No.: **17/301,216**

(22) Filed: **Mar. 29, 2021**

Related U.S. Application Data

(60) Provisional application No. 63/200,480, filed on Mar. 9, 2021, provisional application No. 63/030,149, filed on May 26, 2020.

(51) **Int. Cl.**
C12N 5/071 (2010.01)
A23C 9/20 (2006.01)
C12N 5/078 (2010.01)

(52) **U.S. Cl.**
CPC **A23C 9/203** (2013.01); **A23C 9/206** (2013.01); **C12N 5/0631** (2013.01); **C12N 5/0634** (2013.01); **C12N 2501/315** (2013.01); **C12N 2502/095** (2013.01); **C12N 2502/11** (2013.01); **C12N 2513/00** (2013.01)

(58) **Field of Classification Search**
CPC C12N 5/0631; C12N 2502/095; C12N 2502/11; C12N 2513/00; C12N 2501/315
See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

6,358,920 B1	3/2002	Blaschuk et al.
2013/0344490 A1	12/2013	Kim
2015/0079584 A1	3/2015	Gevaert et al.
2017/0267970 A1	9/2017	Gupta et al.
2018/0066220 A1	3/2018	Nath et al.
2019/0211296 A1	7/2019	Allbritton
2021/0207090 A1	7/2021	Strickland

FOREIGN PATENT DOCUMENTS

EP	0911389 A2	4/1999
WO	WO-20210142241 A1	7/2021

OTHER PUBLICATIONS

Watson. Immunological Functions of the Mammary Gland and Its Secretion-Comparative Review. Aust. J. Biol. Sci., 1980, 33, 403-22 (Year: 1980).*

Gourdou et al. Development of a Constitutively Active Mutant Form of the Prolactin Receptor, a Member of the Cytokine Receptor Family. Molecular Endocrinology. 1996. vol. 10, No. 1, p. 45-56 (Year: 1996).*

Shennan et al. Transport of Milk Constituents by the Mammary Gland. Physiological Reviews vol. 80, No. 3, Jul. 2000 (Year: 2000).*

Arèvalo Turrubiarte, M., et al., Phenotypic and functional characterization of two bovine mammary epithelial cell lines in 2D and 3D models, Am J Physiol, 310: C348-C356, (2016).

Blatchford D.R., et al., Milk Secretion in Cultured Mammary Epithelial Cells, In: Kitagawa, Y., et al. (eds), Animal Cell Technology: Basic & Applied Aspects, Springer, Dordrecht, 10: 141-145 (1999).

Chen, G., et al., Isolation, culture, and differentiation of mammary epithelial stem/progenitor cells from fresh or ex vivo cultured human breast tissue, Curr Protoc Cell Biol, 82(1): e65 (2019).

Cho, et al., Constructions of a 3D mammary duct based on spatial localization of the extracellular matrix, NPG Asia Mat, 10:970-981 (2018).

Co-Pending U.S. Appl. No. 17/247,672, filed Dec. 18, 2020.

Morada, M., et al., Continuous culture of Cryptosporidium parvum using hollow fiber technology, Int J for Parasitol, 46 (1): 21-29 (2016).

Sharfstein, S.T., et al., Functional Differentiation and Primary Metabolism of Mouse Mammary Epithelial Cells in Extended-Batch and Hollow-Fiber Culture, Biotechnol and Bioeng, 40(6): 672-680 (1992).

U.S. Appl. No. 17/247,672 Office Action dated Apr. 5, 2021.

Yonemura, S., Differential sensitivity of epithelial cells to extracellular matrix in polarity establishment, PLOS One, 9 (11): e112922 (2014).

Gourdou, I., et al., Expression by transgenesis of a constitutively active mutant form of the prolactin receptor induces premature abnormal development of the mouse mammary gland and lactation failure, Biol Reprod, 70(3): 718-728 (2004).

Kozlowski, M., et al., Differences in growth and transcriptomic profile of bovine mammary epithelial monolayer and three-dimensional cell cultures, J Physiol Pharmacol, 60(Suppl 1): 5-14 (2009).

O'Brien, L.E., et al., Rac1 orientates epithelial apical polarity through effects on basolateral laminin assembly, Nat Cell Biol, 3(9): 831-838 (2001).

PCT/US2020/066209 International Search Report and Written Opinion dated Apr. 22, 2021.

Yang, N.S., et al., Growth of human mammary epithelial cells on collagen gel surfaces, Cancer Res, 41(10): 4093-4100 (1983).

* cited by examiner

Primary Examiner — Taeyoon Kim
(74) *Attorney, Agent, or Firm* — Goodwin Procter LLP

(57) **ABSTRACT**

Disclosed herein are milk compositions that comprise protein, lipid, and oligosaccharide components at concentrations that mimic and/or are substantially similar to human breast milk as produced by a lactating female. The milk compositions include one or more milk components produced in vitro and/or ex vivo from cultured mammary cells.

13 Claims, 7 Drawing Sheets
Specification includes a Sequence Listing.

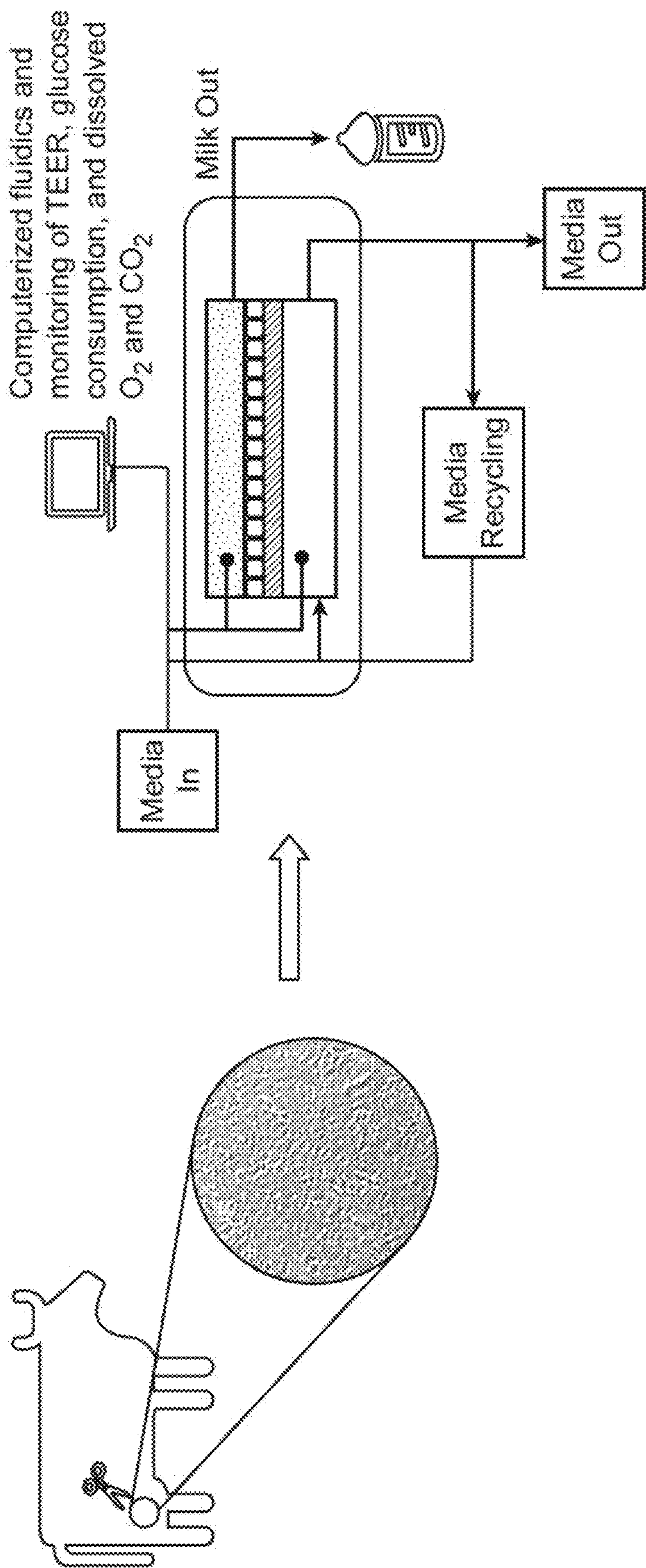


FIG. 1

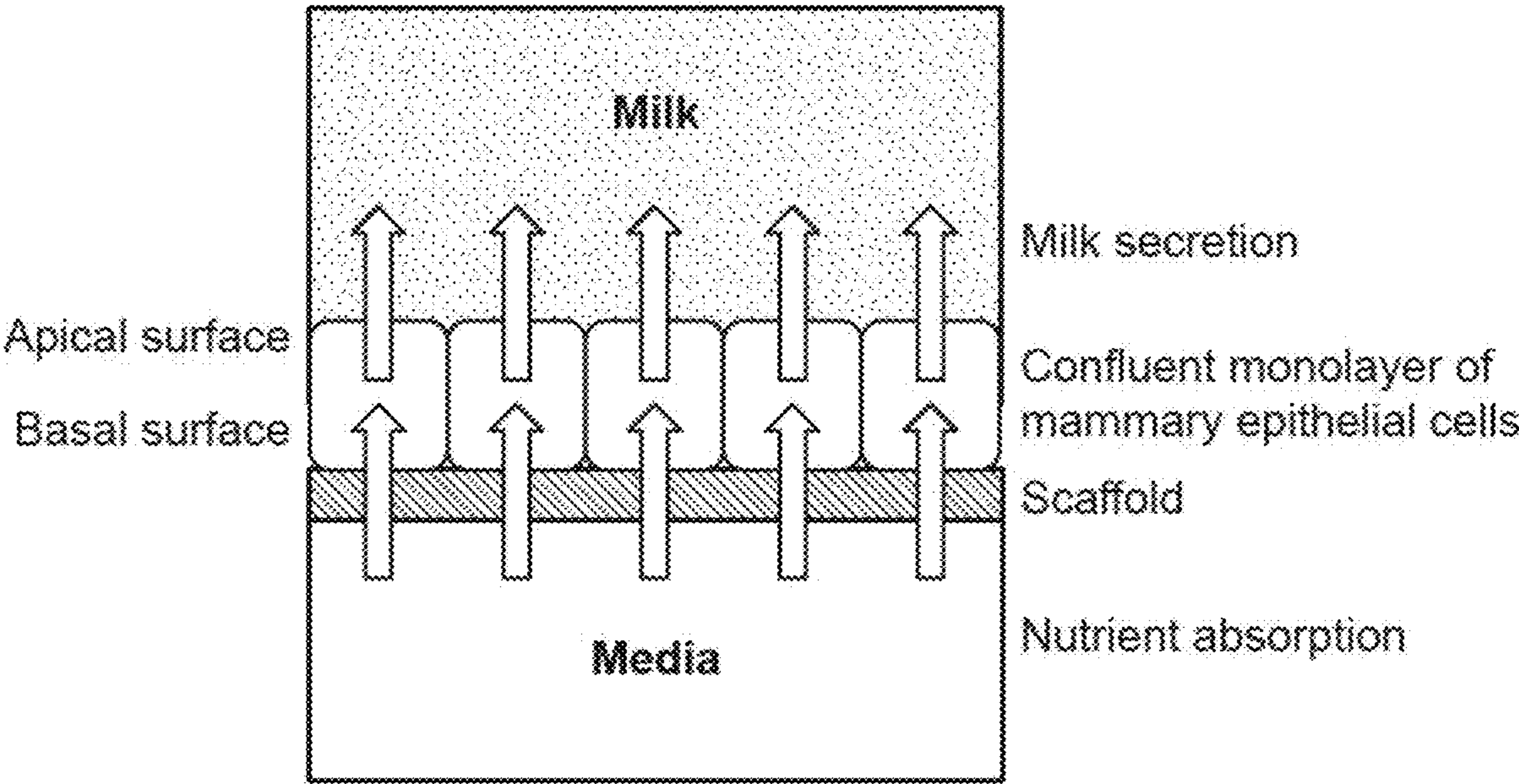


FIG. 2

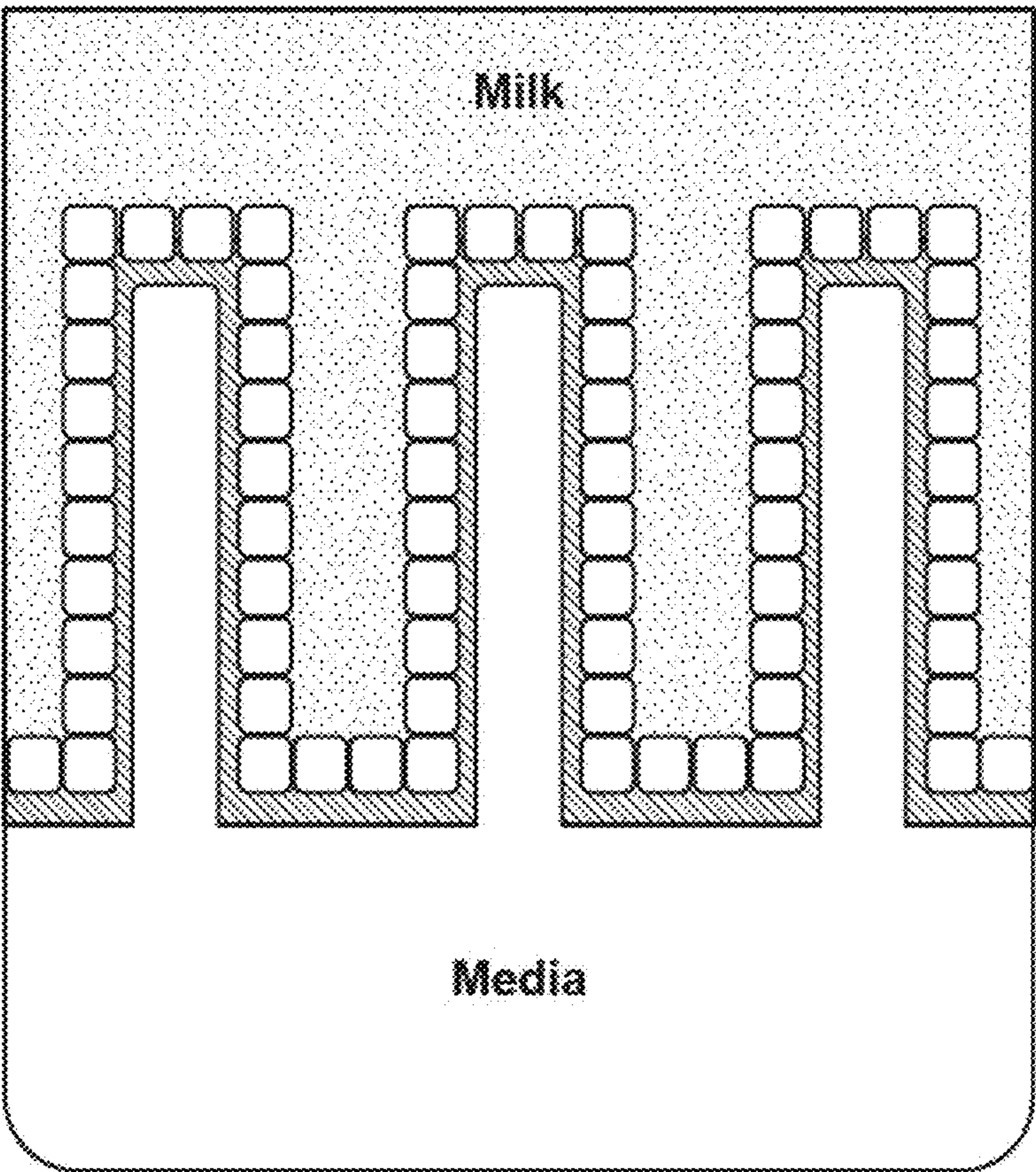


FIG. 3

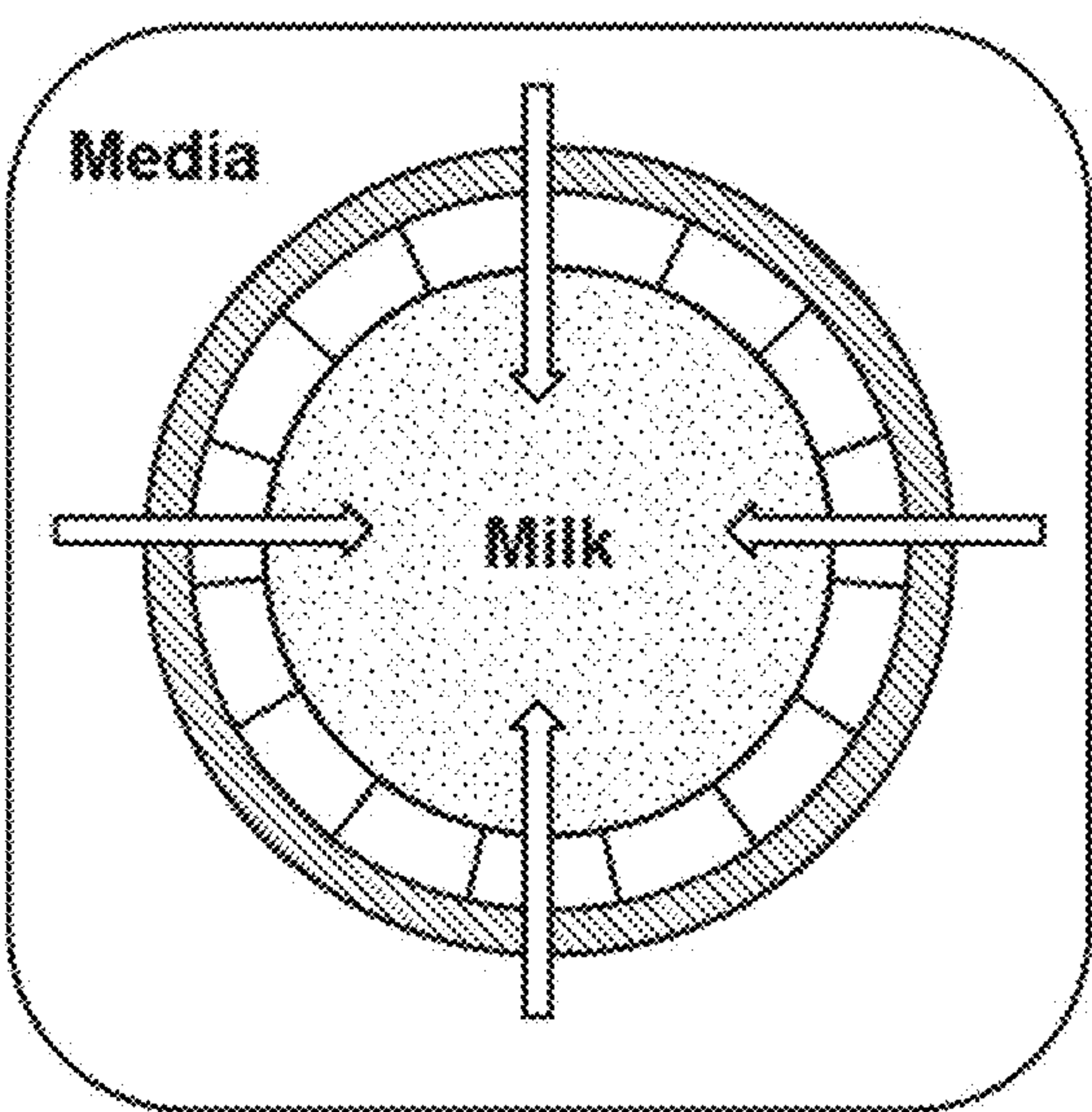
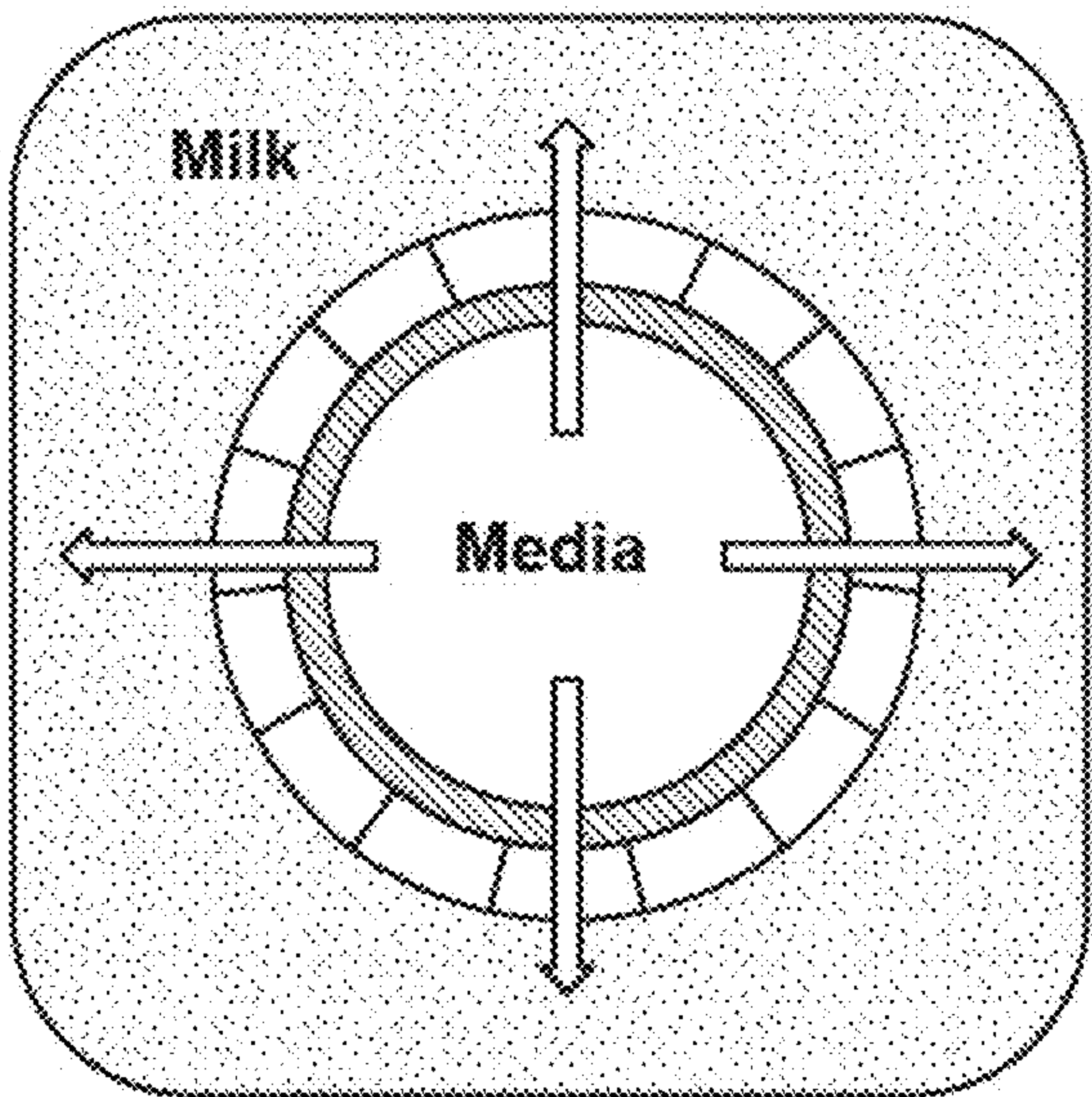
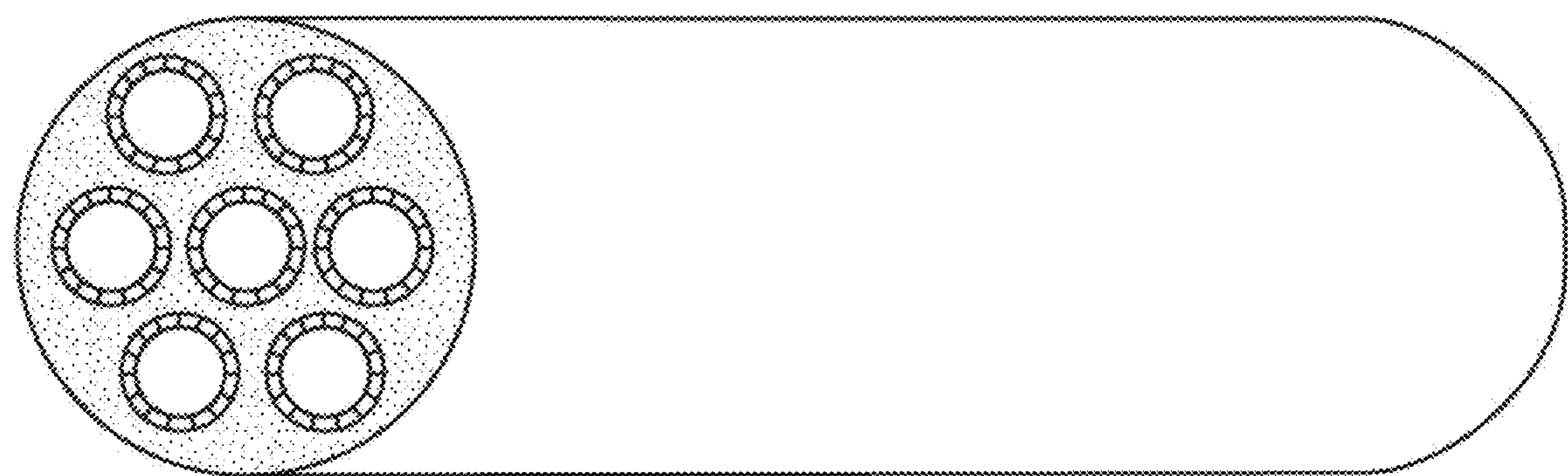


FIG. 4

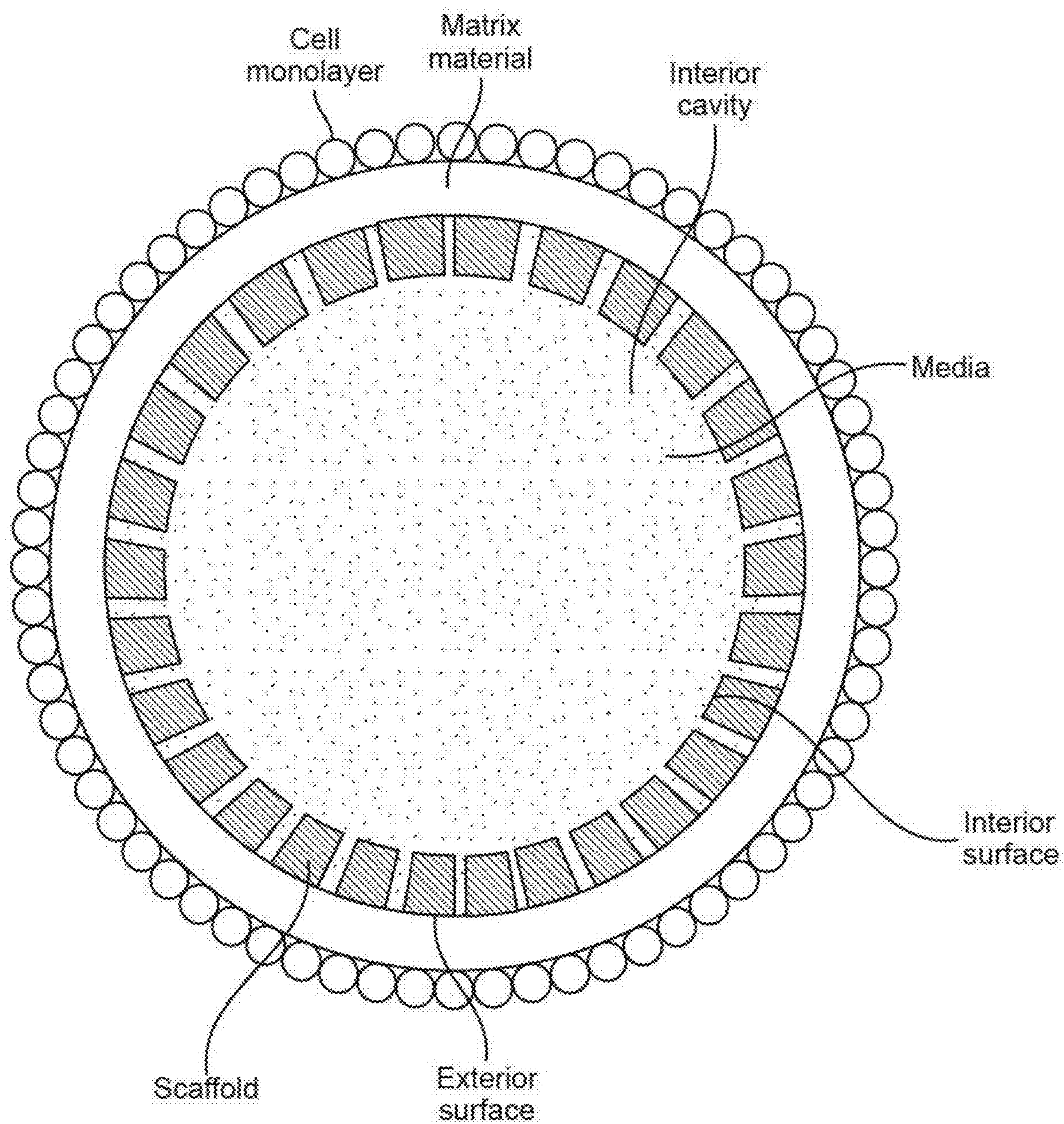


FIG. 5

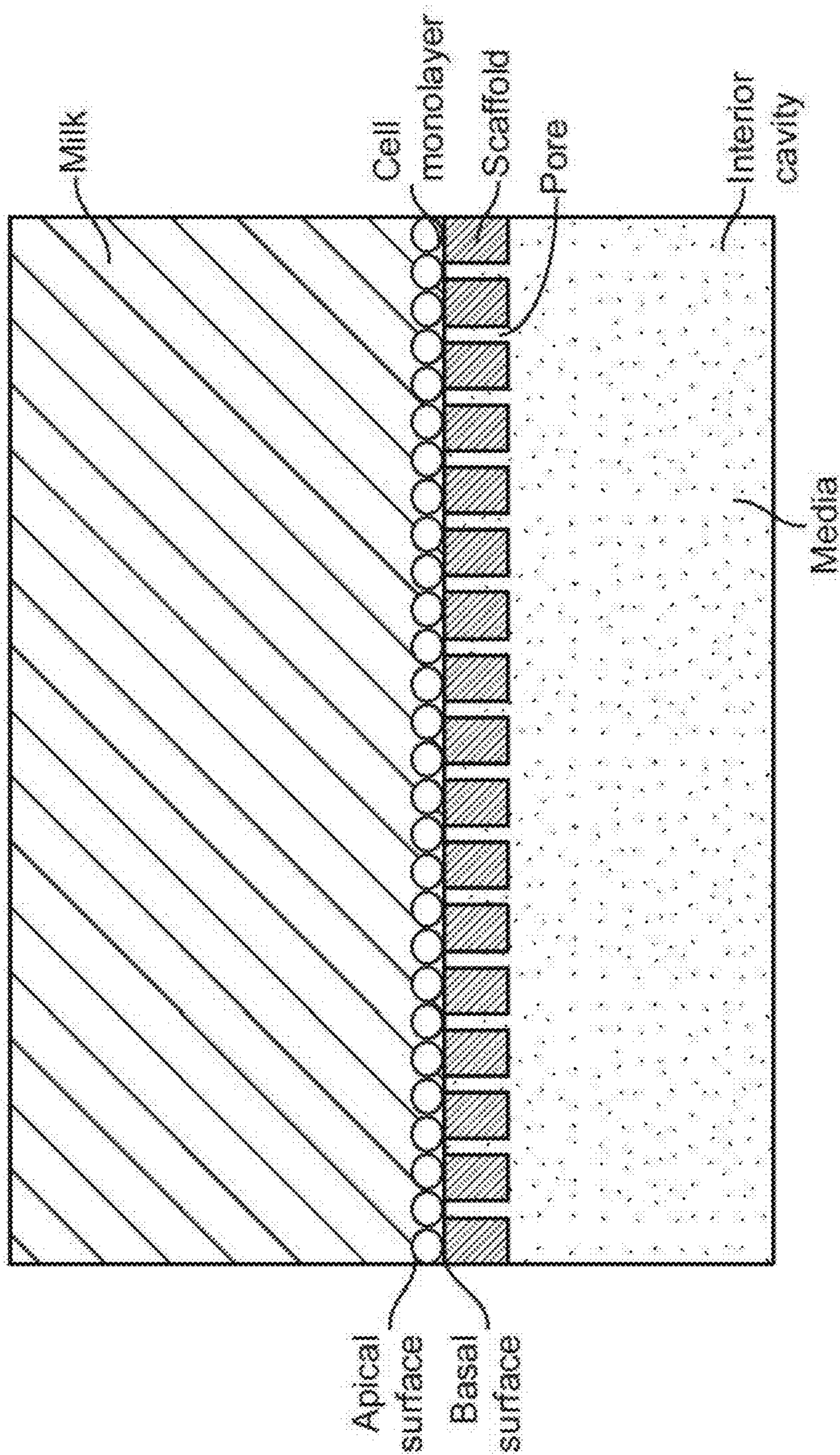


FIG. 6

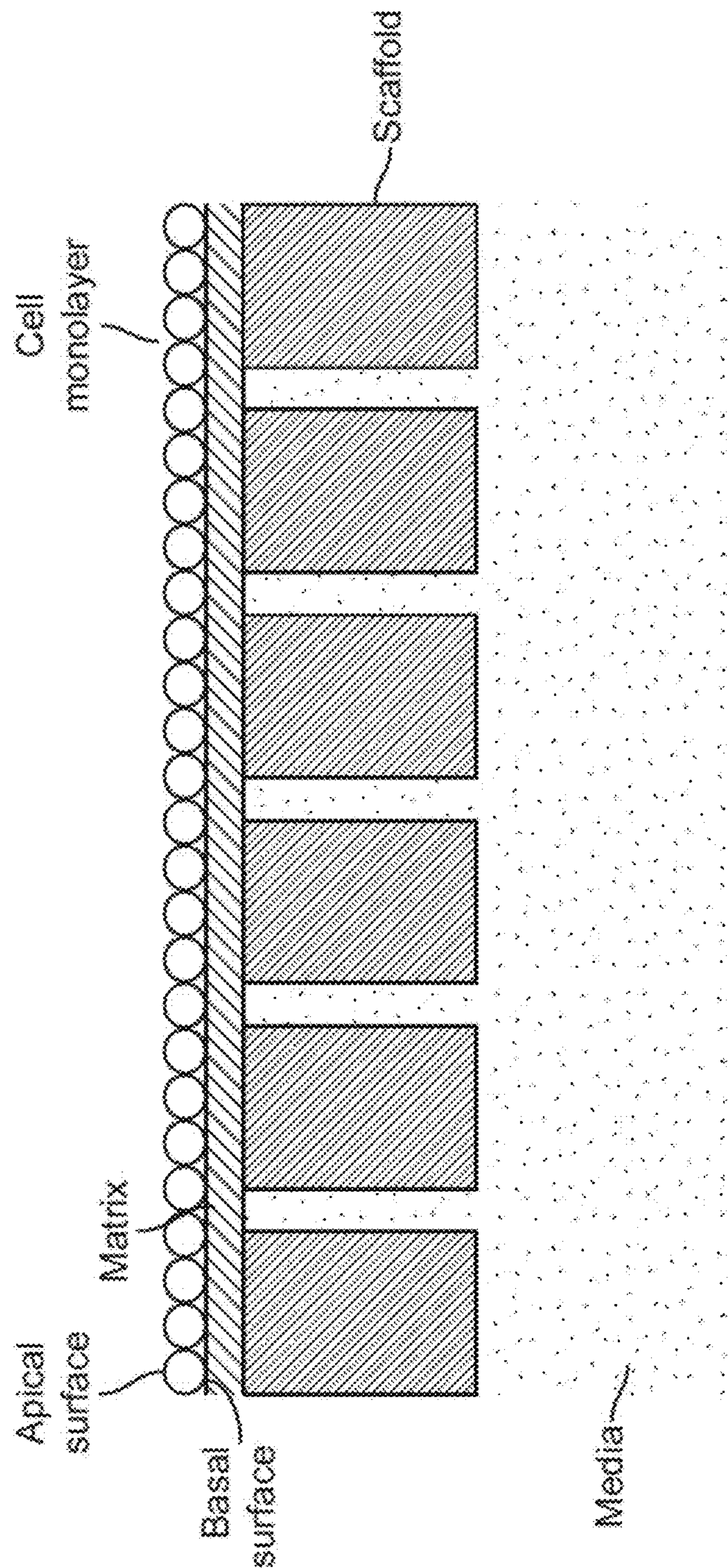


FIG. 7

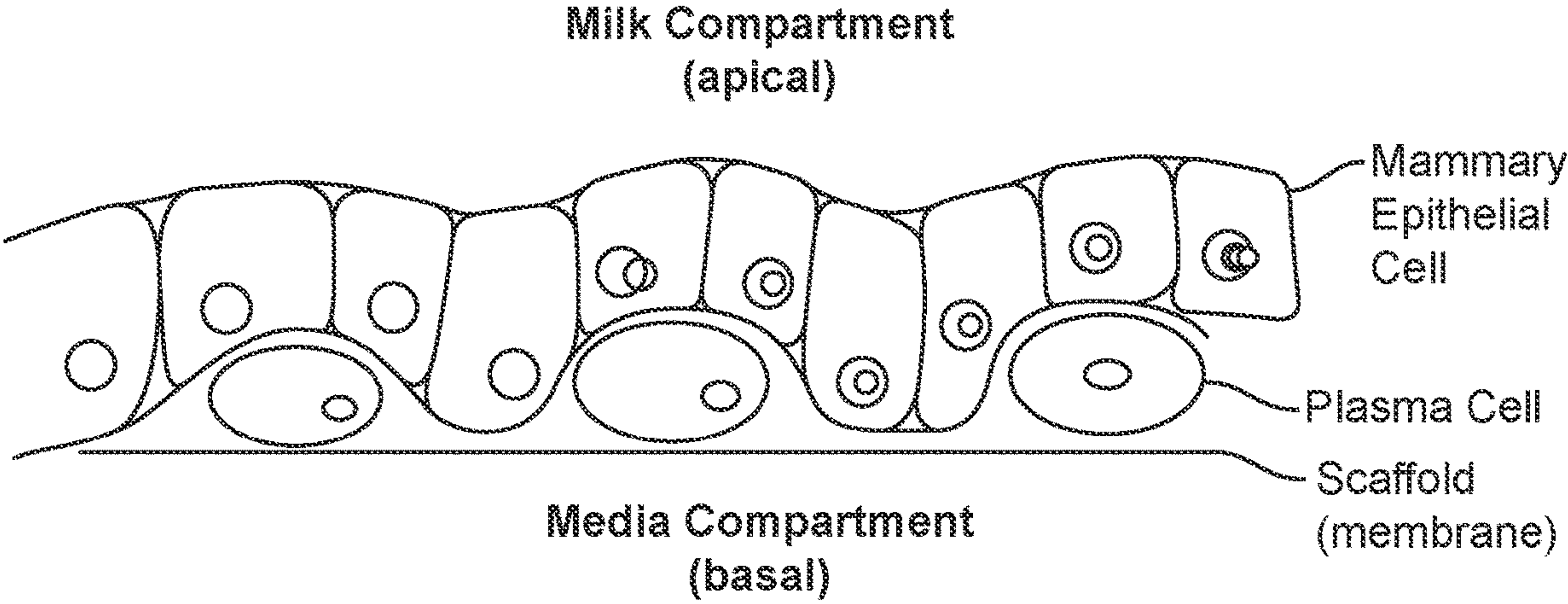


FIG. 8

1

MILK PRODUCT COMPOSITIONS**CROSS-REFERENCE**

This application claims the benefit of priority from U.S. Provisional Application No. 63/030,149 filed on May 26, 2020, and U.S. Provisional Application No. 63/200,480 filed on Mar. 9, 2021, the contents of each are incorporated herein by reference in their entireties.

SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted in ASCII format and is hereby incorporated by reference in its entirety. Said ASCII copy, created on Apr. 26, 2021, is named BMQ_001_SL.txt and is 29,417 bytes in size.

FIELD

This disclosure relates to milk product compositions that comprise protein, lipid, and oligosaccharide components and component concentrations that mimic human breast milk as produced by a lactating female, which milk compositions are produced in vitro and/or ex vivo from cultured mammary cells.

BACKGROUND

Milk is a staple of the human diet, both during infancy and throughout life. The American Academy of Pediatrics and World Health Organization recommend that infants be exclusively breastfed for the first 6 months of life, and consumption of dairy beyond infancy is a mainstay of human nutrition, representing a 700 billion dollar industry worldwide. However, lactation is a physiologically demanding and metabolically intensive process that can present biological and practical challenges for breastfeeding mothers, and milk production is associated with environmental, social, and animal welfare impacts in agricultural contexts.

The possibility of using mammalian cell culture to produce food has gained increasing interest in recent years, with the development of several successful prototypes of meat and sea food products from cultured muscle and fat cells (Stephens et al. 2018 *Trends Food Sci Technol.* 78:155-166). Additionally, efforts are underway to commercialize the production of egg and milk proteins using microbial expression systems. However, this fermentation-based process relies on the genetically engineered expression and purification of individual components and is unable to reproduce the full molecular profile of milk or dairy.

SUMMARY OF THE DISCLOSURE

The present disclosure relates to milk compositions that comprise protein, lipid, and oligosaccharide components and component concentrations that mimic and/or are substantially similar to human breast milk as produced by a lactating female and are produced in vitro and/or ex vivo from cultured mammary cells.

Thus, one aspect of the disclosure relates to milk products comprising specified concentrations or amounts by weight of protein components, lipid components human milk oligosaccharide components and lactose, wherein at least one of the protein components, lipid components, HMOs, and lactose is produced by cultured human mammary epithelial cells. In an embodiment, disclosed herein are methods of

2

producing an isolated cultured milk product from mammary cells, the method comprising: (a) culturing a cell construct in a bioreactor under conditions which produce the cultured milk product, said cell construct comprising: (i) a three-dimensional scaffold having an exterior surface, an interior surface defining an interior cavity, and a plurality of pores extending from the interior surface to the exterior surface; (ii) a matrix material disposed on the exterior surface of the three-dimensional scaffold; (iv) a plurality of plasma cells disposed on the matrix material; and (v) a confluent monolayer of polarized mammary cells disposed on the plurality of plasma cells, wherein the mammary cells are selected from the group consisting of: mammary epithelial cells, mammary myoepithelial cells, mammary progenitor cells, wherein the polarized mammary cells comprise an apical surface and a basal surface; and (b) isolating the cultured milk product. In some embodiments, the cultured milk product comprises secretory IgA (sIgA).

In some embodiments, the bioreactor comprises an apical compartment that is substantially isolated from the internal cavity of the cell construct. In some embodiments, the basal surface of the mammary cells is in fluidic contact with the culture media. In some embodiments, the apical compartment is in fluidic contact with the apical surface of the mammary cells. In some embodiments, the cultured milk product is secreted from the apical surface of the mammary cells into the apical compartment. In some embodiments, the culture media substantially does not contact the cultured milk product. In some embodiments, total cell density of mammary cells within the bioreactor is at least 10^{11} . In some embodiments, total surface area of mammary cells within the bioreactor is at least 1.5 m^2 . In some embodiments, the matrix material comprises one or more extracellular matrix proteins. In some embodiments, total cell density of plasma cells in the bioreactor is about 200 to 500 plasma cells per mm^2 . In some embodiments, the culturing is carried out at a temperature of about 27°C . to about 39°C . In some embodiments, the culturing is carried out at an atmospheric concentration of CO_2 of about 4% to about 6%.

In certain embodiments, described herein are cell constructs, comprising: (a) a three dimensional scaffold having an exterior surface, an interior surface defining an interior cavity/basal chamber, and a plurality of pores extending from the interior surface to the exterior surface; (b) a matrix material disposed on the exterior surface of the three-dimensional scaffold; (c) a culture media disposed within the interior cavity/basal chamber and in fluidic contact with the internal surface; and (d) a plurality of plasma cells disposed on the matrix material; and (e) an at least 70% confluent monolayer of polarized mammary cells disposed on the plurality of plasma cells, wherein the mammary cells are selected from the group consisting of: mammary epithelial cells, mammary myoepithelial cells, and mammary progenitor cells. In some embodiments, the polarized mammary cells comprise an apical surface and a basal surface. In some embodiments, the basal surface of the mammary cells is in fluidic contact with the culture media.

In some embodiments, at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or 100% of the mammary cells are polarized in the same orientation. In some embodiments, the monolayer of polarized mammary cells is at least 70% confluent, at least 80% confluent, at least 90% confluent, at least 95% confluent, at least 99% confluent, or 100% confluent. In some embodiments, the mammary cells comprise a constitutively active prolactin receptor protein. In some embodiments, the culture medium comprises a carbon source, a chemical buffering system, one or more essential

amino acids, one or more vitamins and/or cofactors, and one or more inorganic salts. In some embodiments, the culture medium further comprises prolactin. In some embodiments, the matrix material comprises one or more extracellular matrix proteins. In some embodiments, the three-dimensional scaffold comprises a natural polymer, a biocompatible synthetic polymer, a synthetic peptide, a composite derived from any of the preceding, or any combination thereof. In some embodiments, the natural polymer is collagen, chitosan, cellulose, agarose, alginate, gelatin, elastin, heparan sulfate, chondroitin sulfate, keratan sulfate, and/or hyaluronic acid. In some embodiments, the biocompatible synthetic polymer is polysulfone, polyvinylidene fluoride, polyethylene co-vinyl acetate, polyvinyl alcohol, sodium polyacrylate, an acrylate polymer, and/or polyethylene glycol.

In certain aspects, described herein is a milk product comprising: about 6-14 grams per liter (g/L) protein components; about 18-89 g/L lipid components; about 7-14 g/L human milk oligosaccharides (HMOs); and about 64-77 g/L lactose, wherein at least one of the protein components, lipid components, HMOs, and lactose is produced by cultured human mammary epithelial cells.

In some embodiments, the milk product further comprises one or more immunoglobulins. In some embodiments, the immunoglobulin is secretory IgA.

In some embodiments, the protein component comprises about 55-65% dry weight of the milk product. In some embodiments, the protein component comprises beta-casein, kappa-casein and alpha-casein. In some embodiments, the beta-casein has a concentration of about 0.5-1.5 g/L, the kappa-casein has a concentration of about 0.5-0.6 g/L, and the alpha-casein has a concentration of about 0.1-0.5 g/L in the milk product. In some embodiments, the beta-casein, kappa-casein and alpha-casein together comprise about 35-45% dry weight percent of the protein component of the milk product. In some embodiments, the beta-casein comprises greater than about 50% of total casein content.

In some embodiments, the protein component further comprises one or more of alpha-lactalbumin, lysozyme, lactoferrin, haptocorrin, butyrophilin, osteopontin, mucin MC5, mucin BrE3, and lactadherin. In some embodiments, the protein component further comprises serum albumin. In some embodiments, the alpha-lactalbumin has a concentration of about 2.7-3.3 g/L in the milk product. In some embodiments, the lysozyme has a concentration of about 0.2-0.5 g/L in the milk product. In some embodiments, the lactoferrin has a concentration of about 1.0-2.0 g/L in the milk product. In some embodiments, the haptocorrin has a concentration of about 0.07-0.7 g/L in the milk product. In some embodiments, the butyrophilin has a concentration of about 0.03-0.05 g/L in the milk product. In some embodiments, the osteopontin has a concentration of about 0.05-0.2 g/L in the milk product. In some embodiments, the mucin MC5 has a concentration of about 0.5-0.6 g/L in the milk product. In some embodiments, the mucin BrE3 has a concentration of about 0.5-0.7 g/L in the milk product. In some embodiments, the lactadherin has a concentration of about 0.06-0.07 g/L in the milk product. In some embodiments, the serum albumin has a concentration of about 0.025-3.5 g/L, or about 0.01-2 g/L, or about 0.15-1 g/L, or about 0.2-0.7 g/L. In some embodiments, the protein components are of human origin.

In some embodiments, the lipid component comprises one or more of saturated fatty acids, monounsaturated fatty acids, polyunsaturated fatty acids, cholesterol, phospholipids, plasmalogens and sphingolipids. In some embodiments,

the saturated fatty acids has a concentration of about 5-34 g/L in the milk product. In some embodiments, the saturated fatty acids comprise palmitic acid, stearic acid, and lauric acid. In some embodiments, the palmitic acid comprises at least about 50% sn-2 form. In some embodiments, palmitic acid has a concentration of about 3-18 g/L in the milk product. In some embodiments, stearic acid has a concentration of about 0.7-5 g/L in the milk product. In some embodiments, lauric acid has a concentration of about 0.5-5 g/L in the milk product. In some embodiments, monounsaturated fatty acids have a concentration of about 7-46 g/L in the milk product. In some embodiments, the monounsaturated fatty acid comprises oleic acid. In some embodiments, the oleic acid comprises at least about 50% sn-1 form. In some embodiments, the oleic acid has a concentration of about 7-45 g/L in the milk product. In some embodiments, polyunsaturated fats have a concentration of about 2-20 g/L in the milk product. In some embodiments, the polyunsaturated fats comprise one or more of linoleic acid, alpha-linolenic acid, eicosadienoic acid, arachidonic acid, dihomo-gamma-linolenic acid, and docosahexadienoic acid. In some embodiments, linoleic acid has a concentration of about 2-19 g/L in the milk product. In some embodiments, the linoleic acid comprises at least about 50% sn-3 form. In some embodiments, alpha-linolenic acid has a concentration of about 0.5-0.7 g/L in the milk product. In some embodiments, eicosadienoic acid has a concentration of about 0.5-0.7 g/L in the milk product. In some embodiments, arachidonic acid has a concentration of about 0.5-0.7 g/L in the milk product. In some embodiments, dihomo-gamma-linolenic acid has a concentration of about 0.3-0.5 g/L in the milk product. In some embodiments, docosahexadienoic acid has a concentration of about 0.02-0.4 g/L in the milk product. In some embodiments, linoleic acid has a concentration of about 2-19 g/L, alpha-linolenic acid has a concentration of about 0.5-0.7 g/L, eicosadienoic acid has a concentration of about 0.5-0.7 g/L, arachidonic acid has a concentration of about 0.5-0.7 g/L, dihomo-gamma-linolenic acid has a concentration of about 0.3-0.5 g/L, and docosahexadienoic acid has a concentration of about 0.02-0.4 g/L in the milk product. In some embodiments, the milk product comprises oleic acid, palmitic acid, and linoleic acid, wherein the oleic acid comprises at least about 50% sn-1 form, the palmitic acid comprises at least about 50% sn-2 form, and the linoleic acid comprises at least about 50% sn-3 form. In some embodiments, the milk product comprises cholesterol having a concentration of about 0.09-0.15 g/L in milk product. In some embodiments, phospholipids, plasmalogens and sphingolipids together have a concentration of about 0.1-0.4 g/L in the milk product.

In some embodiments, the human milk oligosaccharide component comprises one or more neutral oligosaccharides. In some embodiments, the human milk oligosaccharide component comprises one or more acidic oligosaccharides. In some embodiments, the human milk oligosaccharide component comprises neutral oligosaccharides and/or acidic oligosaccharides. In some embodiments, the one or more neutral oligosaccharides comprise TF-LNH (trifucosyllacto-N-hexose), 2'-FL (2'-fucosyllactose), DF-LNHII (difucosyllacto-N-hexaose), LNFP I (lacto-N-fucopentaose I), LND-FHI (lacto-N-difucosylhexaose I), LNT (lacto-N-tetraose), LNnT (lacto-N-neotetraose), DF-L (Difucosyllactose), and 3-FL (3-fucosyllactose).

In some embodiments, the one or more acidic oligosaccharides comprise 6'-SL (6'-sialyllactose), DS-LNT (disialyllacto-N-tetraose), FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), LST c (sialyl-lacto-N-tetraose c), and 3'-SL

5

(3'-sialyllactose). In some embodiments, the one or more neutral oligosaccharides comprises TF-LNH (trifucosyllacto-N-hexose), which oligosaccharide has a concentration of about 1-4 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises 2'-FL (2'-fucosyllactose), which oligosaccharide has a concentration of about 1-4 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises DF-LNHII (difucosyllacto-N-hexaose), which oligosaccharide has a concentration of about 1-4 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises LNFP I (lacto-N-fucopentaose I), which oligosaccharide has a concentration of about 0.5-2 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises LNDFHI (lacto-N-difucosyllactose I), which oligosaccharide has a concentration of about 0.2-2 g/L in the milk product.

In some embodiments, the one or more neutral oligosaccharides comprises LNT (lacto-N-tetraose), which oligosaccharide has a concentration of about 0.3-1.5 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises LNnT (lacto-N-neotetraose), which oligosaccharide has a concentration of about 0.5-1.5 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises DF-L (Difucosyllactose), which oligosaccharide has a concentration of about 0.1-1 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises 3-FL (3-fucosyllactose), which oligosaccharide has a concentration of about 0.2-1.5 g/L in the milk product. In some embodiments, the one or more acidic oligosaccharides comprises 6'-SL (6'-sialyllactose), which oligosaccharide has a concentration of about 0.2-1.2 g/L in the milk product. In some embodiments, the one or more acidic oligosaccharides comprises DS-LNT (disialyllacto-N-tetraose), which oligosaccharide has a concentration of about 0.05-1 g/L in the milk product. In some embodiments, the one or more acidic oligosaccharides comprises FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), which oligosaccharide comprises about 0.05-0.7 g/L in the milk product. In some embodiments, the one or more acidic oligosaccharides comprises LST c (sialyl-lacto-N-tetraose c), which oligosaccharide has a concentration of about 0.05-0.7 g/L in the milk product. In some embodiments, the one or more acidic oligosaccharides comprises 3'-SL (3'-sialyllactose), which oligosaccharide has a concentration of about 0.1-0.3 g/L in the milk product.

In some embodiments, the one or more neutral oligosaccharides comprise TF-LNH (trifucosyllacto-N-hexose), 2'-FL (2'-fucosyllactose), DF-LNH II (difucosyllacto-N-hexaose), LNFP I (lacto-N-fucopentaose I), LNDFHI (lacto-N-difucosyllactose I), LNT (lacto-N-tetraose), LNnT (lacto-N-neotetraose), DF-L (Difucosyllactose), and 3-FL (3-fucosyllactose) and wherein the one or more acidic oligosaccharides comprises 6'-SL (6'-sialyllactose), DS-LNT (disialyllacto-N-tetraose), FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), LST c (sialyl-lacto-N-tetraose c), and 3'-SL (3'-sialyllactose). In some embodiments, the milk product comprises about 1-4 g/L TF-LNH (trifucosyllacto-N-hexose), about 1-4 g/L 2'-FL (2'-fucosyllactose), about 1-4 g/L DF-LNH II (difucosyllacto-N-hexaose), about 0.5-2 g/L LNFP I (lacto-N-fucopentaose I), about 0.22 g/L LNDFHI (lacto-N-difucosyllactose I), about 0.3-1.5 g/L LNT (lacto-N-tetraose), about 0.5-1.5 g/L LNnT (lacto-N-neotetraose), about 0.1-1 g/L DF-L (Difucosyllactose), about 0.2-1.5 g/L 3-FL (3-fucosyllactose), about 0.2-1.2 g/L 6'-SL (6'-sialyllactose), about 0.05-1 g/L DS-LNT (disialyllacto-N-tetraose), about 0.05-0.7 g/L FS-LNnH I (fucosyl-

6

sialyl-lacto-N-neohexaose I), about 0.05-0.7 g/L LST c (sialyl-lacto-N-tetraose c), and about 0.1-0.3 g/L 3'-SL (3'-sialyllactose). In some embodiments, the neutral oligosaccharides content comprises at least about 2-fold, or about 3-fold, or about 4-fold, or about 5-fold, or about 6-fold, or about 7-fold, or about 8-fold, or about 9-fold, or about 10-fold, or about 11-fold, or about 12-fold, or about 13-fold, or about 14-fold, or about 15-fold more by weight than acidic oligosaccharide content.

In certain aspects, described herein are milk products comprising: about 6-14 grams per liter (g/L) protein components; about 18-89 g/L lipid components; about 7-14 g/L human milk oligosaccharides (HMOs); and about 64-77 g/L lactose, wherein the protein components comprise beta-casein, kappa-casein, and alpha-casein, alpha-lactalbumin, lysozyme, lactoferrin, haptocorrin, butyrophilin, osteopontin, mucin MC5, mucin BrE3, and lactadherin, wherein the lipid components comprise palmitic acid, stearic acid and lauric acid, oleic acid, linoleic acid, alpha-linolenic acid, eicosadienoic acid, arachidonic acid, dihomo-gamma-linolenic acid, docosahexadienoic acid, cholesterol, phospholipids, plasmalogens and sphingolipids, wherein the human milk oligosaccharides comprise TF-LNH (trifucosyllacto-N-hexose), 2'-FL (2'-fucosyllactose), DF-LNHII (difucosyllacto-N-hexaose), LNFP I (lacto-N-fucopentaose I), LNDFHI (lacto-N-difucosyllactose I), LNT (lacto-N-tetraose), LNnT (lacto-N-neotetraose), DF-L (Difucosyllactose), and 3-FL (3-fucosyllactose), 6'-SL (6'-sialyllactose), DS-LNT (disialyllacto-N-tetraose), FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), LST c (sialyl-lacto-N-tetraose c), and 3'-SL (3'-sialyllactose), and wherein at least one of the protein components, lipid components, HMOs, and lactose is produced by cultured human mammary epithelial cells.

In some embodiments, the milk product comprises about 0.5-1.5 g/L beta-casein, about 0.5-0.6 g/L kappa-casein, about 0.1-0.5 g/L alpha-casein, about 2.7-3.3 g/L alpha-lactalbumin, about 0.2-0.5 g/L lysozyme, about 1.0-2.0 g/L lactoferrin, about 0.07-0.7 g/L haptocorrin, about 0.03-0.05 g/L butyrophilin, about 0.05-0.2 g/L osteopontin, about 0.5-0.6 g/L mucin MC5, about 0.5-0.7 g/L mucin BrE3, about 0.06-0.07 g/L lactadherin, about 2-19 g/L linoleic acid, about 0.5-0.7 g/L alpha-linolenic acid, about 0.5-0.7 g/L eicosadienoic acid, about 0.5-0.7 g/L arachidonic acid, about 0.3-0.5 g/L dihomo-gamma-linolenic acid, about 0.02-0.4 g/L docosahexadienoic acid, about 0.09-0.15 g/L cholesterol, about 0.1-0.4 g/L phospholipids, plasmalogens, and sphingolipids combined, about 1-4 g/L TF-LNH (trifucosyllacto-N-hexose), about 1-4 g/L 2'-FL (2'-fucosyllactose), about 1-4 g/L DF-LNHII (difucosyllacto-N-hexaose), about 0.5-2 g/L LNFP I (lacto-N-fucopentaose I), about 0.2-2 g/L LNDFHI (lacto-N-difucosyllactose I), about 0.3-1.5 g/L LNT (lacto-N-tetraose), about 0.5-1.5 g/L LNnT (lacto-N-neotetraose), about 0.1-1 g/L DF-L (Difucosyllactose), about 0.2-1.5 g/L 3-FL (3-fucosyllactose), about 0.2-1.2 g/L 6'-SL (6'-sialyllactose), about 0.05-1 g/L DS-LNT (disialyllacto-N-tetraose), about 0.05-0.7 g/L FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), about 0.05-0.7 g/L LST c (sialyl-lacto-N-tetraose c), and about 0.1-0.3 g/L 3'-SL (3'-sialyllactose).

In certain aspects, described herein are milk products comprising: about 3-15 percent protein by weight; about 9-92 percent lipid by weight; about 4-15 percent by weight human milk oligosaccharides (HMOs); and about 33-80 percent by weight lactose, wherein at least one of the protein, lipid, HMOs, and lactose is produced by cultured human mammary epithelial cells. In some embodiments, the milk product further comprises serum albumin. In some

embodiments, the milk product comprises about 0.025-3.5 g/L, or about 0.01-2 g/L, or about 0.15-1 g/L, or about 0.3-0.7 g/L serum albumin.

In some embodiments, the milk product further comprises one or more immunoglobulins. In some embodiments, the immunoglobulins comprise one or more of IgA, IgG and IgM. In some embodiments, the IgA comprises one or more of IgA2 (secretory) and IgA1 (non-secretory). In some embodiments, the milk product comprises about 0.2-1.0 g/L secretory IgA. In some embodiments, the milk product comprises about 0.15-1.6 g/L total IgA. In some embodiments, the milk product comprises about 0.03-0.3 g/L IgG. In some embodiments, the milk product comprises about 0.01-0.1 g/L IgM. In some embodiments, the milk product comprises about 0.2-2 percent by weight immunoglobulins.

In some embodiments, the protein components, lipid components, HMOs, and lactose are isolated from cultured mammary epithelial cells. In some embodiments, the cultured mammary epithelial cells comprise one or more immortalized mammary cell lines. In some embodiments, the cultured mammary epithelial cells are derived from one or more primary mammary tissue samples. In some embodiments, the one or more primary mammary tissue samples is derived from a surgical explant of mammary gland tissue. In some embodiments, the one or more primary mammary tissue samples comprises alveolar and/or luminal tissue or cells collected from the alveolar structure of the mammary gland. In some embodiments, the one or more primary mammary tissue samples is derived from a needle aspiration of mammary gland tissue. In some embodiments, the primary mammary tissue further comprises one or more myo-epithelial cells. In some embodiments, the primary mammary tissue further comprises one or more stem cells. In some embodiments, the primary mammary tissue further comprises one or more immune cells. In some embodiments, the immune cells comprise B cells. In some embodiments, the immune cells comprise plasma cells.

In some embodiments, the milk product is isolated from cultured mammary epithelial cells from one or more specific persons. In some embodiments, the milk product isolated from cultured mammary epithelial cells from a specific donor is provided only to that donor or persons authorized by that donor. In some embodiments, the cultured mammary epithelial cells are co-cultured with one or more plasma cells lines. In some embodiments, the cultured mammary epithelial cells are co-cultured with one or more myoepithelial cell lines. In some embodiments, the cultured mammary epithelial cells are co-cultured with one or more stem cell lines.

In some embodiments, the milk product comprises at least about 80%, or at least about 85%, or at least about 90%, or at least about 95%, or at least about 97%, or at least about 98%, or at least about 99%, of the overall macromolecular composition of human breast milk. In some embodiments, non-protein nitrogen content comprises at least about 10%, or at least about 15%, or at least about 20%, or at least about 25%, or at least about 30% of total nitrogen content.

In certain aspects, described herein are frozen milk products, comprising a milk product described herein that has been frozen. In some embodiments, the milk product has been lyophilized. In certain aspects, described herein is a containerized milk product, comprising a milk product described herein that is packaged into a container. In certain aspects, described herein is a containerized frozen milk product, comprising the frozen milk product that is packaged into a container. In certain aspects, described herein is a containerized lyophilized milk product, comprising the lyophilized milk product that is packaged into a container.

In certain aspects, described herein are extracted milk products, comprising one or more components extracted from a milk product described herein. In some embodiments, the one or more components extracted from the collected milk product are lyophilized or concentrated to produce a lyophilized or a concentrated extracted milk product component. In some embodiments, the one or more components extracted from the collected milk product are concentrated by membrane filtration or reverse osmosis. In some embodiments, the one or more extracted components from the collected milk product comprise milk protein, lipid, carbohydrate, vitamin, and minerals.

In certain aspects, described herein are containerized extracted milk products, comprising the extracted milk product described herein that is packaged in a container. In some embodiments, the container is sterile. In some embodiments, the container is vacuum-sealed. In some embodiments, the container is a food grade container. In some embodiments, the container is a canister, ajar, a bottle, a bag, a box, or a pouch.

In some embodiments, the milk product comprises about 500-1150 kcal/L available energy content. In some embodiments, between about 40-55% of the available energy content of the milk product is from lipids. In some embodiments, the milk product comprises between about 95.8 and 195.2 g/L macromolecular content.

In certain aspects, described herein are methods of treating a patient suffering from an infectious disease, comprising: administering to the patient an effective amount of a milk product comprising: about 6-14 grams per liter (g/L) protein components; about 18-89 g/L lipid components; about 7-14 g/L human milk oligosaccharides (HMOs); and about 64-77 g/L lactose, wherein at least one of the protein components, lipid components, HMOs, and lactose. In some embodiments, the milk product further comprises one or more immunoglobulins. In some embodiments, the immunoglobulins comprise one or more of IgA, IgG and IgM. In some embodiments, the IgA comprises one or more of IgA2 (secretory) and IgA1 (non-secretory).

In some embodiments, the milk product comprises about 0.2-1.0 g/L secretory IgA. In some embodiments, milk product comprises about 0.15-1.6 g/L total IgA. In some embodiments, the milk product comprises about 0.03-0.3 g/L IgG. In some embodiments, the milk product comprises about 0.01-0.1 g/L IgM. In some embodiments, the milk product comprises about 0.2-2 percent by weight immunoglobulins. In some embodiments, the infectious disease is a gastrointestinal infection. In some embodiments, the patient is immune compromised.

In certain aspects, described herein are methods of treating an infant suffering from a gastrointestinal infection comprising administering an effective amount of a milk product comprising: about 6-14 grams per liter (g/L) protein components; about 18-89 g/L lipid components; about 7-14 g/L human milk oligosaccharides (HMOs); and about 64-77 g/L lactose, wherein at least one of the protein components, lipid components, HMOs, and lactose. In some embodiments, the milk product further comprises one or more immunoglobulins. In some embodiments, the immunoglobulins comprise one or more of IgA, IgG and IgM. In some embodiments, the IgA comprises one or more of IgA2 (secretory) and IgA1 (non-secretory). In some embodiments, the milk product comprises about 0.2-1.0 g/L secretory IgA. In some embodiments, the milk product comprises about 0.15-1.6 g/L total IgA. In some embodiments, the milk product comprises about 0.03-0.3 g/L IgG. In some embodiments, the milk product comprises about 0.01-0.1 g/L IgM.

In some embodiments, the milk product comprises about 0.2-2 percent by weight immunoglobulins.

These and other aspects of the disclosure are set forth in more detail in the description of the disclosure below.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows an example of the collection of milk for nutritional use from mammary epithelial cells grown as a confluent monolayer in a compartmentalizing culture apparatus in which either fresh or recycled media is provided to the basal compartment and milk is collected from the apical compartment. TEER, transepithelial electrical resistance.

FIG. 2 shows an example of polarized absorption of nutrients and secretion of milk across a confluent monolayer of mammary epithelial cells anchored to a scaffold at the basal surface.

FIG. 3 shows an example micropatterned scaffold that provides increased surface area for the compartmentalized absorption of nutrients and secretion of milk by a confluent monolayer of mammary epithelial cells.

FIG. 4 shows three examples of a hollow fiber bioreactor depicted as a bundle of capillary tubes (top), which can support mammary epithelial cells lining either the external (top and lower left) or internal (lower right) surface of the capillaries, providing directional and compartmentalized absorption of nutrients and secretion of milk.

FIG. 5 exemplifies a cross-section of three-dimensional cell construct. The construct is made up of a scaffold having an interior surface defining an interior cavity/basal chamber and an exterior surface. The interior cavity/basal chamber comprises cell culture media. A matrix material sits on top of the exterior surface of the scaffold. Pores transverse the scaffold from the interior surface to the exterior surface, allowing cell media to contact the basal surface of the cells of the cell monolayer disposed on the matrix material.

FIG. 6 exemplifies a bioreactor for producing a cultured milk product. The bioreactor is made up of a cell construct and an apical chamber. The cell construct is made up of a scaffold having an interior surface defining an interior cavity/basal chamber and an exterior surface. The cavity comprises cell culture media. A matrix material sits on top of the exterior surface of the scaffold. Pores transverse the scaffold from the interior surface to the exterior surface, allowing cell media to contact the basal surface of the cells of the cell monolayer disposed on the matrix material. The apical surface of the cells of the cell monolayer secrete the milk/cultured milk product into the apical chamber. The apical chamber and the interior cavity/basal chamber are separated by the cell monolayer.

FIG. 7 exemplifies a cell construct. The construct is made up of a scaffold having an interior surface defining an interior cavity/basal chamber and an exterior surface. The interior cavity/basal chamber comprises cell.

FIG. 8 exemplifies a cell construct having mammary epithelial cells (MECs) and plasma cells. The plasma cells are adjacent to the scaffold. The MECs form a confluent monolayer above (and in some instances, in between) the plasma cells, with the apical side of the MECs facing the apical compartment (or, milk compartment). The plasma cells secrete IgA, which then binds to a receptor on the basolateral surface of the MECs, triggering internalization of the antibody-receptor complex and further processing of the antibody into sIgA as it transits toward the apical surface (not shown).

DETAILED DESCRIPTION

Milk is a nutrient-rich liquid food produced in the mammary glands of mammals. It is a primary source of nutrition

for infant mammals (including humans who are breastfed) before they are able to digest other types of food. Milk is a complex biological matrix composed of thousands of unique molecules, with nutritional and functional properties that are ideal to support the growth and development of a mammalian infant.

Natural milk contains many macronutrients, including proteins, lipids, polysaccharides and lactose. Each species produces milk with a unique composition that reflects its distinct physiological needs (Beck K L, et al., *J Proteome Res.* 2015; 14(5):2143-2157). As milk composition is optimized for each species, breast milk is considered the gold standard for human infant nutrition and is recommended as the exclusive nutrition source for the first 6 months by the World Health Organization (Mordor Intelligence, Global dairy market (2016-2024)).

Compared with bovine milk, human milk is lower in protein but higher in fat and carbohydrate content (Ballard O, et al., *Pediatr Clin North Am.* 2013; 60(1):49-74). Many components overlap but undergo different processing steps in humans and cows and therefore differ in their final conformations and physiological properties. For example, the positions of specific fatty acids in triglycerides differ between bovine milk and breast milk, and these differences have been shown to affect absorption (Andreas N J, et al., *Early Hum Dev.*; 91 (11):629-635). Additionally, human milk contains hundreds of unique oligosaccharides that contribute to gut maturation and immunity in the developing infant, which are lacking in bovine-based formula (Totten S M, et al., *J Proteome Res.* 2012; 11(12):6124-6133).

Human milk is not merely nutritional. Rather, human milk contains a variety of factors with bioactive qualities that have a profound role in infant survival and health. Early milk from mammals contains antibodies that provide protection to the newborn baby. Additional factors contributing to the disease fighting potential of natural milk include leukocytes, hormones, antimicrobial peptides, cytokines, chemokines, and other bioactive factors.

Secretory IgA (sIgA) is found in naturally-occurring breast milk. IgA is produced by plasma cells located within the breast tissue. The IgA binds to a receptor (polymeric Ig receptor) on the basal surface of mammary epithelial cells. The IgA and receptor are transported into the mammary epithelial cells where the mammary cells process the IgA by cleaving the extracellular domain of the polymeric Ig receptor bound to the IgA, leaving the soluble ectodomain of the Ig receptor (the "secretory component") bound to the IgA. The sIgA is secreted from the apical surface of the mammary epithelial cells. sIgA transferred to children via breast milk is an integral component of an infant's immune system enabling them to fight off infections. Further, sIgA is an important component of the immune systems of all humans and sIgA deficiencies are associated with increased susceptibility to illness. sIgA-based therapies offer the opportunity to prevent and treat infectious diseases at their sites of entry.

Immune cells, including CD20+ B cells, which are precursors for sIgA-secreting plasma cells, are present in normal, non-lactating mammary tissue. During lactation, immune cell homing to the mammary gland contributes to a local, mucosal immunity that helps to prevent infection within the mammary gland. Further, immune cell homing to the mammary gland enables the transfer of sIgA antibodies to an infant via breast feeding.

Unfortunately, efforts at mass production of sIgA are hampered by the fact that it is a multivalent and heavily post-translationally modified protein assembly that requires cooperation with a mucosal epithelium for processing into

its bioactive form. Thus, it is difficult to manufacture sIgA for use with non-naturally occurring breast milk or as a therapeutic for humans.

These are several of the factors that contribute to the inability of infant formula to replicate the properties of breast milk. Therefore, there is a need for improved milk products for developing infants and children that more closely mimic the composition of human breast milk as produced by a lactating female.

Mammary epithelial cells (MECs) in culture have been previously demonstrated to display organization and behavior similar to that observed in vivo (Arevalo et al. 2016 *Am J Physiol Cell Physiol.* 310(5):C348-3 56; Chen et al. 2019 *Curr Protoc Cell Biol.* 82(1):e65). In Arevalo et al., specific biomarkers of MEC populations were detected in immortalized bovine mammary epithelial cells (BME-UV1) and immortalized bovine mammary alveolar cells (MAC-T) cultured on adherent 2-D plates, ultralow attachment surface 3D microplates, and 3D plates coated with Matrigel. Additionally, in Chen et al., protocols are detailed for isolation and culture of human primary mammary epithelial stem/progenitor cells from human breast tissue and subsequent generation of mammospheres using 3D organoid culture on gelatin sponges and Matrigel matrices. However, neither Arevalo nor Chen attempted to stimulate the production of milk from these MEC cultures.

In particular, when grown on an appropriate extracellular matrix and stimulated with prolactin, cultured bovine mammary epithelial cells polarize and organize into structures capable of secreting certain milk components (Blatchford et al. 1999 *Animal Cell Technology: Basic & Applied Aspects* 10:141-145). In Blatchford et al, bovine MECs polarized and formed mammospheres. Casein and butyrophilin were isolated from the cultures. However, the cells did not polarize in one uniform direction. Blatchford, et al. noted that the milk proteins were distributed in between the cells and dispersed throughout the mammospheres. Due to the lack of a uniform polarization orientation, Blatchford had to isolate the secreted proteins from the culture media.

Furthermore, in vitro two-dimensional models, such as those used in Blatchford et al. provide a low surface area-to-volume ratio (low density format). The surface area available for cell attachment limits the number of cells that can be grown

The only known attempt to culture mouse mammary epithelial cells in a high-density format, such as the hollow fiber bioreactor, failed to achieve compartmentalization necessary for the production and extraction of a cultured milk product (Sharfstein et al. 1992 *Biotechnology and Bioengineering* 40:672-680). In Sharfstein et al., growth, long-term expression of functional differentiation, and metabolism of COMMA-1D (an immortalized mouse mammary epithelial cell line) was examined in two different systems: extended batch culture and hollow-fiber reactor culture. Using COMMA-1D seeded onto Costar Transwell® polycarbonate membrane cell culture inserts, Sharfstein et al. created a confluent monolayer capable of barrier formation and polarized metabolism between the apical and basal side that maintained gradients of glucose and lactate. However, using a hollow-fiber bioreactor culture, Sharfstein et al. was unable to achieve separation of basal and apical compartments. Furthermore, it was not determined if nutrient uptake was polarized in a hollow-fiber culture (Sharfstein et al. 1992). Importantly, no prior work has been able to culture mammary epithelial cells from humans or other nutritionally relevant species in a high-density, three-dimensional, compartmentalizing format.

The present disclosure is explained in greater detail below. This description is not intended to be a detailed catalog of all the different ways in which the disclosure may be implemented, or all the features that may be added to the instant disclosure. For example, features illustrated with respect to one embodiment may be incorporated into other embodiments, and features illustrated with respect to a particular embodiment may be deleted from that embodiment. Numerous variations and additions to the various embodiments suggested herein will be apparent to those skilled in the art in light of the instant disclosure, which do not depart from the instant disclosure. Hence, the following specification is intended to illustrate some particular embodiments of the disclosure, and not to exhaustively specify all permutations, combinations, and variations thereof.

Unless the context indicates otherwise, it is specifically intended that the various features of the disclosure described herein can be used in any combination. Moreover, the present disclosure also contemplates that in some embodiments of the disclosure, any feature or combination of features set forth herein can be excluded or omitted. To illustrate, if the specification states that a complex comprises components A, B and C, it is specifically intended that any of A, B or C, or a combination thereof, can be omitted and disclaimed singularly or in any combination.

Definitions

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this disclosure belongs. The terminology used in the description of the disclosure herein is for the purpose of describing particular embodiments only and is not intended to be limiting of the disclosure.

The terminology used herein is for the purpose of describing particular embodiments only and is not intended to be limiting. As used herein, the singular forms “a,” “an,” and “the” are intended to include the plural forms as well, unless the context clearly indicates otherwise. It will be further understood that the terms “comprises” and/or “comprising,” when used in this specification, specify the presence of stated features, elements, and/or components, but do not preclude the presence or addition of one or more other features, elements, components, and/or groups thereof. As used herein, the term “and/or” includes any and all combinations of one or more of the associated listed items. As used herein, phrases such as “between X and Y” and “between about X and Y” should be interpreted to include X and Y. As used herein, phrases such as “between about X and Y” mean “between about X and about Y.” As used herein, phrases such as “from about X to Y” mean “from about X to about Y.”

Moreover, the present disclosure also contemplates that in some embodiments, any feature or combination of features set forth herein can be excluded or omitted.

Furthermore, the term “about,” as used herein when referring to a measurable value such as an amount of a compound or agent of this disclosure, dose, time, temperature, and the like, is meant to encompass variations of $\pm 10\%$, $\pm 5\%$, $\pm 1\%$, $\pm 0.5\%$, or even $\pm 0.1\%$ of the specified amount.

As used herein, the transitional phrase “consisting essentially of” is to be interpreted as encompassing the recited materials or steps and those that do not materially affect the basic and novel characteristic(s) of the disclosure. Thus, the

term “consisting essentially of” as used herein should not be interpreted as equivalent to “comprising.”

As used herein, the compositions described in the present disclosure are referred to interchangeably as (the singular or plural forms of) “nutritional compositions substantially similar to human milk,” “milk products,” “milk compositions,” “cultured milk products,” or equivalent as made clear by the context.

As used herein, by “isolate” (or grammatical equivalents, e.g., “extract”) a product, it is meant that the product is at least partially separated from at least some of the other components in the starting material.

As used herein, the term “polypeptide” encompasses both peptides and proteins, and does not require any particular amino acid length or tertiary structure unless indicated otherwise.

The term “polarized” as used herein in reference to cells and/or monolayers of cells refers to a spatial status of the cell wherein there are two distinct surfaces of the cell, e.g., an apical surface and a basal surface, which may be different. In some embodiments, the distinct surfaces of a polarized cell comprises different surface and/or transmembrane receptors and/or other structures. In some embodiments, individual polarized cells in a continuous monolayer have similarly-oriented apical surfaces and basal surfaces. In some embodiments, individual polarized cells in a continuous monolayer have communicative structures between individual cells (e.g., tight junctions) to allow cross communication between individual cells and to create separation (e.g., compartmentalization) of the apical compartment and basal compartment.

As used herein, “apical surface” means the surface of a cell that faces an external environment or toward a cavity or chamber, for example the cavity of an internal organ. With respect to mammary epithelial cells, the apical surface is the surface from which the cultured milk product is secreted.

As used herein, “basal surface” means the surface of a cell that is in contact with a surface, e.g., the matrix of a bioreactor.

As used herein, “bioreactor” means a device or system that supports a biologically active environment that enables the production of a cultured milk product described herein from mammary cells described herein.

The term “lactogenic” as used herein refers to the ability to stimulate production and/or secretion of milk. A gene or protein (e.g., prolactin) may be lactogenic, as may any other natural and/or synthetic product. In some embodiments, a lactogenic culture medium comprises prolactin, thereby stimulating production of milk by cells in contact with the culture medium.

As used herein, the term “food grade” refers to materials considered non-toxic and safe for consumption (e.g., human and/or other animal consumption), e.g., as regulated by standards set by the U.S. Food and Drug Administration.

Cell Constructs

Included herein are cell constructs for producing a cultured milk product representing the biosynthetic output of cultured mammary epithelial cells (immortalized or from primary tissue samples) and IgA producing cells, for example plasma cells. Disclosed herein, in certain embodiments, are cell constructs for producing cultured milk products comprising sIgA, the cell constructs comprising: (a) a three dimensional scaffold having an exterior surface, an interior surface defining an interior cavity/basal chamber, and a plurality of pores extending from the interior surface to the exterior surface; (b) a matrix material disposed on the exterior surface of the three-dimensional scaffold; (c) a

culture media disposed within the interior cavity/basal chamber and in fluidic contact with the internal surface; (d) a population of plasma cells (PCs) disposed on the matrix material and (e) a continuous monolayer of mammary cells disposed on the population of plasma cells, the mammary cells selected from the group consisting of: (i) mammary epithelial cells, (ii) mammary myoepithelial cells, and (iii) mammary progenitor cells.

Mammary Cells

In some embodiments, the mammary cells comprise milk-producing mammary epithelial cells (MECs), contractile myoepithelial cells, and/or progenitor cells that can give rise to both mammary epithelial cells (MECs) and mammary contractile myoepithelial cells. Mammary epithelial cells (MECs) are the only cells that produce milk. In some embodiments, the mammary cells comprise mammary epithelial cells (MECs), primary mammary epithelial cells, mammary myoepithelial cells and mammary progenitor cells. In some embodiments, the mammary cells are obtained from a tissue biopsy of a mammary gland.

In some embodiments, the mammary cells are derived from breast milk-derived stem cells or breast stem cells originating from tissue biopsy of a mammary gland. The epithelial component of breast milk includes not only mature epithelial cells, but also their precursors and stem cells in culture. A subpopulation of breast milk-derived stem cells displays very high multilineage potential, resembling those typical for human embryonic stem cells (hESCs). Breast stem cells may also originate from tissue biopsy of the mammary gland, and include terminally differentiated MECs. Both breast milk-derived stem cells and breast stem cells originating from tissue biopsy of the mammary gland are multi-potent cells that can give rise to MECs or myoepithelial cells.

In some embodiments, at least 50% of the mammary cells of the cells culture are polarized. In some embodiments, at least 55% of the mammary cells of the cell culture are polarized. In some embodiments, at least 60% of the mammary cells of the cell culture are polarized. In some embodiments, at least 65% of the mammary cells of the cell culture are polarized. In some embodiments, at least 70% of the mammary cells of the cell culture are polarized. In some embodiments, at least 75% of the mammary cells of the cell culture are polarized. In some embodiments, at least 80% of the mammary cells of the cell culture are polarized. In some embodiments, at least 85% of the mammary cells of the cell culture are polarized. In some embodiments, at least 90% of the mammary cells of the cell culture are polarized. In some embodiments, at least 95% of the mammary cells of the cell culture are polarized. In some embodiments, at least 100% of the mammary cells of the cell culture are polarized. In some embodiments, substantially all of the mammary cells of the cell construct are polarized (i.e., have an apical surface and a basal surface). In some embodiments, substantially all of the mammary cells of the cell construct are polarized and substantially all of the polarized cells are oriented in the same direction. For example, in some embodiments, substantially all of the mammary cells have an apical surface and a basal surface, wherein the apical surface of substantially all of the cells is oriented in the same direction and the basal surface of substantially all of the cells is oriented in the same direction.

In some embodiments, the continuous monolayer of mammary cells has at least 70% confluence over the scaffold. In some embodiments, the continuous monolayer of mammary cells has at least about 75% confluence over the scaffold. In some embodiments, the continuous monolayer

15

of mammary cells has at least about 80% confluence over the scaffold. In some embodiments, the continuous monolayer of mammary cells has at least about 85% confluence over the scaffold. In some embodiments, the continuous monolayer of mammary cells has at least about 90% confluence over the scaffold. In some embodiments, the continuous monolayer of mammary cells has at least about 95% confluence over the scaffold. In some embodiments, the continuous monolayer of mammary cells has at least about 99% confluence over the scaffold. In some embodiments, the continuous monolayer of mammary cells has 100% confluence over the scaffold. Genetic Modifications to Mammary Cells

In some embodiments, the mammary cells comprise a constitutively active prolactin receptor protein. In some embodiments, the mammary cells comprise a constitutively active human prolactin receptor protein. Where the primary mammary epithelial cell or immortalized mammary epithelial cells comprise a constitutively active prolactin receptor, the culture medium does not contain prolactin.

In some embodiments, the constitutively active human prolactin receptor protein comprises a deletion of amino acids 9 through 187, wherein the numbering is based on the reference amino acid sequence of a human prolactin receptor identified as SEQ ID NO: 1.

SEQ ID NO: 1: Human prolactin receptor (GenBank accession number AAD32032.1)

MKENVASATVFTLLFLNTCLLNGQLPPGK-
PEIFKCRSPNKETFTCWWRPGTDGGLP TNYSLT-
YHREGETLMHECPDYITGGPN-
SCHFGKQYTSMWRTYIMMVNATNQMGSSFSDEL
YVDVTYIVQPDPPLELAVEVKQPE-
DRKPYLWIKWSPPTLIDLKTGWFTLLYEIRLKPE-
KAAE WEIHFAQQTEFKILSLHPGQKYLQVRCK-
PDHGYWSAWSPATFIQIPSDFTMNDTTVWIS
VAVLSAVICLIIVWAVALKGYSMVTCIFPPVPGP-
KIKGFD AHLLEK GKSEELLSALGCQDFPP
TSDYEDLLVEYLEVDDSEDQHLMSVHSKEHP-
SQGMKPTYLDPDTSGRGSCDSPSLLEK
EEPQANPSTFYDPEVIEKPENPETHTTWDPQCISMEG-
KIPYFHAGGSKCSTWPLPQPSQHNPR
SSYHNITDVCELAVGPAGPATLLNEAGKDALKSSQ-
TIKSREEGKATQQREVESFHSETDQD
TPWLLPQEKTPFGSAKPLDYVEIHKVND-
GALSLLPKQRENSGKPKPGTPENNKEYAKVS
GVMDNNILVLPDPHAKNVACFEESAKE-
APPSLEQNQA EKALANFTATSSKCRLQLGGLDY
LDPACFTHSFH

In some embodiments, the constitutively active human prolactin receptor protein comprising a deletion of the following amino acids: VFTLLFLNTCLLNGQLPPGK-PEIFKCRSPNKETFTCWWRPGTDGGLPTNYSLT-YHREGETLMHECPDYITGGPN-SCHFGKQYTSMWRTYIMMVNATNQMGSSFSDELYVDVTYIVQPDPPLELAVEVKQPE-DRKPYLWIKWSPPTLIDLKTGWFTLLYEIRLKPEKAA (e.g., amino acid positions 10 through 178 of SEQ ID NO: 1).

In some embodiments, the mammary cells comprise a loss of function mutation introduced into a circadian related gene PER2. In some embodiments, the loss of function mutation introduced into a circadian related gene PER2 promotes increased synthesis of cultured milk components. In some embodiments, the loss of function mutation in the PER2 gene comprises an 87-amino acid deletion from position 348 to 434 in PER2, wherein the numbering is based on the reference amino acid sequence of a human PER2 identified as SEQ ID NO: 2.

16

SEQ ID NO: 2: Human Period Circadian Protein Homolog 2 (GenBank Accession Number NM 022817)

MNGYAEFPSPSPNPTKEPVEPQP-
SQVPLQEDVDMSSGSSGHETNENCSTGRDSQGS
CDDSGKJELGMLVEPPDARQSPDTFSLMMAKSEHNP
STSGCSSDQSSKVDTHKEL1KTLKEL KVHLPADK-
KAKGKASTLATLKYALRSVKQVKANEEYYQLL-
MSSEGHPCGADVPSYTVEE MESVTSE-
HIVKNADMFAVAVSLVSGKILYISDQVASIFHCKRD
AFSDAKFVEFLAPHDVGVF
HSFTSPYKLPLWSMCSGADSFTQEC-
MEEKSFFCRVSVRKSHENEIRYHPFRMTPYLVKVRD
QQGAESQLCCLLLAERVHSGYEAPRIPPEKRIFTTTH-
TPNCLFQDQVDERAVPLLGYLPQD LIETPVLVQLHPS-
DRPLMLAIHKKILQSGGQPFDDYSPIRFRARNG-
EYITLDTSWSSFINP
WSRKISFIIGRHKVRVGPLNEDVFAAHPCTEEKALHP-
SIQELTEQIIIRLLLQPVPHSGSSG YGSLGSNG-
SHEHLSQTSSSDSNGHEDSRRRRAEICKNGNKT-
KNRSHYSHESGEQKKKSVT
EMQTNPPAEKKAVPAMEKD-
SLGVSFPEELACKNQPTCSYQQISCLDS-
VIRYLESCNEAATLK RKCEFPANVPALRSSDKR-
KATVSPGPHAGEAEPPSRVNSRTGVGTHLTSLALPG
KAESVASL TSQCSYSS-
TIVHVGDKKPQPELEMVEDAASGPESLD-
CLAGPALACGLSKEKEPFKKLGLTKE VLAHTQKE-
EQSFLQKFKEIRKLSIFQSHCHYYLQERSKGGQPSERTA
PGLRNTSGIDSPWKK TGKNRKLKSKRVKPRDSSES-
TGSGGPVSARPLVGLNATAWSPSDTSQSS-
CPAVPFPAVPVA AYSLPVFPAPGTVAAPPAP-
PHASFTVPAVPVDLQHQFAVQPPFPAPLAPVMAFM
LPSYSP SGTPNLPAFFPSQPQFSPHPTLTSE-
MASASQPEFPEGGTGAMGTTGATETA AVGADCKPGT
SRDQQPKAPLTRDEPSDTQNSDALST-
SSGLNLLL NEDLCSASGSAASESLGSGSLGCDASPS
GAGSSDTSHTSKYFGSIDSENNHKAKMNTG-
MEESEHFIKCVLQDPIWLLMADADSSVMM
TYQLPSRNLEAVLKEDREKLKLLQKLQPRFT-
ESQKQELREVHQWMQTGGLPAAIDVAECV YCEN-
KEKGNICIPYEEDIPSLGLSEVSDTKEDENG-
SPLNHRIEEQT

In some embodiments, the loss of function mutation introduced into PER2 comprises a deletion of the following amino acids: CLFQDQVDERAVPLLGYLPQDLIETPVLVQLHPSDRPLMLAIHKKILQSGGQPFDDYSPIRFRAR NGEYITLDTSWSSFINPWSRKISFIIGRHKV (e.g., amino acid positions 341 through 434 of SEQ ID NO: 2).

In some embodiments, the mammary cells comprise a polynucleotide encoding a prolactin receptor comprising a modified intracellular signaling domain. In some embodiments, the loss of function mutation introduced into a circadian related gene PER2 promotes increased synthesis of individual cultured milk components. In some embodiments, the prolactin receptor comprises a truncation wherein position 154 of exon 10 has been spliced to the 3' sequence of exon 11. In some embodiments, the prolactin receptor comprises a sequence according to SEQ ID NO: 3.

SEQ ID NO: 3: Human Isoform 4 of Prolactin Receptor (GenBank Accession Number AF416619; Trott et al. 2003 *J. Mol. Endocrinol* 30(1):31-47)

MKENVASATVFTLLFLNTCLLNGQLPPGK-
PEIFKCRSPNKETFTCWWRPGTDGGLP TNYSLT-
YHREGETLMHECPDYITGGPN-
SCHFGKQYTSMWRTYIMMVNATNQMGSSFSDEL
YVDVTYIVQPDPPLELAVEVKQPE-

17

DRKPYLWIKWSPPTLIDLTGWFTLLYEIRLKPE-
KAAE WEIIFAGQQTEFKILSLHPGQKYLQVRCK-
PDHGYWSAWSPATFIQIPSDFTMNDTTVWIS
VAVLSAVICLIIVWAVALKGYSMTVCIFPPVPGP-
KIKGFDAHLLKKGKSEELLSALGCQDFPP
TSDYEDLLVEYLEVDDSEDQHLMSVHSKEHP-
SQGDPLMLGASHYKNLKSYPKISSQGRL
AVFTKATLTTVQ

In some embodiments, the mammary cells comprise a polynucleotide encoding a modified (e.g., recombinant) effector of a prolactin protein. In some embodiments, the modified effector of the prolactin protein comprises a janus kinase-2 (JAK2) tyrosine kinase domain. In some embodiments, the modified effector comprises a JAK2 tyrosine kinase domain fused to a signal transducer and activator of transcription-5 (STAT5) tyrosine kinase domain (e.g., a polynucleotide encoding a JAK2 tyrosine kinase domain linked to the 3' end of a polynucleotide encoding the STAT5 tyrosine kinase domain). In some embodiments, the modified effector of a prolactin protein promotes increased synthesis of individual cultured milk components. In some embodiments, the modified effector has a sequence according to SEQ ID NO: 4. Bolded amino acids correspond to the JAK2 kinase domain of amino acid positions 757 through 1129 of a reference human JAK2 amino acid sequence.

SEQ ID NO: 4. STA5A Human Signal Transducer and Activator of Transcription 5A Fused at 3' End to Amino Acids 757-1129 of JAK2 Human Tyrosine-Protein Kinase

MAGWIAQQL	QGDALRQMQR	LYGQHFPIEV
RHYLAQWIES	QPWDAIDLDN	PQDRAQATQL
LEGLVQELQK	KAEHQVGEDG	FLKKIKLGHY
ATQLQKTYDR	CPELVRCIR	HILYNEQRLV
REANNCSSPA	GILVDAMSQK	HLQINQTFEE
LRLVTQDTEN	ELKKLQQQTQE	YFIIQYQESL
RIQAQFAQLA	QLSPQERLSR	ETALQQKQVS
LEAWLQREAA	TLQQYRVELA	EKHQKTLQLL
RKQQTIILDD	ELIQWKRRQQ	LAGNGGPPEG
SLDVLQSWCE	KLAEIHWQNR	QQIRRAEHL
QQLPIGPVE	EMLAEVNATI	TDIISALVTS
TFIIEKQPPQ	VLKTQTKFAA	TVRLLVGGKL
NVHMNPPQVK	ATIISEQQA	SLLKNENTRN
ECSGEILNNC	CVMEYHQATG	TLSAHFRNMS
LKRIKRRDRR	GAESVTEEF	TVLFESQFSV
GSNELVFQVK	TLSPWVIV	HGSQDHNATA
NAFAE	PGRVPFAVPD	KVLWPQLCEA
SNRGLTKENL	VFLAQKLFNN	SSSHLEDYSG
LSVSWSQFNR	ENLPGWNYTF	WQWFDGVM
LKKHHKPHWN	DGAILGFVNK	QQAHDLLINK
FLLRFS	DSEIGGITIA	WKFDSPERNL
FSIRSLADRL	GDLSYLIYVF	PDRPKDEVFS
LAKA	VDGYVKPQIK	QWPEFVNAS
MDQAPSPAVC	PQAPYNMYPQ	NPDHVLDQDG
DETMDV	ARHVEELLRR	PMDSLDSRLS
SAR	GSLSLDSQ	RKLQFYEDRH
ANLINNCMDY	EPDFRPSFRA	IIRDLNSLFT
TEND	MLPNMRIGAL	GFSGAFEDRD
FLQQLGKGNF	GSVEMCRYDP	LQDNTGEWA
VKKLQHSTEE	HLRDFEREIE	ILKSLQHDNI
VKYKGVCSYA	GRNKLKIME	YLPYGSLRDY
KERIDH	IKLLQYTSQI	CKGMEYLGTK
NILVENENRV	KIGDFGLTKV	LPQDKEYYKV
KEPGESPIFW	YAPESLTESK	FSVASDVWSF
ELFTY	IEKSKSPPAE	FMRMIGNDKQ
LLKNNGRLLPR	PDGCPDEIYM	IMTECWNNNV
NQRPSFRDLA	LRVDQIRDN	

In some embodiments, the mammary cells are immortalized. In some embodiments, the mammary cells comprise

18

one or more nucleic acids encoding human telomerase reverse transcriptase (hTERT) or simian virus 40 (SV40). In some embodiments, the mammary cells comprise a small hairpin RNA (shRNA) to p16 (Inhibitor of Cyclin-Dependent Kinase 4) (p16(INK4)) and Master Regulator of Cell Cycle Entry and Proliferative Metabolism (c-MYC).

In some embodiments, the method comprises introducing into the mammary cell: (a) a polynucleotide encoding a prolactin receptor comprising a modified intracellular signaling domain, optionally wherein the prolactin receptor comprises a truncation wherein position 154 of exon 10 has been spliced to the 3' sequence of exon 11; (b) a polynucleotide encoding a chimeric prolactin receptor that binds to a ligand, which is capable of activating milk synthesis in the absence of prolactin; (c) a polynucleotide encoding a constitutively or conditionally active prolactin receptor protein, optionally wherein the polynucleotide encodes a constitutively active human prolactin receptor protein comprising a deletion of amino acids 9 through 187 (e.g., a deletion of amino acids 9 through 187, wherein the numbering is based on the reference amino acid sequence of a human prolactin receptor identified as SEQ ID NO: 1); (d) a polynucleotide encoding a modified (e.g., recombinant) effector of a prolactin protein comprising (i) a janus kinase-2 (JAK2) tyrosine kinase domain, optionally wherein the JAK2 tyrosine kinase domain is fused to a signal transducer and activator of transcription-5 (STAT5) tyrosine kinase domain (e.g., a polynucleotide encoding a JAK2 tyrosine kinase domain linked to the 3' end of a polynucleotide encoding the STAT5 tyrosine kinase domain); and/or (ii) a prolactin receptor intracellular domain fused to a JAK2 tyrosine kinase domain; (e) a loss of function mutation into a circadian related gene PER2 (period circadian protein homolog 2); and/or (f) a polynucleotide encoding one or more glucose transporter genes GLUT1 and/or GLUT12, thereby increasing the rate of nutrient uptake at the basal surface of the monolayer.

Plasma Cells

Plasma cells are derived from a human donor. In some embodiments, the plasma cells are derived from bone marrow, spleen, and/or a lymph node. a primary mammary tissue sample. In certain embodiments, the plasma cells are derived from mucosal epithelial cells other than mammary cells (e.g., from oronasal, gastrointestinal, or respiratory tissue). In some embodiments, the plasma cells are derived from a plasma cell line. In certain embodiments, the plasma cells are derived from a plasmacyte cell line. In some embodiments, the plasma cells are isolated and sorted from non-plasma cells via fluorescence-activated cell sorting, magnetic-activated cell sorting, and/or microfluidic cell sorting. In some embodiments, plasma cells, plasmablasts, or pre-plasmablasts are sorted and isolated by FACS analysis using markers known in the art (e.g., CD38, CD138 and/or CD19). In certain embodiments, the plasma cells are cultivated with the immortalized mammary epithelial cells on a scaffold, thereby producing a cell construct for producing a cultured milk product with secretory products of the plasma cells and mammary cells (e.g., sIgA). In certain embodiments, the plasma cells are grown on a scaffold below a monolayer of mammary cells. In certain embodiments, the plasma cells are grown as dispersed populations of plasma cells overlaid by a monolayer of mammary cells. In certain embodiments, the plasma cells are stimulated to produce immunoglobins during co-culture with mammary cells. In certain embodiments, the plasma cells produce one or more immunoglobins of a class selected from IgG, IgM and IgA. In certain embodiments the plasma cells produce IgA. In

certain embodiments, plasma cells produce IgA, and the IgA is processed by mammary epithelial cells to yield sIgA that is bound to secretory component, and the sIgA is secreted by the apical surface of the mammary cells.

Scaffolds

In some embodiments, the cell construct further comprises a scaffold having a top surface/exterior surface and a bottom surface/interior surface. In some embodiments, the scaffold is a 2-dimensional surface or a 3-dimensional surface (e.g., a 3-dimensional micropatterned surface, and/or as a cylindrical structure that is assembled into bundles). A non-limiting example of a 2-dimensional surface scaffold is a Transwell® filter. In some embodiments, the scaffold is a 3-dimensional surface. Non-limiting examples of a 3-dimensional micropatterned surface include a microstructured bioreactor, a decellularized tissue (e.g., a decellularized mammary gland or decellularized plant tissue), micropatterned scaffolds fabricated through casting or three-dimensional printing with biological or biocompatible materials, textured surface. In some embodiments, the scaffold is produced by electrospinning cellulose nanofibers and/or a cylindrical structure that can be assembled into bundles (e.g., a hollow fiber bioreactor). In some embodiments, the scaffold is porous. In some embodiments, the scaffold is a 3D scaffold. In some embodiments, the 3-dimensional scaffold is any structure which has an enclosed hollow interior/central cavity. In some embodiments, the three dimensional scaffold joins with one or more surfaces to form an enclosed interior chamber/basal compartment. For example, the scaffold can join with one or more walls of a bioreactor to form the interior chamber/basal compartment. In some embodiments, the scaffold is a hollow fiber bioreactor. In some embodiments, the 3D scaffold is a tube in which the central cavity is defined by the interior surface of the scaffold. In some embodiments, the 3D scaffold is a hollow sphere in which the central cavity is defined by the interior surface of the scaffold.

For in vitro culture methods for studies of intestinal absorption, 2-dimensional surface scaffold such as Transwells® have long been used as the standard as they provide both apical and basolateral spaces to simulate the gut-blood-barrier and enable both active and passive transport of drugs and nutrients. However, cells seeded onto flat supports exhibit markedly different phenotypes to cells in vivo, partly due to the poor representation of the 3-D extracellular microenvironments.

A 3-dimensional scaffold allows the cells (e.g., MECs and plasma cells) to grow or interact with their surroundings in all three dimensions. Unlike 2D environments, a 3D cell culture allows cells in vitro to grow in all directions, approximating the in vivo mammary environment. Further, the 3D scaffold allows for a larger surface area for culture of the cells and for metabolite and gas exchange, plus it enables necessary compartmentalization—enabling the cultured milk product to be secreted into one compartment, while the cell culture media is contacted with the mammary cells and plasma cells in another compartment. To date, a confluent monolayer with polarized separation of basal and apical cell surfaces using mammary epithelial cell on a 3D surface has not been achieved (Sharfstein et al. 1992).

In some embodiments, the scaffold is porous. In some embodiments, the scaffold is permeable to the cell media, allowing the cell media to contact the cells of the cell monolayer. In some embodiments, the scaffold is transversely by at least one pore that allows the cell media to contact the basal surface of the cells of the cell monolayer.

In some embodiments, the top surface/exterior surface of the scaffold is coated with a matrix material. In some embodiments, the matrix is made up of one or more extracellular matrix proteins. Non-limiting examples of extracellular matrix proteins include collagen, laminin, entactin, tenascin, and/or fibronectin. In some embodiments, the scaffold comprises a natural polymer, a biocompatible synthetic polymer, a synthetic peptide, and/or a composite derived from any combination thereof. In some embodiments, a natural polymer useful with this invention includes, but is not limited to, collagen, chitosan, cellulose, agarose, alginate, gelatin, elastin, heparan sulfate, chondroitin sulfate, keratan sulfate, and/or hyaluronic acid. In some embodiments, a biocompatible synthetic polymer useful with this invention includes, but is not limited to, cellulose, polysulfone, polyvinylidene fluoride, polyethylene co-vinyl acetate, polyvinyl alcohol, sodium polyacrylate, an acrylate polymer, and/or polyethylene glycol. In some embodiments, the top of the scaffold is coated with laminin and collagen.

In some embodiments, the matrix material is porous. In some embodiments, the matrix material is permeable to the cell media, allowing the cell media to contact the cells of the cell monolayer. In some embodiments, the matrix material is transversely by at least one pore that allows the cell media to contact the basal surface of the cells of the cell monolayer.

In some embodiments, the pore size of the scaffold and/or matrix material is at least about 0.1 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 0.2 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 0.3 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 0.4 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 0.5 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 0.6 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 0.7 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 0.8 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 0.9 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 1.0 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 1.1 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 1.2 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 1.3 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 1.4 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 1.5 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 1.6 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 1.7 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 1.8 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 1.9 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 2.0 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 2.1 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 2.2 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 2.2 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 2.3 μm . In some embodiments, the pore size of the scaffold

21

and/or matrix material is at least about 2.4 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 2.5 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 2.6 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 2.7 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 2.8 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 2.9 μm . In some embodiments, the pore size of the scaffold and/or matrix material is at least about 3.0 μm .

In some embodiments, the cell construct comprises: (a) a three dimensional scaffold having an exterior surface, an interior surface defining an interior cavity/basal chamber, and a plurality of pores extending from the interior surface to the exterior surface; (b) a matrix material disposed on the exterior surface of the three-dimensional scaffold; (c) a culture media disposed within the interior cavity/basal chamber and in fluidic contact with the internal surface; (d) a population of plasma cells (PCs) disposed on the matrix material and (e) a continuous monolayer of mammary cells disposed on the population of plasma cells, the mammary cells selected from the group consisting of: (i) mammary epithelial cells, (ii) mammary myoepithelial cells, and (iii) mammary progenitor cells; wherein the continuous monolayer mammary epithelial cells has an apical surface and a basal surface (e.g., the cells form a polarized and confluent cell monolayer).

Bioreactor

Disclosed herein, in certain embodiments, are bioreactors, comprising: (a) an apical compartment comprising a cultured milk product; and (b) at least one cell construct comprising: (a) a three dimensional scaffold having an exterior surface, an interior surface defining an interior cavity/basal chamber, and a plurality of pores extending from the interior surface to the exterior surface; (b) a matrix material disposed on the exterior surface of the three-dimensional scaffold; (c) a culture media disposed within the interior cavity/basal chamber and in fluidic contact with the internal surface; (d) a population of plasma cells (PCs) disposed on the matrix material and (e) a continuous monolayer of mammary cells disposed on the population of plasma cells, the mammary cells selected from the group consisting of: (i) mammary epithelial cells, (ii) mammary myoepithelial cells, and (iii) mammary progenitor cells. In certain embodiments, the cell construct of the bioreactor comprises at least a 70% confluent monolayer of polarized mammary cells disposed on the matrix material, wherein the mammary cells are selected from the group consisting of: mammary epithelial cells, mammary myoepithelial cells, and mammary progenitor cells; wherein the apical surface of the mammary cells is in fluidic contact with the apical compartment.

In some embodiments, the bioreactor is an enclosed bioreactor. In some embodiments, the apical chamber is substantially isolated from the interior cavity/basal compartment.

A hollow fiber bioreactor is an exemplary bioreactor for use with the methods disclosed here. The hollow fiber bioreactor is a high-density, continuous perfusion culture system that closely approximates the environment in which cells grow in vivo. It consists of thousands of semi-permeable 3D scaffolds (i.e., hollow fibers) in a parallel array within a cartridge shell fitted with inlet and outlet ports. These fiber bundles are potted or sealed at each end so that any liquid entering the ends of the cartridge will necessarily

22

flow through the interior of the fibers. Cells are generally seeded outside the fibers within the cartridge in the extra capillary space (ECS).

Three fundamental characteristics differentiate hollow fiber cell culture from other methods: (1) cells are bound to a porous matrix much as they are in vivo, not a plastic dish, microcarrier or other impermeable support, (2) the molecular weight cut off of the support matrix can be controlled, and (3) extremely high surface area to volume ratio (150 cm^2 or more per mL) which provides a large area for metabolite and gas exchange for efficient growth of host cells.

The bioreactor structure provides a fiber matrix that allows permeation of nutrients, gases and other basic media components, as well as cell waste products, but not cells, where the cells can be amplified. Hollow fiber bioreactor technology has been used to obtain high density cell amplification by utilizing hollow fibers to create a semi-permeable barrier between the cell growth chamber and the medium flow. Since the surface area provided by this design is large, using this fiber as a culture substrate allows the production of large numbers of cells. Cells growing in the 3-dimensional environment within the bioreactor are bathed in fresh medium as it perfuses through the hollow fibers.

To replicate the topography of the intestine, Costello et al. developed a 3-D printed bioreactor that can both contain porous villus scaffolds via micromolding (Costello et al. 2017 *Scientific Reports* 7(12515): 1-10). This geometrically complex molded scaffold provided separation of the apical and basolateral spaces in a manner in which fluid flow exposes intestinal epithelial cells to physiologically relevant shear stresses (Costello et al. 2017). Similarly, a long-term culture in vitro culture in a simulated gut-like environment was created by Morada et al. using a hollow fiber bioreactor which allowed for two controlled separate environments (biphasic) to provide host cells with oxygen and nutrients from the basal layer, while allowing a low oxygen nutrient rich environment to be developed on the apical surface (Morada et al. 2016 *International Journal for Parasitology* 26: 21-29).

In configuring the hollow fiber bioreactor, there are design considerations and parameters that can be varied depending upon the goals associated with expansion of the cells. One such design consideration is the size of the pores in the fiber wall. This is generally designed to allow the passage of nutrients to the cells, carry away waste, provide desired products to the cells (such as growth factors), to remove desired products from the cells, and exclude certain factors that may be present from reaching the cells. Accordingly, the pore size of the fiber walls can be varied to modify which components will pass through the walls. For example, pore size can allow the passage of large proteinaceous molecules, including growth factors, including, but not limited to, epidermal growth factor and platelet-derived growth factor. The person of ordinary skill in the art would understand how to vary the pore size depending upon the components that it is desirable to pass through the fiber walls to reach the cells or to carry material from the cells.

In some embodiments, the pore size is about 0.2 μm . In some embodiments, the pore size is about 0.1 μm . In some embodiments, the pore size is about 0.2 μm . In some embodiments, the pore size is about 0.3 μm . In some embodiments, the pore size is about 0.4 μm . In some embodiments, the pore size is about 0.5 μm . In some embodiments, the pore size is about 0.6 μm . In some embodiments, the pore size is about 0.7 μm . In some embodiments, the pore size is about 0.8 μm . In some embodiments, the pore size is about 0.9 μm . In some

23

embodiments, the pore size is about 1.0 μm . In some
embodiments, the pore size is about 1.1 μm . In some
embodiments, the pore size is about 1.2 μm . In some
embodiments, the pore size is about 1.3 μm . In some
embodiments, the pore size is about 1.4 μm . In some
embodiments, the pore size is about 1.5 μm . In some
embodiments, the pore size is about 1.6 μm . In some
embodiments, the pore size is about 1.7 μm . In some
embodiments, the pore size is about 1.8 μm . In some
embodiments, the pore size is about 1.9 μm . In some
embodiments, the pore size is about 2.0 μm . In some
embodiments, the pore size is about 2.1 μm . In some
embodiments, the pore size is about 2.2 μm . In some
embodiments, the pore size is about 2.2 μm . In some
embodiments, the pore size is about 2.3 μm . In some
embodiments, the pore size is about 2.4 μm . In some
embodiments, the pore size is about 2.5 μm . In some
embodiments, the pore size is about 2.6 μm . In some
embodiments, the pore size is about 2.7 μm . In some
embodiments, the pore size is about 2.8 μm . In some
embodiments, the pore size is about 2.9 μm . In some
embodiments, the pore size is about 3.0 μm .

Methods of Making Cell Constructs

Disclosed herein, in certain embodiments, are methods of
making a cell construct for producing a cultured milk
product comprising immunoglobulins. In some embodi-
ments, the method comprises (a) depositing (i) isolated
mammary epithelial cells, mammary myoepithelial cells
and/or mammary progenitor cells, and (ii) isolated plasma
cells on the upper surface of a scaffold having an upper
surface and lower surface to produce a mixed population of
plasma cells and mammary cells (i.e., mammary epithelial
cells, mammary myoepithelial cells and/or mammary pro-
genitor cells); (b) cultivating the mixed population of mam-
mary cells and plasma cells of (a) on the scaffold, to produce
a monolayer of polarized mammary cells located adjacent to
and above the plasma cells, wherein the plasma cells are
located adjacent to and above the upper surface of the
scaffold, wherein the upper surface is located adjacent to and
above the lower surface of the scaffold, and wherein the
polarized mammary cells comprise an apical surface and a
basal surface, thereby producing a cell construct for pro-
ducing the cultured milk product. In some embodiments, the
mammary cells are primary mammary cells. In some
embodiments, the mammary cells are derived from a cell
culture. In some embodiments, the mammary epithelial
cells, myoepithelial cells and/or mammary progenitor cells
are isolated from bone marrow, spleen tissue, lymph node
tissue, mammary explants from mammary tissue (e.g.,
breast, udder, teat tissue), or raw breastmilk. In some
embodiments, the mammary cells comprise mammary epi-
thelial cells. In some embodiments, the mammary cells,
comprise mammary myoepithelial cells. In some embodi-
ments, the mammary cells, comprise mammary progenitor
cells. In some embodiments, the plasma cells are isolated
from any suitable human tissue or a cell culture. In some
embodiments, the mammary cells and plasma cells are
deposited concurrently. In some embodiments, the plasma
cells are deposited onto the surface of the scaffold prior to
the deposition of the mammary cells.

In some embodiments, the method comprises (a) depos-
iting (i) isolated immortalized mammary epithelial cells,
mammary myoepithelial cells and/or mammary progenitor
cells, and (ii) isolated plasma cells on the upper surface of
a scaffold having an upper surface and lower surface to
produce a mixed population of plasma cells and immortal-
ized mammary cells (i.e., immortalized mammary epithelial

24

cells, immortalized mammary myoepithelial cells and/or
immortalized mammary progenitor cells); (b) cultivating the
mixed population of immortalized mammary cells and
plasma cells of (a) on the scaffold, to produce a monolayer
of polarized immortalized mammary cells located adjacent
to and above the plasma cells, wherein the plasma cells are
located adjacent to and above the upper surface of the
scaffold, wherein the upper surface is located adjacent to and
above the lower surface of the scaffold, and wherein the
polarized immortalized mammary cells comprise an apical
surface and a basal surface, thereby producing a cell con-
struct for producing the cultured milk product. In some
embodiments, the immortalized mammary cells comprise
immortalized mammary epithelial cells. In some embodi-
ments, the immortalized mammary cells, comprise immor-
talized mammary myoepithelial cells. In some embodi-
ments, the immortalized mammary cells, comprise
immortalized mammary progenitor cells. In some embodi-
ments, the plasma cells are isolated from any suitable human
tissue or a cell culture. In some embodiments, the immor-
talized mammary cells and plasma cells are deposited con-
currently. In some embodiments, the plasma cells are depos-
ited onto the surface of the scaffold prior to the deposition
of the immortalized mammary cells. In certain embodi-
ments, plasma cells are added to the culture of immortalized
mammary epithelial cells to produce a co-culture of mam-
mary cells and plasma cells. In certain embodiments, the
plasma cells are cultivated with the immortalized mammary
epithelial cells on the scaffold, thereby producing a cell
construct for producing a cultured milk product with secre-
tory products of the immune cells and mammary cells (e.g.,
sIgA). In certain embodiments, the isolated mammary cells
are immortalized prior to co-culture of the cells.

In certain embodiments, the immune cells are stimulated
to produce immunoglobins during co-culture. In certain
embodiments, the immune cells produce one or more immu-
noglobins of a class selected from IgG, IgM and IgA. In
certain embodiments the immune cells produce secretory
IgA. Classes of immunoglobins produced by the immune
cells include one or more IgA, IgM, and IgG. In certain
embodiments, immune cells are co-cultured with MECs in a
bioreactor according to methods described herein. In certain
embodiments, the bioreactor is a hollow fiber bioreactor
described herein.

In certain embodiments, mammary cells are modified
and/or stimulated with prolactin according to the methods
described herein to stimulate and optimize milk production.
In certain embodiments, the mammary cells are modified to
express a constitutively active prolactin receptor protein.

In certain embodiments, mammary cells are identified and
isolated from mammary tissue samples. In some embodi-
ments, the mammary cells are isolated and sorted via fluo-
rescence-activated cell sorting, magnetic-activated cell sort-
ing, and/or microfluidic cell sorting. In certain
embodiments, the mammary epithelial cell populations are
sorted by FACS analysis using markers known in the art for
identifying the cell populations. In certain embodiments,
myoepithelial mammary cells and luminal epithelial mam-
mary cells are isolated by FACS analysis. In certain embodi-
ments, progenitor myoepithelial mammary cells and/or pro-
genitor luminal epithelial mammary cells are isolated by
FACS analysis. Any suitable method known in the art for
sorting mammary epithelial cells (e.g., luminal epithelial
cells), myoepithelial cells, progenitor cells, and immune
cells can be used. For example, mammary cells can be sorted
using CD24, EPCAM and/or CD49f, cell surface markers.

In some embodiments, plasma cells are identified and isolated from primary mucosal tissue (e.g., oronasal, gastrointestinal, respiratory or mammary). In some embodiments, plasma cells are identified and isolated from primary mammary tissue samples. In some embodiments, the plasma cells are isolated and sorted via fluorescence-activated cell sorting, magnetic-activated cell sorting, and/or microfluidic cell sorting. In certain embodiments, plasma cells are sorted and isolated by FACS analysis. In certain embodiments plasma cells, plasmablasts, or pre-plasmablasts are sorted and isolated by FACS analysis using markers known in the art (e.g., CD20, CD38, CD138 and/or CD19).

In some embodiments, the culturing and/or cultivating of the mammary cells and plasma cells for the cell construct is carried out at a temperature of about 35° C. to about 39° C. (e.g., a temperature of about 35° C., 35.5° C., 36° C., 36.5° C., 37° C., 37.5° C., 38° C., 38.5° C. or about 39° C., or any value or range therein, e.g., about 35° C. to about 38° C., about 36° C. to about 39° C., about 36.5° C. to about 39° C., about 36.5° C. to about 37.5° C., or about 36.5° C. to about 38° C.). In some embodiments, the culturing and/or cultivating is carried out at a temperature of about 37° C.

In some embodiments, the culturing and/or cultivating of the mammary cells and plasma cells for the cell construct is carried out at an atmospheric concentration of CO₂ of about 4% to about 6%, e.g., an atmospheric concentration of CO₂ of about 4%, 4.25%, 4.5%, 4.75%, 5%, 5.25%, 5.5%, 5.75%, or 6% or any value or range therein, e.g., about 4% to about 5.5%, about 4.5% to about 6%, about 4.5% to about 5.5%, or about 5% to about 6%). In some embodiments, the culturing and/or cultivating is carried out at an atmospheric concentration of CO₂ of about 5%.

In some embodiments, the culturing and/or cultivating of the mammary cells and the plasma cells for the cell construct comprises culturing and/or cultivating in a culture medium that is exchanged about every day to about every 10 days (e.g., every 1 day, every 2 days, every 3 days, every 4 days, every 5 days, every 6 days, every 7 days, every 8 days, every 9 days, every 10 days, or any value or range therein, e.g., about every day to every 3 days, about every 3 days to every 10 days, about every 2 days to every 5 days). In some embodiments, the culturing and/or cultivating further comprises culturing in a culture medium that is exchanged about every day to about every few hours to about every 10 days, e.g., about every 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, or 24 hours to about every 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 days or any value or range therein. For example, in some embodiments, the culturing and/or cultivating further comprises culturing and/or cultivating in a culture medium that is exchanged about every 12 hours to about every 10 days, about every 10 hours to about every 5 days, or about every 5 hours to about every 3 days.

In some embodiments, the cell construct is stored in a freezer or in liquid nitrogen. The storage temperature depends on the desired storage length. For example, freezer temperature (e.g., storage at a temperature of about 0° C. to about -80° C. or less, e.g., about 0° C., -10° C., -20° C., -30° C., -40° C., -50° C., -60° C., -70° C., -80° C., -90° C., -100° C. or any value or range therein) may be used if the cells are to be used within 6 months (e.g., within 1, 2, 3, 4, 5, or 6 months). For example, liquid nitrogen may be used (e.g., storage at a temperature of -100° C. or less (e.g., about -100° C., -110° C., -120° C., -130, -140, -150, -160, -170, -180, -190° C., -200° C., or less) for longer term storage (e.g., storage of 6 months or longer, e.g., 6, 7, 8, 9, 10, 11, or 12 months, or 1, 2, 3, 4, 5, 6 or more years).

Cultured Milk Products

The present disclosure relates to cultured milk products that comprise protein, lipid, and oligosaccharide components and component concentrations that mimic human breast milk as produced by a lactating female, which compositions are produced, at least in part, by in vitro and/or ex vivo cultured mammary cells.

In some embodiments, two compositionally defined products can be obtained depending on the cellular inputs and culture conditions: (a) a functional nutrition product representing the biosynthetic output of cultured mammary epithelial cells (immortalized or from primary tissue samples); and (b) a similar product further comprising immunoglobulins, produced when plasma cells (PCs) are co-cultured with mammary epithelial cells. Plasma cells can be derived from a primary mammary tissue sample or, e.g., from a plasma cell line.

Contemplated product compositions of the present disclosure can be defined by total levels of protein, lipid, and carbohydrate (Tables 1A-1C) and/or by a signature of specific macronutrient components (Tables 2A-2C) present in concentrations and proportions consistent with human milk.

TABLE 1A

Macromolecular Composition of Functional Nutrition Products Collected from Human Mammary Epithelial Cells Cultured With or Without Plasma Cells.			
Macromolecular Fraction	Concentration, g/L	% wt	Source
Protein	3-21 ^a	1-44	MECs
Total Immunoglobulin ^b	0.4-1.8	0.14-3.75	PCs
Lipid	9-135	3.1-95+	MECs
HMO	3.5-21	1.2-44	MECs
Lactose	32-115	11-95+	MECs
Total macromolecular	48-293		MECs ±
content			PCs
Energy (kcal/L)	375-1725		

HMO, human milk oligosaccharide. MEC, mammary epithelial cell; PC, plasma cell.
^aNon-immunoglobulin protein content.
^bFormulations lacking immunoglobulin content can be derived from MEC culture in the absence of plasma cells. Formulations with immunoglobulin content can be obtained by coculture of MECs with plasmacytes.
^cLong-chain fatty acids linoleic acid and alpha-linoleic acid are not synthesized by mammalian cells and are supplemented in cell culture media.

TABLE 1B

Macromolecular Composition of Functional Nutrition Products Collected from Human Mammary Epithelial Cells Cultured With or Without Plasma Cells.			
Macromolecular Fraction	Concentration, g/L	% wt	Source
Protein	4.5-17.5 ^a	1.8-24	MECs
Total Immunoglobulin ^b	0.6-1.5	0.25-2.1	PCs
Lipid	13.5-111	5.5-95+	MECs
HMO	5.25-17.5	2.2-24	MECs
Lactose	48-96	20-95+	MECs
Total macromolecular	71.9-244		MECs ±
content			PCs
Energy (kcal/L)	375-1438		

HMO, human milk oligosaccharide. MEC, mammary epithelial cell; PC, plasma cell.
^aNon-immunoglobulin protein content.
^bFormulations lacking immunoglobulin content can be derived from MEC culture in the absence of plasma cells. Formulations with immunoglobulin content can be obtained by co-culture of MECs with plasmacytes.
^cLong-chain fatty acids linoleic acid and alpha-linoleic acid are not synthesized by mammalian cells and are supplemented in cell culture media.

TABLE 1C

Macromolecular Composition of Functional Nutrition Products Collected from Human Mammary Epithelial Cells Cultured With or Without Plasma Cells.				
Macromolecular Fraction	Concentration, g/L	% wt	% vol	Source
Protein	6-14 ^a	3-15	1	MECs
Total Immunoglobulin ^b	0.8-1.2	0.4-1.3	0.1	PCs
Lipid	18-89	9-92	4	MECs
HMO	7-14	4-15	2.4	MECs
Lactose	64-77	33-80	4.6	MECs
Total macromolecular	95.8-195.2		10-15	MECs ±
content				PCs
Energy (kcal/L)	500-1150			

HMO, human milk oligosaccharide. MEC, mammary epithelial cell; PC, plasma cell.
^aNon-immunoglobulin protein content.
^bFormulations lacking immunoglobulin content can be derived from MEC culture in the absence of plasma cells. Formulations with immunoglobulin content can be obtained by co-culture of MECs with plasmacytes.
^cLong-chain fatty acids linoleic acid and alpha-linoleic acid are not synthesized by mammalian cells and are supplemented in cell culture media.

In some embodiments, the concentrations of components indicated in Tables 1A-1C can vary, each individually, for example, by having a concentration that is greater than that indicated by 0.1 fold, or 0.2 fold, or 0.3 fold, or 0.4 fold, or 0.5 fold, or 0.6 fold, or 0.7 fold, or 0.8 fold, or 0.9 fold, or 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 fold.

In some embodiments, the concentrations of components indicated in Tables 1A-1C can vary, each individually, for example, by having a concentration that is less than that indicated by 0.1 fold, or 0.2 fold, or 0.3 fold, or 0.4 fold, or 0.5 fold, or 0.6 fold, or 0.7 fold, or 0.8 fold, or 0.9 fold.

In some embodiments, milk products are contemplated herein that include a subset of the components (i.e., macromolecular fractions) of Tables 1A-1C. In other embodiments, milk products contemplated herein can exclude one or more of the components (i.e., macromolecular fractions) of Tables 1A-1C.

TABLE 2A

Macromolecular Content and Concentration Ranges of Functional Nutrition Products Derived from Human Mammary Epithelial Cells Cultured With or Without Plasma Cells.				
Macromolecular Fraction	Concentration, g/L			
Protein				
P-Casein	0.25-1.9			
K-Casein	0.25-0.9			
a-Casein	0.05-0.75			
a-Lactalbumin	1.35-4.9			
Lysozyme	0.2-0.75			
Lactoferrin	0.5-3			
Haptocorrin	0.03-1.1			
Butyrophilin	0.02-0.075			
Osteopontin	0.025-0.3			
Mucin MC5	0.25-0.9			
Mucin BrE3	0.25-1.1			
Lactadherin	0.03-1.1			
Immunoglobulins	0.1-3			
Secretory IgA	0.1-1.5			
	0.07-2.4			
	0.02-0.45			
Total IgA Total IgG Total IgM	0.005-0.2			
Lipids	2.5-51			
Saturated fatty acids	2-27			
Palmitic acid (C16:0)	0.35-7.5			
Stearic acid (C18:0)	0.25-7.5			
Lauric acid (C12:0)	3.5-69			
Monounsaturated fatty acids	3.5-68			
Oleic acid (C18:1 n-9 Z)	1-30			

TABLE 2A-continued

Macromolecular Content and Concentration Ranges of Functional Nutrition Products Derived from Human Mammary Epithelial Cells Cultured With or Without Plasma Cells.	
Macromolecular Fraction	Concentration, g/L
Polyunsaturated fatty acids	
Linoleic acid, LA	1-29
(C18:2 n-6 Z)	0.25-1.1
a-Linolenic acid, ALA (C18:3 n-3)	0.25-1.1
Eicosadienoic acid (C20:2)	0.25-1.1
Arachidonic acid, AA (C20:4 n-6)	0.15-8
Dihomo-y-linolenic acid, DGLA (C20:3 n-6)	0.01-0.6
Docosahexadienoic acid,	0.045-0.22
DHA (C22:6 n-3) Cholesterol	
Phospholipids, plasmalogens, sphingolipids	0.05-0.6
Human Milk Oligosaccharides	0.8-4.8
Neutral	0.8-4.8
TF-LNH (trifucosyllacto-N-hexose)	0.8-4.8
2'-FL (2'-fucosyllactose)	
DF-LNHII (difucosyllacto-N-hexaose)	0.4-2.4
LNFP I (1 acto-A-fucopentaose I)	0.16-2.4
LNDFHI (lacto-N-difucosylhexaose I)	0.23-1.8
LNT (lacto-A-tetraose)	0.4-1.8
LNnT (lacto-A-neotetraose)	0.08-1.2
DF-L (Difucosyllactose) 3-FL	0.16-1.8
(3-fucosyllactose) Acidic	
6'-SL (6'-sialyllactose)	0.16-1.44
DS-LNT (disialyllacto-A-tetraose)	0.04-1.2
FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I) LST c	0.04-0.84
(sialyl-lacto-A-tetraose c)	0.04-0.84
3'-SL (3'-sialyllactose)	0.08-0.36
Lactose	34-104

TABLE 2B

Macromolecular Content and Concentration Ranges of Functional Nutrition Products Derived from Human Mammary Epithelial Cells Cultured With or Without Plasma Cells.	
Macromolecular Fraction	Concentration, g/L
	0.38-1.9
	0.38-0.75
Protein P-Casein K-Casein a-Casein	0.08-0.63
a-Lactalbumin	2.0-4.13
Lysozyme	0.15-0.63
Lactoferrin	0.8-2.5
Haptocorrin	0.05-0.89
Butyrophilin	0.02-0.063
Osteopontin	0.04-0.25
Mucin MC5	0.38-0.75
Mucin BrE3	0.38-0.89
Lactadherin	0.045-0.09
Immunoglobulins	0.15-2.5
Secretory IgA	0.15-1.25
Total IgA	0.11-2
Total IgG	0.02-0.38
Total IgM	0.0075-0.13
Lipids	
Saturated fatty acids	3.75-43
Palmitic acid (C16:0)	2.25-23
Stearic acid (C18:0)	0.53-6.3
Lauric acid (C12:0)	0.38-6.3
Monounsaturated fatty acids	5.25-58
Oleic acid (C18:1 n-9 Z)	5.3-56
Polyunsaturated fatty acids	1.5-25
Linoleic acid, LA	
(C18:2 n-6 Z)	1.5-25
a-Linolenic acid, ALA (C18:3 n-3)	0.38-0.89
Eicosadienoic acid (C20:2)	0.38-0.89
Arachidonic acid, AA (C20:4 n-6)	0.38-0.89
Dihomo-y-linolenic acid, DGLA (C20:3 n-6)	0.23-0.63
Docosahexadienoic acid, DHA (C22:6 n-3)	0.015-0.5
Cholesterol	0.07-0.19

TABLE 2B-continued

Macromolecular Content and Concentration Ranges of Functional Nutrition Products Derived from Human Mammary Epithelial Cells Cultured With or Without Plasma Cells.	
Macromolecular Fraction	Concentration, g/L
Phospholipids, plasmalogens, sphingolipids	0.075-0.5
Human Milk Oligosaccharides	
Neutral	
TF-LNH (trifucosyllacto-N-hexose)	0.9-4.4
2'-FL (2'-fucosyllactose)	0.9-4.4
DF-LNHII (difucosyllacto-N-hexaose)	0.9-4.4
LNFP I (1 acto-A-fucopentaose I)	0.45-2.2
LNDFHI (lacto-N-difucosylhexaose I)	0.18-2.2
LNT (lacto-A-tetraose)	0.27-1.7
LNnT (lacto-A-neotetraose)	0.4-1.7
DF-L (Difucosyllactose)	0.09-1.1
3-FL (3-fucosyllactose)	0.18-1.7
Acidic	
6'-SL (6'-sialyllactose)	0.18-1.3
DS-LNT (disialyllacto-A-tetraose)	0.04-1.1
FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I)	0.04-0.8
LST c (si alyl-lacto-A-tetraose c)	0.04-0.8
3'-SL (3'-sialyllactose)	0.09-0.33
Lactose	50-98

TABLE 2C

Macromolecular Content and Concentration Ranges of Functional Nutrition Products Derived from Human Mammary Epithelial Cells Cultured With or Without Plasma Cells.	
Macromolecular Fraction	Concentration, g/L
Protein	
P-Casein	0.5-1.5
K-Casein	0.5-0.6
a-Casein	0.1-0.5
a-Lactalbumin	2.7-3.3
Lysozyme	0.2-0.5
Lactoferrin	1-2
Haptocorrin	0.07-0.7
Butyrophilin	0.03-0.05
Osteopontin	0.05-0.2
Mucin MCS	0.5-0.6
Mucin BrE3	0.5-0.7
Lactadherin	0.06-0.07
Immunoglobulins	0.2-2
Secretory IgA	0.2-1.0
Total IgA	0.15-1.6
Total IgG	0.03-0.3
Total IgM	0.01-0.1
Lipids	
Saturated fatty acids	5-34
Palmitic acid (C16:0)	3-18
Stearic acid (C18:0)	0.7-5
Lauric acid (C12:0)	0.5-5
Monounsaturated fatty acids	7-46
Oleic acid (C18:1 n-9 Z)	7-45
Polyunsaturated fatty acids	2-20
Linoleic acid, LA (C18:2n-6 Z)	2-19
a-Linolenic acid, ALA (C18:3 n-3)	0.5-0.7
Eicosadienoic acid (C20:2)	0.5-0.7
Arachidonic acid, AA (C20:4 n-6)	0.5-0.7
Dihomo-γ-linolenic acid, DGLA (C20:3 n-6)	0.3-0.5
Docosahexadienoic acid, DHA (C22:6 n-3)	0.02-0.4
Cholesterol	0.09-0.15
Phospholipids, plasmalogens, sphingolipids	0.1-0.4
Human Milk Oligosaccharides	
Neutral	
TF-LNH (trifucosyllacto-N-hexose)	1-4
2'-FL (2'-fucosyllactose)	1-4
DF-LNHII (difucosyllacto-N-hexaose)	1-4
LNFP I (1 acto-A-fucopentaose I)	0.5-2
LNDFHI (lacto-N-difucosylhexaose I)	0.2-2
LNT (lacto-A-tetraose)	0.3-1.5

TABLE 2C-continued

Macromolecular Content and Concentration Ranges of Functional Nutrition Products Derived from Human Mammary Epithelial Cells Cultured With or Without Plasma Cells.	
Macromolecular Fraction	Concentration, g/L
LNnT (lacto-A-neotetraose)	0.5-1.5
DF-L (Difucosyllactose)	0.1-1
3-FL (3-fucosyllactose) Acidic	0.2-1.5
6'-SL (6'-sialyllactose)	0.2-1.2
DS-LNT (disialyllacto-A-tetraose)	0.05-1
FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I)	0.05-0.7
LST c (sialyl-lacto-A-tetraose c)	0.05-0.7
3'-SL (3'-sialyllactose)	0.1-0.3
Lactose	67-78

In some embodiments, the concentrations of components indicated in Tables 2A-C can vary, each individually, for example, by having a concentration that is greater than that indicated by 0.1 fold, or 0.2 fold, or 0.3 fold, or 0.4 fold, or 0.5 fold, or 0.6 fold, or 0.7 fold, or 0.8 fold, or 0.9 fold, or 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 fold.

In some embodiments, the concentrations of components indicated in Tables 2A-C can vary, each individually, for example, by having a concentration that is less than that indicated by 0.1 fold, or 0.2 fold, or 0.3 fold, or 0.4 fold, or 0.5 fold, or 0.6 fold, or 0.7 fold, or 0.8 fold, or 0.9 fold.

In some embodiments, milk products are contemplated herein that include a subset of the components (i.e., macromolecular fractions) of Tables 2A-C. In other embodiments, milk products contemplated herein can exclude one or more of the components (i.e., macromolecular fractions) of Tables 2A-C.

In some embodiments, milk products are contemplated herein that further comprise serum albumin. In some embodiments, the serum albumin can have a concentration of about 0.025-3.5 g/L. In some embodiments, the serum albumin can have a concentration of about 0.01-2 g/L. In some embodiments, the serum albumin can have a concentration of about 0.15-1 g/L, and in some embodiments, the serum albumin can have a concentration of about 0.2-0.7 g/L.

One aspect of the present disclosure is directed to a milk product comprising about 6-14 grams per liter (g/L) protein components, about 18-89 g/L lipid components, about 7-14 g/L human milk oligosaccharides (HMOs), and about 64-77 g/L lactose, wherein at least one of the protein components, lipid components, HMOs, and lactose is produced by cultured human mammary epithelial cells.

In some embodiments, the protein component comprises about 55-65% dry weight of the milk product. In some embodiments, the protein component comprises one or more of beta-casein, kappa-casein and alpha-casein, and in some embodiments, the beta-casein can have a concentration of about 0.5-1.5 g/L, the kappa-casein can have a concentration of about 0.5-0.6 g/L and the alpha-casein can have a concentration of about 0.1-0.5 g/L in the milk product. In some embodiments of the milk product, the beta-casein, kappa-casein and alpha-casein together comprise about 35-45% dry weight percent of the protein component. In some embodiments, the beta-casein comprises greater than about 50% of total casein content.

In some embodiments, the protein component further comprises, for example, one or more of alpha-lactalbumin, lysozyme, lactoferrin, haptocorrin, butyrophilin, osteopontin, mucin MC5, mucin BrE3, and lactadherin. Some

embodiments further comprise serum albumin. In some embodiments of the milk product, the alpha-lactalbumin can have a concentration of about 2.7-3.3 g/L, and in some embodiments, the lysozyme can have a concentration of about 0.2-0.5 g/L, and in some embodiments, the lactoferrin can have a concentration of about 1.0-2.0 g/L. In some embodiments, the haptocorrin can have a concentration of about 0.07-0.7 g/L, and in some embodiments, the butyrophilin can have a concentration of about 0.03-0.05 g/L. In some embodiments, the osteopontin can have a concentration of about 0.05-0.2 g/L, and in some embodiments, the mucin MC5 can have a concentration of about 0.5-0.6 g/L. In some embodiments, the mucin BrE3 can have a concentration of about 0.5-0.7 g/L, and in some embodiments, the lactadherin can have a concentration of about 0.06-0.07 g/L. In some embodiments serum albumin can have a concentration of about 0.025-3.5 g/L, or about 0.01-2 g/L, or about 0.15-1 g/L, or about 0.2-0.7 g/L.

In some embodiments of the milk product, the protein components are of human origin.

In some embodiments, the lipid component comprises one or more of saturated fatty acids, monounsaturated fatty acids, polyunsaturated fatty acids, cholesterol, phospholipids, plasmalogens, and sphingolipids. In some embodiments of the milk product, the saturated fatty acids can have a concentration of about 5-34 g/L. The saturated fatty acids can comprise, for example, one or more of palmitic acid, stearic acid and lauric acid and combinations thereof. In some embodiments, the palmitic acid comprises at least about 50% sn-2 form. In some embodiments of the milk product, palmitic acid can have a concentration of about 3-18 g/L, in some embodiments, stearic acid can have a concentration of about 0.7-5 g/L, and in some embodiments, lauric acid can have a concentration of about 0.5-5 g/L.

In some embodiments, monounsaturated fatty acids can have a concentration of about 7-46 g/L of the milk product. The saturated fatty acid component can comprise, for example, oleic acid, in some embodiments, the oleic acid comprises at least about 50% sn-1 form, and in some embodiments, oleic acid can have a concentration of about 7-45 g/L in the milk product.

In some embodiments of the milk product, polyunsaturated fats can have a concentration of about 2-20 g/L. The polyunsaturated fats can comprise, for example, one or more of linoleic acid, alpha-linolenic acid, eicosadienoic acid, arachidonic acid, dihomo-gamma-linolenic acid, and docosahexadienoic acid. In some embodiments, the linoleic acid comprises at least about 50% sn-3 form, and in some embodiments, linoleic acid can have a concentration of about 2-19 g/L, and in some embodiments, alpha-linolenic acid can have a concentration of about 0.5-0.7 g/L. In some embodiments, eicosadienoic acid can have a concentration of about 0.5-0.7 g/L, and in some embodiments arachidonic acid can have a concentration of about 0.5-0.7 g/L. In some embodiments, dihomo-gamma-linolenic acid can have a concentration of about 0.3-0.5 g/L, and in some embodiments, docosahexadienoic acid can have a concentration of about 0.02-0.4 g/L. Some embodiments of the milk product comprise all of the above polyunsaturated fats, wherein linoleic acid can have a concentration of about 2-19 g/L, alpha-linolenic acid can have a concentration of about 0.5-0.7 g/L, eicosadienoic acid can have a concentration of about 0.5-0.7 g/L, arachidonic acid can have a concentration of about 0.5-0.7 g/L, dihomo-gamma-linolenic acid can have a concentration of about 0.3-0.5 g/L, and docosahexadienoic acid can have a concentration of about 0.02-0.4 g/L in the milk product.

In some embodiments, cholesterol can have a concentration of about 0.09-0.15 g/L in milk product, and in some embodiments, phospholipids, plasmalogens, and sphingolipids together can have a concentration of about 0.1-0.4 g/L in the milk product.

In some embodiments, the milk product comprises one or more neutral oligosaccharides, one or more acidic oligosaccharides or one or more each of neutral oligosaccharides and acidic oligosaccharides.

In some embodiments, the one or more neutral oligosaccharides comprise TF-LNH (trifucosyllacto-N-hexose), 2'-FL (2'-fucosyllactose), DF-LNHII (difucosyllacto-N-hexaose), LNFP I (lacto-N-fucopentaose I), LNDFHI (lacto-N-difucosylhexaose I), LNT (lacto-N-tetraose), LNnT (lacto-N-neotetraose), DF-L (Difucosyllactose), and 3-FL (3-fucosyllactose).

In some embodiments, the one or more acidic oligosaccharides comprise 6'-SL (6'-sialyllactose), DS-LNT (disialyllacto-N-tetraose), FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), LST c (sialyl-lacto-N-tetraose c), and 3'-SL (3'-sialyllactose).

In some embodiments, the one or more neutral oligosaccharides comprises TF-LNH (trifucosyllacto-N-hexose), which oligosaccharide can have a concentration of about 1-4 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises 2'-FL (2'-fucosyllactose), which oligosaccharide can have a concentration of about 1-4 g/L in the milk product, and in some embodiments, the one or more neutral oligosaccharides comprises DF-LNH II (difucosyllacto-N-hexaose), which oligosaccharide can have a concentration of about 1-4 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises LNFP I (lacto-N-fucopentaose I), which oligosaccharide can have a concentration of about 0.5-2 g/L in the milk product, and in some embodiments, the one or more neutral oligosaccharides comprises LNDFH I (lacto-N-difucosylhexaose I), which oligosaccharide can have a concentration of about 0.2-2 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises LNT (lacto-N-tetraose), which oligosaccharide can have a concentration of about 0.3-1.5 g/L in the milk product, and in some embodiments, the one or more neutral oligosaccharides comprises LNnT (lacto-N-neotetraose), which oligosaccharide can have a concentration of about 0.5-1.5 g/L in the milk product. In some embodiments, the one or more neutral oligosaccharides comprises DF-L (Difucosyllactose), which oligosaccharide can have a concentration of about 0.1-1 g/L in the milk product, and in some embodiments, the one or more neutral oligosaccharides comprises 3-FL (3-fucosyllactose), which oligosaccharide can have a concentration of about 0.2-1.5 g/L of the milk product.

In some embodiments, the one or more acidic oligosaccharides comprises 6'-SL (6'-sialyllactose), which oligosaccharide can have a concentration of about 0.2-1.2 g/L in the milk product, and in some embodiments, the one or more acidic oligosaccharides comprises DS-LNT (disialyllacto-N-tetraose), which oligosaccharide can have a concentration of about 0.05-1 g/L in the milk product. In some embodiments, the one or more acidic oligosaccharides comprises FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), which oligosaccharide can have a concentration of about 0.05-0.7 g/L in the milk product, and in some embodiments, the one or more acidic oligosaccharides comprises LST c (sialyl-lacto-N-tetraose c), which oligosaccharide can have a concentration of about 0.05-0.7 g/L in the milk product. In some embodiments, the one or more acidic oligosaccharides com-

prises 3'-SL (3'-sialyllactose), which oligosaccharide can have a concentration of about 0.1-0.3 g/L in the milk product.

In some embodiments of the milk product the one or more neutral oligosaccharides comprise TF-LNH (trifucosyllacto-N-hexose), 2'-FL (2'-fucosyllactose), DF-LNH II (difucosyllacto-N-hexaose), LNFP I (lacto-N-fucopentaose I), LNDFHI (lacto-N-difucosylhexaose I), LNT (lacto-N-tetraose), LNnT (lacto-N-neotetraose), DF-L (Difucosyllactose), and 3-FL (3-fucosyllactose) and the one or more acidic oligosaccharides comprises 6'-SL (6'-sialyllactose), DS-LNT (disialyllacto-N-tetraose), FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), LST c (sialyl-lacto-N-tetraose c), and 3'-SL (3'-sialyllactose).

In some embodiments, the milk product comprises about 1-4 g/L TF-LNH (trifucosyllacto-N-hexose), comprises about 1-4 g/L 2'-FL (2'-fucosyllactose), comprises about 1-4 g/L DF-LNH II (difucosyllacto-N-hexaose), comprises about 0.5-2 g/L LNFP I (lacto-N-fucopentaose I), comprises about 0.2-2 g/L LNDFH I (lacto-N-difucosylhexaose I), comprises about 0.3-1.5 g/L LNT (lacto-N-tetraose), comprises about 0.5-1.5 g/L LNnT (lacto-N-neotetraose), comprises about 0.1-1 g/L DF-L (Difucosyllactose), comprises about 0.2-1.5 g/L 3-FL (3-fucosyllactose), comprises about 0.2-1.2 g/L 6'-SL (6'-sialyllactose), comprises about 0.05-1 g/L DS-LNT (disialyllacto-N-tetraose), comprises about 0.05-0.7 g/L FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), comprises about 0.05-0.7 g/L LST c (sialyl-lacto-N-tetraose c), and comprises about 0.1-0.3 g/L 3'-SL (3'-sialyllactose).

In some embodiments, the neutral oligosaccharides content comprises at least about 2-fold, or about 3-fold, or about 4-fold, or about 5-fold, or about 6-fold, or about 7-fold, or about 8-fold, or about 9-fold, or about 10-fold, or about 11-fold, or about 12-fold, or about 13-fold, or about 14-fold, or about 15-fold more by weight than acidic oligosaccharide content.

In another aspect of the disclosure, a milk product is provided, comprising about 614 grams per liter (g/L) protein components, about 18-89 g/L lipid components, about 7-14 g/L human milk oligosaccharides (HMOs), and about 64-77 g/L lactose, wherein the protein components comprise one or more of beta-casein, kappa-casein, and alpha-casein, alphasalalbumin, lysozyme, lactoferrin, haptocorrin, butyrophilin, osteopontin, mucin MC5, mucin BrE3, and lactadherin, wherein the lipid components comprise one or more of palmitic acid, stearic acid and lauric acid, oleic acid, linoleic acid, alpha-linolenic acid, eicosadienoic acid, arachidonic acid, dihomo-gamma-linolenic acid, docosahexadienoic acid, cholesterol, phospholipids, plasmalogens and sphingolipids, wherein the human milk oligosaccharides comprise one or more of TF-LNH (trifucosyllacto-N-hexose), 2'-FL (2'-fucosyllactose), DF-LNHII (difucosyllacto-N-hexaose), LNFP I (lacto-N-fucopentaose I), LNDFHI (lacto-N-difucosylhexaose I), LNT (lacto-N-tetraose), LNnT (lacto-N-neotetraose), DF-L (Difucosyllactose), and 3-FL (3-fucosyllactose), 6'-SL (6'-sialyllactose), DS-LNT (disialyllacto-N-tetraose), FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), LST c (sialyl-lacto-N-tetraose c), and 3'-SL (3'-sialyllactose), and wherein at least one of the protein components, lipid components, HMOs, and lactose is produced by cultured human mammary epithelial cells. In some embodiments, the milk product further comprises serum albumin.

In some embodiments, the milk product comprises about 0.5-1.5 g/L beta-casein, comprises about 0.5-0.6 g/L kappa-casein, comprises about 0.1-0.5 g/L alpha-casein, comprises

about 2.7-3.3 g/L alpha-lactalbumin, comprises about 0.2-0.5 g/L lysozyme, comprises about 1.0-2.0 g/L lactoferrin, comprises about 0.07-0.7 g/L haptocorrin, comprises about 0.03-0.05 g/L butyrophilin, comprises about 0.05-0.2 g/L osteopontin, comprises about 0.5-0.6 g/L mucin MC5, comprises about 0.5-0.7 g/L mucin BrE3, comprises about 0.06-0.07 g/L lactadherin, comprises about 2-19 g/L linoleic acid, comprises about 0.5-0.7 g/L alpha-linolenic acid, comprises about 0.5-0.7 g/L eicosadienoic acid, comprises about 0.5-0.7 g/L arachidonic acid, comprises about 0.3-0.5 g/L dihomo-gamma-linolenic acid, comprises about 0.02-0.4 g/L docosahexadienoic acid, comprises about 0.09-0.15 g/L cholesterol, together comprise about 0.1-0.4 g/L phospholipids, plasmalogens and sphingolipids, comprises about 1-4 g/L TF-LNH (trifucosyllacto-N-hexose), comprises about 1-4 g/L 2'-FL (2'-fucosyllactose), comprises about 1-4 g/L DF-LNH II (difucosyllacto-N-hexaose), comprises about 0.5-2 g/L LNFP I (lacto-N-fucopentaose I), comprises about 0.2-2 g/L LNDFHI (lacto-N-difucosylhexaose I), comprises about 0.3-1.5 g/L LNT (lacto-N-tetraose), comprises about 0.5-1.5 g/L LNnT (lacto-N-neotetraose), comprises about 0.1-1 g/L DF-L (Difucosyllactose), comprises about 0.2-1.5 g/L 3-FL (3-fucosyllactose), comprises about 0.2-1.2 g/L 6'-SL (6'-sialyllactose), comprises about 0.05-1 g/L DS-LNT (disialyllacto-N-tetraose), comprises about 0.05-0.7 g/L FS-LNnH I (fucosyl-sialyl-lacto-N-neohexaose I), comprises about 0.05-0.7 g/L LST c (sialyl-lacto-N-tetraose c), and comprises about 0.1-0.3 g/L 3'-SL (3'-sialyllactose). In some embodiments, the milk product further comprises serum albumin, which serum albumin has a concentration of about 0.025-3.5 g/L, or about 0.01-2 g/L, or about 0.15-1 g/L, or about 0.3-0.7 g/L.

In another aspect of the disclosure, a milk product is provided comprising about 3-15 percent protein by weight, about 9-92 percent lipid by weight, about 4-15 percent by weight human milk oligosaccharides (HMOs), and about 33-80 percent by weight lactose, wherein at least one of the protein, lipid, HMOs, and lactose is produced by cultured human mammary epithelial cells.

In some embodiments of some aspects of the disclosure, the milk product comprises at least about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, or about 99% of the overall macromolecular composition of human breast milk.

In some embodiments of the milk product of the disclosure, non-protein nitrogen content is at least about 10%, or at least about 15%, or at least about 20%, or at least about 25%, or at least about 30% of total nitrogen content.

In some embodiments, the milk product comprises about 500-1150 kcal/L available energy content, and in some embodiments, between about 40-55% of the available energy content is from lipids. In some embodiments, the milk product comprises between about 95.8 and 195.2 g/L macromolecular content.

In another aspect of the disclosure, a containerized milk product is provided, comprising the milk product according to some embodiments of the disclosure that is packaged into a container.

In another aspect of the disclosure, a frozen milk product is provided, comprising the milk product according to some embodiments of the disclosure that has been frozen. In some embodiments, the frozen milk product is packaged into a container making a containerized frozen milk product.

In another aspect of the disclosure, a lyophilized milk product is provided, comprising the milk product according

35

to some embodiments of the disclosure that has been lyophilized. In some embodiments, the lyophilized milk product is packaged into a container making a containerized lyophilized milk product.

In another aspect of the disclosure, an extracted milk product is provided, comprising one or more components extracted from the milk product according to some embodiments of the disclosure. Non-limiting examples of components that can be extracted from the milk product include protein, lipid, carbohydrate, vitamin, and/or mineral content. In some embodiments, the one or more components extracted from the collected milk product are lyophilized or concentrated to produce a lyophilized or a concentrated extracted milk product component. In some embodiments, the one or more components extracted from the collected milk product are concentrated by membrane filtration or reverse osmosis, while in other embodiments, the whole, un-extracted milk product is concentrated by membrane filtration or reverse osmosis. In some embodiments, the one or more extracted components from the collected milk product comprise milk protein, lipid, carbohydrate, vitamin, and minerals. In some embodiments, the one or more extracted milk product components are packaged in a container, and in some embodiments, concentrated whole, un-extracted milk product is packaged in a container.

In some embodiments, the container is sterile, vacuum-sealed, or classified as food grade, or any combination thereof. Non-limiting examples of containers comprise a canister, ajar, a bottle, a bag, a box, or a pouch. In some embodiments, the container is a canister, ajar, a bottle, a bag, a box, or a pouch.

Immunoglobulins

In some embodiments, the cultured milk product further comprises one or more immunoglobulins or sIgA. In some embodiments, the cultured milk product comprises one or more of IgA, IgG, and IgM. In some embodiments, the cultured milk product comprises IgA2 (secretory) and IgA1 (non-secretory). In some embodiments, the plasma cells of the cell construct produce IgA and the mammary epithelial cells process the IgA to yield sIgA (IgA2). sIgA comprises a secretory component, the extracellular domain of the polymeric Ig receptor, attached to an IgA. Mammary epithelial cells process IgA by cleaving the extracellular domain of a polymeric Ig receptor to generate sIgA. In some embodiments, sIgA is secreted from the apical surface of the mammary epithelial cell.

In certain embodiments, the immunoglobulins bind to an antigen of a microorganism (i.e., bacterium or virus). In certain embodiments, the immunoglobulins bind to viral or bacterial antigens capable of causing an infectious disease in humans. In certain embodiments, the immunoglobulins bind to viral or bacterial antigens that cause infections of respiratory or gastrointestinal epithelium. In certain embodiments, the immunoglobulins bind antigens from microorganisms that cause enterocolitis or sepsis in infants.

In certain embodiments, the milk product comprises about 0.2-1.0 g/L secretory IgA. In certain embodiments, the milk product comprises about 0.2, 0.25, 0.3, 0.35, 0.4, 0.45, 0.5, 0.55, 0.6, 0.65, 0.7, 0.75, 0.8, 0.85, 0.9, 0.95, or 1.0 g/L secretory IgA. In certain embodiments, the milk product comprises about 0.15-1.6 g/L total IgA. In certain embodiments, the milk product comprises about 0.15, 0.2, 0.25, 0.3, 0.35, 0.4, 0.45, 0.5, 0.55, 0.6, 0.65, 0.7, 0.75, 0.8, 0.85, 0.9, 0.95, 1.0, 1.05, 1.1, 1.15, 1.2, 1.25, 1.3, 1.35, 1.4, 1.45, 1.5, 1.55, or 1.6 g/L total IgA.

In some embodiments that comprise IgG, the milk product comprises about 0.03-0.3 g/L IgG. In certain embodi-

36

ments, that comprise IgG, the milk product comprises about 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.11, 0.12, 0.13, 0.14, 0.15, 0.16, 0.17, 0.18, 0.19, 0.2, 0.21, 0.22, 0.23, 0.24, 0.25, 0.26, 0.27, 0.28, 0.29, 0.3 g/L. In some embodiments that comprise IgM, the milk product comprises about 0.01-0.1 g/L IgM. In certain embodiments, the milk product comprises about 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.1 g/L IgM. In some embodiments, the milk product comprises about 0.2-2.0 percent by weight total immunoglobulins. In certain embodiments, the milk product comprises 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2.0 percent by weight total immunoglobulins.

Included herein are immunotherapeutic compositions comprising (a) IgA and sIgA derived from the secreted products of mammary epithelial cells co-cultured with plasma cells, and (b) a pharmaceutically acceptable excipient. In some embodiments, the IgA and sIgA are isolated from the product resulting from co-culturing mammary epithelial cells and plasma cells (e.g., a cultured milk product).

In some embodiments, IgA is produced by the plasma cells in the cell construct. The IgA binds to a receptor (polymeric Ig receptor) on the basal surface of mammary epithelial cells. The IgA and receptor are transported into the mammary epithelial cells. The mammary cells process the IgA. The extracellular domain of the receptor bound to the IgA (secretory component) is cleaved by a proteinase within the mammary epithelial cells and the IgA bound to secretory component is secreted from the apical surface of the mammary epithelial cells to yield sIgA. In some embodiments, the sIgA is secreted as part of the cultured milk product from the apical surface of the mammary cells into the apical compartment. In some embodiments, the sIgA is isolated from the cultured milk product. In some embodiments, the sIgA is not isolated from the cultured milk product.

Basal Culture Media and Lactogenic Media

In some embodiments, the culture medium comprises a carbon source, a chemical buffering system, one or more essential amino acids, one or more vitamins and/or cofactors, and one or more inorganic salts. In some embodiments, the carbon source, chemical buffering system, one or more essential amino acids, one or more vitamins and/or cofactors, and/or one or more inorganic salts are food grade.

In some embodiments, the culture medium is lactogenic culture medium. In some embodiments, the culture medium further comprises prolactin (e.g., mammalian prolactin, e.g., human prolactin), linoleic and alpha-linoleic acid, estrogen and/or progesterone. For example, in some embodiments, the culture medium comprises prolactin (or prolactin is added) in an amount from about 20 ng/mL to about 200 ng/L of culture medium, e.g., about 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, or 200 ng/mL or any value or range therein. In some embodiments, the culture medium comprises prolactin (or prolactin is added) in an amount from about 20 ng/mL to about 195 ng/mL, about 50 ng/mL to about 150 ng/mL, about 25 ng/mL to about 175 ng/mL, about 45 ng/mL to about 200 ng/mL, or about 75 ng/mL to about 190 ng/mL of culture medium. In some embodiments, the culture medium further comprises other factors to improve efficiency, including, but not limited to, insulin, an epidermal growth factor, and/or a hydrocortisone.

In some embodiments, the culture medium comprises a carbon source in an amount from about 1 g/L to about 15 g/L of culture medium (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14 or 15 g/L or any value or range therein), or about

37

1, 2, 3, 4, 5 or 6 g/L to about 7, 8, 9, or 10, 11, 12, 13, 14 or 15 g/L of the culture medium. Non-limiting examples of a carbon source include glucose and/or pyruvate. For example, in some embodiments, the culture medium comprises glucose in an amount from about 1g/L to about 12 g/L of culture medium, e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, or 12 g/L or any value or range therein. In some embodiments, the culture medium comprises glucose in an amount from about 1 g/L to about 6 g/L, about 4 g/L to about 12 g/L, about 2.5 g/L to about 10.5 g/L, about 1.5 g/L to about 11.5 g/L, or about 2 g/L to about 10 g/L of culture medium. In some embodiments, the culture medium comprises glucose in an amount from about 1, 2, 3, or 4 g/L to about 5, 6, 7, 8, 9, 10, 11, or 12 g/L or about 1, 2, 3, 4, 5, or 6 g/L to about 7, 8, 9, 10, 11, or 12 g/L. In some embodiments, the culture medium comprises pyruvate in an amount from about 5 g/L to about 15 g/L of culture medium, e.g., about 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, or 15 g/L or any value or range therein. In some embodiments, the culture medium comprises pyruvate in an amount from about 5 g/L to about 14.5 g/L, about 10 g/L to about 15 g/L, about 7.5 g/L to about 10.5 g/L, about 5.5 g/L to about 14.5 g/L, or about 8 g/L to about 10 g/L of culture medium. In some embodiments, the culture medium comprises pyruvate in an amount from about 5, 6, 7, or 8 g/L to about 9, 10, 11, 12, 13, 14 or 15 g/L or about 5, 6, 7, 8, 9, or 10 g/L to about 11, 12, 13, 14 or 15 g/L.

In some embodiments, the culture medium comprises a chemical buffering system in an amount from about 1 g/L to about 4 g/L (e.g., about 1, 1.5, 2, 2.5, 3, 3.5, or 4 g/L or any value or range therein) of culture medium or about 10 mM to about 25 mM (e.g., about 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 mM or any value or range therein). In some embodiments, the chemical buffering system includes, but is not limited to, sodium bicarbonate and/or 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES). For example, in some embodiments, the culture medium comprises sodium bicarbonate in an amount from about 1 g/L to about 4 g/L of culture medium, e.g., about 1, 1.5, 2, 2.5, 3, 3.5, or 4 g/L or any value or range therein. In some embodiments, the culture medium comprises sodium bicarbonate in an amount from about 1 g/L to about 3.75 g/L, about 1.25 g/L to about 4 g/L, about 2.5 g/L to about 3 g/L, about 1.5 g/L to about 4 g/L, or about 2 g/L to about 3.5 g/L of culture medium. In some embodiments, the culture medium comprises HEPES in an amount from about 10 mM to about 25 mM, e.g., about 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 mM or any value or range therein. In some embodiments, the culture medium comprises HEPES in an amount from about 11 mM to about 25 mM, about 10 mM to about 20 mM, about 12.5 mM to about 22.5 mM, about 15 mM to about 20.75 mM, or about 10 mM to about 20 mM.

In some embodiments, the culture medium comprises one or more essential amino acids in an amount from about 0.5 mM to about 5 mM (e.g., about 0.5, 1, 1.5, 2, 2.5, 3, 3.5, 4, 4.5, or 5 mM or any value or range therein) or about 0.5, 1, 1.5, 2 mM to about 2.5, 3, 3.5, 4, 4.5, or 5 mM. In some embodiments, the one or more essential amino acids is histidine, isoleucine, leucine, lysine, methionine, phenylalanine, threonine, tryptophan, valine, and/or arginine. For example, in some embodiments, the culture medium comprises arginine in an amount from about 0.5 mM to about 5 mM, e.g., about 0.5, 1, 1.5, 2, 2.5, 3, 3.5, 4, 4.5, or 5 mM or any value or range therein. In some embodiments, the culture medium comprises an essential amino acids in an amount from about 0.5 mM to about 4.75 mM, about 2 mM

38

to about 3.5 mM, about 0.5 mM to about 3.5 mM, about 1 mM to about 5 mM, or about 3.5 mM to about 5 mM.

In some embodiments, the culture medium comprises one or more vitamins and/or cofactors in an amount from about 0.01 μ M to about 50 μ M (e.g., about 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.1, 2.2, 2.3, 2.4, 2.5, 3, 4, 5, 6, 7, 8, 9, 10, 12.5, 15, 17.5, 20, 25, 30, 35, 40, 45, 46, 47, 48, 49, 49.025, 49.05, 49.075, or 50 μ M or any value or range therein) or about 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, or 0.9 μ M to about 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.1, 2.2, 2.3, 2.4, 2.5, 3, 4, 5, 6 μ M or about 0.02, 0.025, 0.05, 0.075, 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 9, 10 μ M to about 12.5, 15, 17.5, 20, 25, 30, 35, 40, 45, 46, 47, 48, 49, 49.025, 49.05, 49.075, or 50 μ M. In some embodiments, one or more vitamins and/or cofactors include, but are not limited to, thiamine and/or riboflavin. For example, in some embodiments, the culture medium comprises thiamine in an amount from about 0.025 μ M to about 50 μ M, e.g., about 0.025, 0.05, 0.075, 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12.5, 15, 17.5, 20, 25, 30, 35, 40, 45, 46, 47, 48, 49, 49.025, 49.05, 49.075, or 50 μ M or any value or range therein. In some embodiments, the culture medium comprises thiamine in an amount from about 0.025 μ M to about 45.075 μ M, about 1 μ M to about 40 μ M, about 5 μ M to about 35.075 μ M, about 10 μ M to about 50 μ M, or about 0.05 μ M to about 45.5 μ M. In some embodiments, the culture medium comprises riboflavin in an amount from about 0.01 μ M to about 3 μ M, e.g., about 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.1, 2.2, 2.3, 2.4, 2.5, 2.6, 2.7, 2.8, 2.9, or 3 μ M or any value or range therein. In some embodiments, the culture medium comprises riboflavin in an amount from about 0.01 μ M to about 2.05 μ M, about 1 μ M to about 2.95 μ M, about 0.05 μ M to about 3 μ M, about 0.08 μ M to about 1.55 μ M, or about 0.05 μ M to about 2.9 μ M.

In some embodiments, the culture medium comprises one or more inorganic salts in an amount from about 100 mg/L to about 150 mg/L of culture medium (e.g., about 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 mg/L or any value or range therein) or about 100 mg/L to about 150 mg/L of culture medium (e.g., about 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 mg/L or any value or range therein). In some embodiments, one or more inorganic salts include, but are not limited to, calcium and/or magnesium. For example, in some embodiments, the culture medium comprises calcium in an amount from about 100 mg/L to about 150 mg/L of culture medium, e.g., about 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 mg/L or any value or range therein. In some embodiments, the culture medium comprises arginine in an amount from about 100 mg/L to about 125 mg/L, about 105 mg/L to about 150 mg/L, about 120 mg/L to about 130 mg/L, or about 100 mg/L to about 145 mg/L of culture medium. In some embodiments, the culture medium comprises magnesium in an amount from about 0.01 mM to about 1 mM, e.g., about 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 0.91, 0.92, 0.93, 0.94, 0.95, 0.96, 0.97, 0.98, 0.99, or 1 mM or any value or range therein. In some embodiments, the culture medium comprises magnesium in an amount from about 0.05 mM to about 1 mM, about 0.01 mM to about 0.78 mM, about 0.5 mM to about 1 mM, about 0.03 mM to about 0.75 mM, or about 0.25 mM to about 0.95 mM.

In some embodiments, the culture medium comprises a carbon source in an amount from about 1 g/L to about 15 g/L

39

of culture medium (e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14 or 15 g/L or any value or range therein), or about 1, 2, 3, 4, 5 or 6 g/L to about 7, 8, 9, or 10, 11, 12, 13, 14 or 15 g/L of the culture medium. In some embodiments, the carbon source includes, but is not limited to, glucose and/or pyruvate. For example, in some embodiments, the culture medium comprises glucose in an amount from about 1 g/L to about 12 g/L of culture medium, e.g., about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, or 12 g/L or any value or range therein. In some embodiments, the culture medium comprises glucose in an amount from about 1 g/L to about 6 g/L, about 4 g/L to about 12 g/L, about 2.5 g/L to about 10.5 g/L, about 1.5 g/L to about 11.5 g/L, or about 2 g/L to about 10 g/L of culture medium. In some embodiments, the culture medium comprises pyruvate at an amount of about 5 g/L to about 15 g/L of culture medium, e.g., about 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, or 15 g/L or any value or range therein. In some embodiments, the culture medium comprises pyruvate in an amount from about 5 g/L to about 14.5 g/L, about 10 g/L to about 15 g/L, about 7.5 g/L to about 10.5 g/L, about 5.5 g/L to about 14.5 g/L, or about 8 g/L to about 10 g/L of culture medium.

In some embodiments, the culture medium comprises a chemical buffering system in an amount from about 1 g/L to about 4 g/L (e.g., about 1, 1.5, 2, 2.5, 3, 3.5, or 4 g/L or any value or range therein) of culture medium or about 10 mM to about 25 mM (e.g., about 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 mM or any value or range therein). In some embodiments, the chemical buffering system includes, but is not limited to, sodium bicarbonate and/or HEPES. For example, in some embodiments, the culture medium comprises sodium bicarbonate in an amount from about 1 g/L to about 4 g/L of culture medium, e.g., about 1, 1.5, 2, 2.5, 3, 3.5, or 4 g/L or any value or range therein. In some embodiments, the culture medium comprises sodium bicarbonate in an amount from about 1 g/L to about 3.75 g/L, about 1.25 g/L to about 4 g/L, about 2.5 g/L to about 3 g/L, about 1.5 g/L to about 4 g/L, or about 2 g/L to about 3.5 g/L of culture medium. In some embodiments, the culture medium comprises HEPES in an amount from about 10 mM to about 25 mM, e.g., about 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 mM or any value or range therein. In some embodiments, the culture medium comprises HEPES in an amount from about 1 mM to about 25 mM, about 10 mM to about 20 mM, about 12.5 mM to about 22.5 mM, about 15 mM to about 20.75 mM, or about 10 mM to about 20 mM.

In some embodiments, the culture medium comprises one or more essential amino acids in an amount from about 0.5 mM to about 5 mM (e.g., about 0.5, 1, 1.5, 2, 2.5, 3, 3.5, 4, 4.5, or 5 mM or any value or range therein) or about 0.5, 1, 1.5, 2 mM to about 2.5, 3, 3.5, 4, 4.5, or 5 mM. In some embodiments, one or more essential amino acids is arginine and/or cysteine. For example, in some embodiments, the culture medium comprises arginine in an amount from about 0.5 mM to about 5 mM, e.g., about 0.5, 1, 1.5, 2, 2.5, 3, 3.5, 4, 4.5, or 5 mM or any value or range therein. In some embodiments, the culture medium comprises arginine in an amount from about 0.5 mM to about 4.75 mM, about 2 mM to about 3.5 mM, about 0.5 mM to about 3.5 mM, about 1 mM to about 5 mM, or about 3.5 mM to about 5 mM. For example, in some embodiments, the culture medium comprises cysteine in an amount from about 0.5 mM to about 5 mM, e.g., about 0.5, 1, 1.5, 2, 2.5, 3, 3.5, 4, 4.5, or 5 mM or any value or range therein. In some embodiments, the culture medium comprises cysteine in an amount from about 0.5 mM to about 4.75 mM, about 2 mM to about 3.5 mM,

40

about 0.5 mM to about 3.5 mM, about 1 mM to about 5 mM, or about 3.5 mM to about 5 mM.

In some embodiments, the culture medium comprises one or more vitamins and/or cofactors in an amount from about 0.01 μ M to about 50 μ M (e.g., about 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.1, 2.2, 2.3, 2.4, 2.5, 3, 4, 5, 6, 7, 8, 9, 10, 12.5, 15, 17.5, 20, 25, 30, 35, 40, 45, 46, 47, 48, 49, 49.025, 49.05, 49.075, or 50 μ M or any value or range therein) or about 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, or 0.9 μ M to about 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.1, 2.2, 2.3, 2.4, 2.5, 3, 4, 5, 6 μ M or about 0.02, 0.025, 0.05, 0.075, 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 9, 10 μ M to about 12.5, 15, 17.5, 20, 25, 30, 35, 40, 45, 46, 47, 48, 49, 49.025, 49.05, 49.075, or 50 μ M. In some embodiments, one or more vitamins and/or cofactors includes, but is not limited to, thiamine and/or riboflavin. For example, in some embodiments, the culture medium comprises thiamine in an amount from about 0.025 μ M to about 50 μ M, e.g., 0.025, 0.05, 0.075, 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12.5, 15, 17.5, 20, 25, 30, 35, 40, 45, 46, 47, 48, 49, 49.025, 49.05, 49.075, or 50 μ M or any value or range therein. In some embodiments, the culture medium comprises thiamine in an amount from about 0.025 μ M to about 45.075 μ M, about 1 μ M to about 40 μ M, about 5 μ M to about 35.075 μ M, about 10 μ M to about 50 μ M, or about 0.05 μ M to about 45.5 μ M. In some embodiments, the culture medium comprises riboflavin in an amount from about 0.01 μ M to about 3 μ M, e.g., 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.1, 2.2, 2.3, 2.4, 2.5, 2.6, 2.7, 2.8, 2.9, or 3 μ M or any value or range therein. In some embodiments, the culture medium comprises riboflavin in an amount from about 0.01 μ M to about 2.05 μ M, about 1 μ M to about 2.95 μ M, about 0.05 μ M to about 3 μ M, about 0.08 μ M to about 1.55 μ M, or about 0.05 μ M to about 2.9 μ M.

In some embodiments, the culture medium comprises one or more inorganic salts in an amount from about 100 mg/L to about 150 mg/L of culture medium (e.g., about 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 mg/L or any value or range therein) or about 100 mg/L to about 150 mg/L of culture medium (e.g., about 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 mg/L or any value or range therein). In some embodiments, exemplary one or more inorganic salts is calcium and/or magnesium. For example, in some embodiments, the culture medium comprises calcium in an amount from about 100 mg/L to about 150 mg/L of culture medium, e.g., about 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, or 150 mg/L or any value or range therein. In some embodiments, the culture medium comprises arginine in an amount from about 100 mg/L to about 125 mg/L, about 105 mg/L to about 150 mg/L, about 120 mg/L to about 130 mg/L, or about 100 mg/L to about 145 mg/L of culture medium. In some embodiments, the culture medium comprises magnesium in an amount from about 0.01 mM to about 1 mM, e.g., about 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 0.91, 0.92, 0.93, 0.94, 0.95, 0.96, 0.97, 0.98, 0.99, or 1 mM or any value or range therein. In some embodiments, the culture medium comprises magnesium in an amount from about 0.05 mM to about 1 mM, about 0.01 mM to about 0.78 mM, about 0.5 mM to about 1 mM, about 0.03 mM to about 0.75 mM, or about 0.25 mM to about 0.95 mM.

41

In some embodiments, the carbon source, chemical buffering system, one or more essential amino acids, one or more vitamins and/or cofactors, and/or one or more inorganic salts is food grade.

In some embodiments, the culture medium is lactogenic culture medium, e.g., the culture medium further comprises prolactin (e.g., mammalian prolactin, e.g., human prolactin). For example, in some embodiments, the culture medium comprises prolactin (or prolactin is added) in an amount from about 20 ng/mL to about 200 ng/L of culture medium, e.g., about 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, or 200 ng/mL or any value or range therein. In some embodiments, the culture medium comprises prolactin (or prolactin is added) in an amount from about 20 ng/mL to about 195 ng/mL, about 50 ng/mL to about 150 ng/mL, about 25 ng/mL to about 175 ng/mL, about 45 ng/mL to about 200 ng/mL, or about 75 ng/mL to about 190 ng/mL of culture medium. In some embodiments, the methods further comprise adding prolactin to the culture medium, thereby providing a lactogenic culture medium. In some embodiments, the prolactin is produced by a microbial cell and/or a human cell expressing a recombinant prolactin (e.g., a prolactin comprising a substitution of a serine residue at position 179 of the prolactin gene with aspartate (S179D), e.g., S179D-prolactin). In some embodiments, adding prolactin to the culture medium comprises conditioning culture medium by culturing cells that express and secrete prolactin, and applying the conditioned culture medium comprising prolactin to the basal surface of the monolayer of mammary cells (e.g., mammary epithelial cells, mammary myoepithelial cells and mammary progenitor cells).

In some embodiments, the culture medium further comprises other factors to improve efficiency, including, but not limited to, insulin, an epidermal growth factor, and/or a hydrocortisone. In some embodiments, the methods of the present invention further comprise adding other factors (e.g., insulin, an epidermal growth factor, and/or a hydrocortisone) to the culture medium, e.g., to improve efficiency.

Methods of Manufacturing Cultured Milk Products

Disclosed herein are methods of manufacturing a cultured milk product representing the biosynthetic output of cultured mammary epithelial cells (MECs) and plasma cells.

In some embodiments, the method comprises culturing a cell construct comprising plasma cells and mammary cells disclosed herein in a bioreactor comprising a basal compartment and an apical compartment, wherein the basal compartment comprises a culture media, and the plasma cells and mammary cells secrete the cultured milk product comprising immunoglobulins into the apical compartment.

In some embodiments, the method comprises (a) isolating mammary epithelial cells, myoepithelial cells and/or mammary progenitor cells from mammary explants from mammary tissue (e.g., breast, udder, teat tissue), biopsy sample, or raw breastmilk, to produce isolated mammary epithelial cells, myoepithelial cells and/or mammary progenitor cells; (b) isolating plasma cells from mammary tissue, biopsy sample, or raw breastmilk, to produce isolated primary plasma cells; (c) culturing the isolated mammary epithelial cells, myoepithelial cells and/or mammary progenitor cells; (d) culturing the isolated primary plasma cells; (e) depositing the cultured, isolated mammary epithelial cells, myoepithelial cells and/or mammary progenitor cells and isolated primary plasma cells on a scaffold having an upper surface and lower surface to produce a mixed population of primary plasma cells and mammary epithelial cells, mammary myoepithelial cells and mammary progenitor cells wherein the plasma cells are overlaid by the mammary

42

epithelial cells, mammary myoepithelial cells and mammary progenitor cells; (f) cultivating the mixed population of (e) on the scaffold, to produce a monolayer of polarized mammary cells located adjacent to and above the plasma cells, wherein the plasma cells are located adjacent to and above the upper surface of the scaffold, wherein the upper surface is located adjacent to and above the lower surface of the scaffold, and wherein the polarized mammary cells comprise an apical surface and a basal surface, thereby producing a cell construct for producing the cultured milk product comprising immunoglobulins.

In some embodiments, the method comprises: a) isolating mammary epithelial cells, myoepithelial cells, and/or mammary progenitor cells from mammary explants from mammary tissue (e.g., breast, udder, teat tissue), biopsy sample, or raw breastmilk, to produce isolated mammary epithelial cells, myoepithelial cells, and/or mammary progenitor cells; (b) isolating plasma cells from mammary tissue, biopsy sample, or raw breastmilk, to produce isolated primary plasma cells; (c) culturing the isolated mammary epithelial cells, myoepithelial cells, and/or mammary progenitor cells to produce a mixed population of primary mammary epithelial cells, mammary myoepithelial cells and mammary progenitor cells; (d) culturing the isolated primary plasma cells; (e) sorting the mixed population of mammary epithelial cells, myoepithelial cells, and/or mammary progenitor cells (e.g., selecting the primary mammary epithelial cells) to produce a population of primary mammary epithelial cells; and (f) depositing the sorted population of primary mammary epithelial cells, myoepithelial cells and/or mammary progenitor cells and isolated primary plasma cells on a scaffold having an upper surface and lower surface (g) cultivating the sorted population of mammary epithelial and plasma cells on the scaffold, to produce a monolayer of polarized primary mammary epithelial cells located adjacent to and above the plasma cells, wherein the plasma cells are located adjacent to and above the upper surface of the scaffold, wherein the upper surface is located adjacent to and above the lower surface of the scaffold, and wherein the polarized mammary cells comprise an apical surface and a basal surface, and wherein the polarized monolayer comprises an apical surface and a basal surface, thereby producing a cell construct for producing the cultured milk product comprising immunoglobulins.

In some embodiments, the method comprises (a) culturing immortalized mammary epithelial cells to produce increased numbers of immortalized mammary epithelial cells; and (b) culturing plasma cells to produce increased numbers of plasma cells; (c) cultivating the immortalized mammary epithelial cells and plasma cells of (a) and (b) on a scaffold, the scaffold having an upper surface and lower surface, to produce a monolayer of polarized immortalized mammary epithelial cells above the plasma cells on the upper surface of the scaffold, wherein the polarized monolayer comprises an apical surface and a basal surface, thereby producing a cell construct for producing the cultured milk product comprising immunoglobulins.

In certain embodiments, plasma cells are added to the culture of immortalized mammary epithelial cells. In certain embodiments, the plasma cells are cultivated with the immortalized mammary epithelial cells on the scaffold, thereby producing a cultured milk product with secretory products of the immune cells and mammary cells (e.g., sIgA). In certain embodiments, the isolated mammary cells are immortalized prior to co-culture of the cells with the plasma cells.

43

Plasma cells can be derived from a primary mammary tissue sample or, e.g., from a plasma cell line. In certain embodiments, the plasma cells are stimulated to produce immunoglobulins during co-culture. In certain embodiments, the plasma cells produce IgA. In certain embodiments, the plasma cells produce secretory IgA. In certain embodiments, the one or more classes of immunoglobulins includes secretory IgA (sIgA). In certain embodiments, the immunoglobulins bind to an antigen of a microorganism (i.e., bacterium or virus). In certain embodiments, the immunoglobulins bind to viral or bacterial antigens capable of causing an infectious disease in humans. In certain embodiments, the immunoglobulins bind to viral or bacterial antigens that cause infections of respiratory or gastrointestinal epithelium. In certain embodiments, the immunoglobulins bind antigens from microorganisms that cause enterocolitis or sepsis in infants. In certain embodiments, plasma cells are co-cultured with MECs in a bioreactor according to methods described herein. In certain embodiments, the bioreactor is a hollow fiber bioreactor described herein.

In certain embodiments, mammary cells are modified and/or stimulated with prolactin according to the methods described herein to stimulate and optimize milk production. In certain embodiments, the mammary cells are modified to express a constitutively active prolactin receptor.

In certain embodiments, mammary epithelial cell populations are identified and isolated from primary mammary tissue samples. In some embodiments, the mammary cells are isolated and sorted via fluorescence-activated cell sorting, magnetic-activated cell sorting, and/or microfluidic cell sorting. In certain embodiments, myoepithelial mammary cells and luminal epithelial mammary cells are isolated by FACS analysis. In certain embodiments, progenitor myoepithelial mammary cells and/or progenitor luminal epithelial mammary cells are isolated by FACS analysis. Any suitable method known in the art for sorting mammary epithelial cells (e.g., luminal epithelial cells), myoepithelial cells, progenitor cells, and immune cells can be used. For example, mammary cells can be sorted using CD24, EPCAM and/or CD49f, cell surface markers.

In some embodiments, the plasma cells are isolated and sorted via fluorescence-activated cell sorting, magnetic-activated cell sorting, and/or microfluidic cell sorting. In certain embodiments, the plasma cells are sorted by FACS analysis using markers known in the art for identifying the plasma cells. In certain embodiment, plasma cells, plasma blasts or pre-plasmablasts are sorted and isolated by FACS analysis using markers known in the art (e.g., CD20, CD38, CD138, and/or CD19).

In some embodiments, the cell construct comprises a scaffold comprising an upper surface and a lower surface and a continuous monolayer of polarized mammary epithelial cells, a continuous monolayer of a polarized, mixed population of mammary epithelial cells, mammary myoepithelial cells and mammary progenitor cells, and/or a continuous monolayer of polarized immortalized mammary epithelial cells, wherein the continuous monolayer is located on the upper surface of scaffold.

In some embodiments, the lower surface of the scaffold is adjacent to the basal compartment. In some embodiments, the apical surface of the continuous monolayer is adjacent to the apical compartment. In some embodiments, the continuous monolayer secretes milk and sIgA or IgA through its apical surface into the apical compartment, thereby producing milk comprising IgA and/or sIgA in culture.

In some embodiments, the monolayer of mammary cells forms a barrier that divides the apical compartment and the

44

basal compartment, wherein the basal surface of the mammary cells are attached to the scaffold and the apical surface is oriented toward the apical compartment.

In some embodiments, the basal compartment is adjacent to the lower surface of the scaffold. In some embodiments, the basal compartment comprises a culture medium in fluidic contact with the basal surface of the monolayer of mammary epithelial cells (e.g., the polarized monolayer of mammary epithelial cells, the polarized the monolayer of the mixed population of mammary cells, or the polarized monolayer of immortalized mammary epithelial cells).

In some embodiments, the culture medium comprises a carbon source, a chemical buffering system, one or more essential amino acids, one or more vitamins and/or cofactors, and one or more inorganic salts.

In some embodiments, the bioreactor comprises an apical compartment that is adjacent to the apical surface of the monolayer. In some embodiments, the apical compartment is adjacent to the upper surface of the scaffold.

In some embodiments, the total cell density of mammary cells in the bioreactor is at least 10^{11} mammary cells. In some embodiments, the total cell density of mammary cells in the bioreactor is at least 10^{12} mammary cells. In some embodiments, the total cell density of mammary cells in the bioreactor is at least 10^{13} mammary cells.

In some embodiments, the total cell density of mammary cells in the bioreactor is about 20 to 55 cells per $100 \mu\text{m}^2$. In some embodiments, the total cell density of mammary cells in the bioreactor is about 20 cells per $100 \mu\text{m}^2$. In some embodiments, the total cell density of mammary cells in the bioreactor is about 25 cells per $100 \mu\text{m}^2$. In some embodiments, the total cell density of mammary cells in the bioreactor is about 30 cells per $100 \mu\text{m}^2$. In some embodiments, the total cell density of mammary cells in the bioreactor is about 35 cells per $100 \mu\text{m}^2$. In some embodiments, the total cell density of mammary cells in the bioreactor is about 40 cells per $100 \mu\text{m}^2$. In some embodiments, the total cell density of mammary cells in the bioreactor is about 45 cells per $100 \mu\text{m}^2$. In some embodiments, the total cell density of mammary cells in the bioreactor is about 50 cells per $100 \mu\text{m}^2$. In some embodiments, the total cell density of mammary cells in the bioreactor is about 55 cells per $100 \mu\text{m}^2$.

In some embodiments, the total cell density of plasma cells in the bioreactor is about 200 to 500 plasma cells per mm^2 . In some embodiments, the total cell density of plasma cells in the bioreactor is about 200 plasma cells per mm^2 . In some embodiments, the total cell density of plasma cells in the bioreactor is about 300 plasma cells per mm^2 . In some embodiments, the total cell density of plasma cells in the bioreactor is about 400 plasma cells per mm^2 . In some embodiments, the total cell density of plasma cells in the bioreactor is about 500 plasma cells per mm^2 .

In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 1.5 m^2 . In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 2 m^2 . In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 2.5 m^2 . In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 3 m^2 . In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 4 m^2 . In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 5 m^2 . In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 10 m^2 . In some embodiments, the total surface area of mammary cells

45

within the bioreactor is at least about 15 m². In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 20 m². In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 25 m². In some 5 embodiments, the total surface area of mammary cells within the bioreactor is at least about 50 m². In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 100 m². In some embodiments, the total surface area of mammary cells within the bioreactor is at least about 250 m². In some 10 embodiments, the total surface area of mammary cells within the bioreactor is at least about 500 m².

In some embodiments, the bioreactor maintains a temperature of about 27° C. to about 39° C. (e.g., a temperature of about 27° C., 28° C., 29° C., 30° C., 31° C., 32° C., 33° C., 34° C., 35° C., 35° C., 35.5° C., 36° C., 36.5° C., 37° C., 37.5° C., 38° C., 38.5° C. or about 39° C., or any value or range therein, e.g., about 27° C. to about 38° C., about 36° C. to about 39° C., about 36.5° C. to about 39° C., about 20 36.5° C. to about 37.5° C., or about 36.5° C. to about 38° C.). In some embodiments, the bioreactor maintains a temperature of about 37° C.

In some embodiments, the bioreactor has an atmospheric concentration of CO₂ of about 4% to about 6%, e.g., an 25 atmospheric concentration of CO₂ of about 4%, 4.25%, 4.5%, 4.75%, 5%, 5.25%, 5.5%, 5.75%, or 6% or any value or range therein, e.g., about 4% to about 5.5%, about 4.5% to about 6%, about 4.5% to about 5.5%, or about 5% to about 6%). In some embodiments, the bioreactor has an atmospheric concentration of CO₂ of about 5%.

In some embodiments, the bioreactor has an atmospheric concentration of CO₂ of about 4% to about 6%, e.g., an 30 atmospheric concentration of CO₂ of about 4%, 4.25%, 4.5%, 4.75%, 5%, 5.25%, 5.5%, 5.75%, or 6% or any value or range therein, e.g., about 4% to about 5.5%, about 4.5% to about 6%, about 4.5% to about 5.5%, or about 5% to about 6%). In some embodiments, the bioreactor has an atmospheric concentration of CO₂ of about 5%.

In some embodiments, the method comprises monitoring 40 the concentration of dissolved O₂ and CO₂. In some embodiments, the concentration of dissolved O₂ is maintained between about 10% to about 25% or any value or range therein (e.g., about 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25%). For example, in some embodiments, the concentration of dissolved O₂ is maintained between about 12% to about 25%, about 15% to about 22%, about 10% to about 20%, about 15%, about 20%, or about 22%. In some embodiments, the concentration of CO₂ is maintained 45 between about 4% to about 6%, e.g., a concentration of CO₂ of about 4%, 4.25%, 4.5%, 4.75%, 5%, 5.25%, 5.5%, 5.75%, or 6% or any value or range therein, e.g., about 4% to about 5.5%, about 4.5% to about 6%, about 4.5% to about 5.5%, or about 5% to about 6%). In some embodiments, the concentration of CO₂ is maintained at about 5%.

In some embodiments, the culture medium is exchanged about every day to about every 10 days (e.g., every 1 day, every 2 days, every 3 days, every 4 days, every 5 days, every 6 days, every 7 days, every 8 days, every 9 days, every 10 50 days, or any value or range therein, e.g., about every day to every 3 days, about every 3 days to every 10 days, about every 2 days to every 5 days). In some embodiments, the culture medium is exchanged about every day to about every few hours to about every 10 days, e.g., about every 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, or 24 hours to about every 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 days or any value or range therein. For example, in some 65

46

embodiments, the culture medium is exchanged about every 12 hours to about every 10 days, about every 10 hours to about every 5 days, or about every 5 hours to about every 3 days.

In some embodiments, the method comprises monitoring 5 the glucose concentration and/or rate of glucose consumption in the culture medium and/or in the lactogenic culture medium. In some embodiments, the prolactin is added when the rate of glucose consumption in the culture medium is steady state.

In some embodiments, the method further comprises 10 applying transepithelial electrical resistance (TEER) to measure the maintenance of the monolayer of epithelial cells. TEER measures a voltage difference between the fluids (e.g., media) in two compartments (e.g., between the apical and basal compartments), wherein if the barrier between the compartments loses integrity, the fluids in the two compartments may mix. When there is fluid mixing, the voltage difference will be reduced or eliminated; a voltage difference 15 indicates that the barrier is intact. In some embodiments, upon detection of a loss of voltage by TEER, a scaffold (e.g., a Transwell® filter, a microstructured bioreactor, a decellularized tissue, a hollow fiber bioreactor, etc.) is reinoculated with additional cells and allowed time to reestablish a barrier (e.g., a monolayer) before resuming production of the cultured milk product (e.g., milk production).

In some embodiments, the method further comprises 20 collecting the cultured milk product from the apical compartment to produce collected cultured milk product. In some embodiments, the collecting is via a port, via gravity, and/or via a vacuum. In some embodiments, a vacuum is attached to a port.

In some embodiments, the method further comprises 25 freezing the collected cultured milk product to produce frozen cultured milk product and/or lyophilizing the collected cultured milk product to produce lyophilized cultured milk product.

In some embodiments, the method further comprises 30 packaging the collected cultured milk product, the frozen cultured milk product and/or the lyophilized cultured milk product into a container.

In some embodiments, the method further comprises 35 extracting one or more components from the collected cultured milk product. Non-limiting examples of components from the collected cultured milk product include milk protein, lipid, carbohydrate, vitamin, and/or mineral contents. In some embodiments, the components from the collected cultured milk product are lyophilized and/or concentrated to produce a lyophilized or a concentrated cultured milk product component product. In some embodiments, the components from the collected cultured milk product are concentrated by, e.g., membrane filtration and/or reverse osmosis. In some embodiments, the lyophilized or concentrated cultured milk product component product is packaged 40 in a container, optionally wherein the container is sterile and/or a food grade container. In some embodiments, the container is vacuum-sealed. In some embodiments, the container is a canister, ajar, a bottle, a bag, a box, or a pouch. In some embodiments, the cultured milk product is a standardized, sterile cultured milk product. In some embodiments, the cultured milk product is for nutritional use.

In some embodiments, the cultured milk product is produced by any method disclosed herein.

Breast milk contains low but measurable concentrations 45 of environmental contaminants, health-harming chemicals from industry and manufacturing products that are widely spread in the environment. Environmental contaminants are

47

partly secreted in breast milk. The contaminant levels in breast milk reflect those in the mother's body and are therefore ideal for monitoring exposure levels. Toxic environmental contaminants can be transferred from mother to infant via breastfeeding. Persistent organic pollutants (POPs) are a family of lipophilic stable chemicals that bioaccumulate in adipose tissue and create a lasting toxic body burden. Breastfeeding provides a significant source of exposure to POPs early in human life, the effects of which are unknown.

In some embodiments, the cultured milk product does not comprise or is substantially free of one or more environmental contaminants. In some embodiments, the cultured milk product does not comprise or is substantially free of persistent organic pollutants (POPs). In some embodiments, the cultured milk product does not comprise or is substantially free of polychlorinated dibenzo-p-dioxins (PCDDs), polychlorinated dibenzofurans (PCDFs), polychlorinated biphenyls (PCBs) and pesticides such as DDT.

Heavy metals such as mercury, lead, arsenic, cadmium, nickel, chromium, cobalt, zinc, and other potentially toxic metals that are dispersed throughout the environment also have bioaccumulative features known to accumulate in human milk and thus are of concern to the nursing infant. Metal in breast milk originates from exogenous sources, i.e., uptake via contaminated air, food, and drinking water, and endogenous release along with essential trace elements. For example, lead and mercury are equally dispersed in the human food chain, and their impact on fetal development is heavily determined by the mother's diet and nutritional status. The exposures to toxic metals have significant public health implication, even at small concentrations and acute exposures, these metals remain toxic to humans. A nursing infant may be exposed to toxic metals in a period of highest susceptibility. Nursing infants may be exposed to heavy metals through breast milk in excess of what they should, and exposure may have health implication for the infants. For infants in particular, these exposures may have adverse effect on the developing central nervous system, leaving a life-long defect on their cognitive abilities.

In some embodiments, the cultured milk product does not comprise or is substantially free of one or more heavy metals, such as arsenic, lead, cadmium, nickel, mercury, chromium, cobalt, and zinc. In some embodiments, the cultured milk product does not comprise or is substantially free of arsenic. In some embodiments, the cultured milk product does not comprise or is substantially free of lead. In some embodiments, the cultured milk product does not comprise or is substantially free of cadmium. In some embodiments, the cultured milk product does not comprise or is substantially free of nickel. In some embodiments, the cultured milk product does not comprise or is substantially free of mercury. In some embodiments, the cultured milk product does not comprise or is substantially free of chromium. In some embodiments, the cultured milk product does not comprise or is substantially free of cobalt. In some embodiments, the cultured milk product does not comprise or is substantially free of zinc. In some embodiments, the cultured milk product does not comprise or is substantially free of arsenic, lead, cadmium, nickel, mercury, chromium, cobalt, and zinc.

Foreign allergenic proteins can be difficult to distinguish from endogenous human milk proteins. Food proteins with allergenic potential that have been detected in human milk include hen's egg and peanut proteins. There are eight major food allergens, known as the big 8, that are responsible for most of the serious food allergy reactions in the U.S. The big

48

8 list is comprised of milk, egg, fish, crustacean shellfish, tree nuts, peanuts, wheat, and soybean allergens. Proteins known to cause egg allergy include ovomucoid, ovalbumin, and conalbumin. Peanuts proteins include arachin 6, arachin 3, conarachin, main allergen Arah1, and arachin Arah2. As an example of maternal dietary protein transportation to milk, it has been shown that the consumption of one egg per day leads to higher concentrations of the chicken egg allergen ovalbumin (OVA) in human milk compared to egg-avoiding mothers.

In some embodiments, the cultured milk product does not comprise or is substantially free of one or more food allergens. In some embodiments, the cultured milk product does not comprise or is substantially free of egg, fish, crustacean shellfish, tree nuts, peanuts, wheat, and soybean allergens. In some embodiments, the cultured milk product does not comprise or is substantially free of egg allergens. In some embodiments, the cultured milk product does not comprise or is substantially free of fish allergens. In some embodiments, the cultured milk product does not comprise or is substantially free of crustacean allergens. In some embodiments, the cultured milk product does not comprise or is substantially free of tree nut allergens. In some embodiments, the cultured milk product does not comprise or is substantially free of peanut allergens. In some embodiments, the cultured milk product does not comprise or is substantially free of wheat allergens. In some embodiments, the cultured milk product does not comprise or is substantially free of soybean allergens.

In some embodiments, the cultured milk product does not comprise or is substantially free of arachin 6, arachin 3, conarachin, Arah1, and Arah2.

In some embodiments, the cultured milk product does not comprise or is substantially free of ovalbumin (OVA).

Methods of Use

Disclosed herein, in certain embodiments, are methods of treating and/or preventing infection comprising administration of a cultured milk product comprising IgA or sIgA described herein, or and immunotherapeutic composition comprising isolated IgA or sIgA described herein.

In certain embodiments, the immunotherapeutic compositions and cultured milk products comprising IgA and/or sIgA are administered to patients in an effective amount for the treatment or prevention of a microbial infection. In certain embodiments, the microbial infection is a bacterial infection. Non-limiting examples of bacterial infections that can be treated and/or prevented include infections caused by: *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Moraxella catarrhalis*, *Staphylococcus aureus*, *Streptococcus pyogenes*, *Salmonella*, *Shigella*, *Campylobacter*, *Staphylococcus aureus* and *Helicobacter pylori*. In certain embodiments, the microbial infection is a viral infection. Non-limiting examples of viral infection that can be treated and/or prevented include infections caused by: influenza virus, parainfluenza virus, respiratory syncytial virus, rhinovirus, and human metapneumovirus.

In certain embodiments, the cultured milk product immune comprising IgA or sIgA, or therapeutic composition comprising IgA or sIgA are administered orally to a subject to treat or prevent a gastrointestinal infection. In certain embodiments, the gastrointestinal infection is bacterial or viral gastroenteritis. In certain embodiments, the subject has an ulcer caused by an infection of *Helicobacter pylori*. In certain embodiments, the subject has an infection caused by norovirus, adenovirus, or rotavirus. In certain embodiments, the immunotherapeutic compositions and/or cultured milk

products are administered to an infant for the treatment or prevention of gastrointestinal infection.

In certain embodiments, the immunotherapeutic compositions disclosed herein are administered to a subject as a nasal inhalant to treat or prevent a respiratory infection. In certain embodiments, the subject has pneumonia, bronchitis, and/or lung tissue damage.

In certain embodiments, the subject is immune compromised. The subject can be immune compromised due to a primary immune deficiency disease or disorder (e.g., severe combined immunodeficiency (SCID) or a secondary immune deficiency, such as, but not limited to, chemotherapy treatment, diabetes, aging, malnutrition, and acquired immunodeficiency syndrome. In certain embodiments, the immunotherapeutic compositions and/or cultured milk products are administered to an infant for the treatment or prevention of an infection.

In certain embodiments, the immunotherapeutic compositions and/or cultured milk products are administered in combination with another anti-microbial agent (i.e., antibiotic or anti-viral agent). In certain embodiments, the cultured milk product described herein are administered with an additional nutritional product for treatment and/or prevention of a gastrointestinal infection.

Having described the present disclosure, the same will be explained in greater detail in the following examples, which are included herein for illustration purposes only, and which are not intended to be limiting to the disclosure.

EXAMPLES

The Examples that follow are illustrative of specific embodiments of the disclosure, and various uses thereof. They are set forth for explanatory purposes only and should not be construed as limiting the scope of the disclosure in any way.

Example 1: Production of Milk Components from Human Mammary Epithelium

In this example, human mammary epithelium is recapitulated and milk is produced in vitro. The system and process described is exemplary and can be scaled to produce multiliter volumes of milk components. Mammary epithelial cells are expected to form a polarized monolayer on bioreactor fibers that have been precoated with one or more of laminin and collagen or other extracellular matrix proteins, as well as on uncoated fibers. When confluent, the monolayer forms a barrier that divides the intra- and extracapillary space (ECS), with the basal surface attached to the fibers and the apical surface oriented toward the ECS. Milk component production is stimulated by addition of prolactin to the media. The secreted milk components are collected from the ECS and submitted for downstream analyses of the protein, lipid, and carbohydrate content in comparison to human breast milk produced in vivo.

Materials for use in this Example are shown in Table 3.

Materials		
Item	Supplier	Catalog Number
Primary mammary epithelial cells	ATCC	PCS-600-010
Mammary epithelial cell basal medium	ATCC	PCS-600-30
Mammary epithelial cell growth kit	ATCC	PCS-600-040

-continued

Materials		
Item	Supplier	Catalog Number
Dulbecco's phosphate buffered saline (D-PBS)	ATCC	ATCC 30-2200
Trypsin-EDTA	ATCC	PCS-999-003
Trypsin Neutralizing Solution	ATCC	PCS-999-004
EHS (Laminin-1/111)	Sigma	L2020-1 MG
Collagen-IV	Sigma	C5533-5 MG
Prolactin	Shenandoah Biotechnology	100-45-500 ug

Procedures

Expansion of Primary Human Mammary Epithelia Cells (HMECs)

Human mammary epithelial cells (1 ampoule; 5×10⁵ cells) are expanded into one collagen-IV-coated T300 flask (or two T175 flasks) in mammary epithelial cell medium prepared with supplements provided in the kit listed in Table 3. Once an appropriate cell number is obtained, cells are rinsed with D-PBS and collected from the plates using trypsin-EDTA. Once cells are detached, trypsin activity is halted using Trypsin Neutralizing Solution. Cells are resuspended in medium and seeded into a hollow fiber bioreactor (Fibercell Systems), prepared as described below.

Preparation of Hollow Fiber Bioreactor (C2025D, 20 kD MWCO)

Prior to seeding, a bioreactor cartridge (Fibercell Systems) is prepared by preculturing with PBS for a minimum of 24 hours. The bioreactor cartridge is optionally pre-coated by adding about 50-100 pg of one or more of collagen I, collagen IV, laminin-1 11 (e.g., laminin-111 isolated from Engelbreth-Holm Swarm tumor), alpha-4, alpha-5, fibronectin, and/or entactin in 3.2 mL of PBS and allowing ultrafiltration across the fiber at room temperature overnight. The uncoated or precoated cartridge is exchanged with medium and incubated overnight at room temperature. The medium is then exchanged with the cells collected from the T300 (or T175) flask(s). The reservoir volume is no more than 125 mL. The cartridge is rotated 180 degrees after seeding the cells.

Cell Growth in the Bioreactor and Prolactin Stimulation

After seeding the bioreactor, cells are grown in mammary epithelial cell growth medium supplemented with 10-25 mL DMEM/10% CDM-HD (Dulbecco's Modified Eagle Medium; Chemically Defined Medium for High Density Cell Culture) per 100 mL medium. The proportion of DMEM/CDM-HD can be adjusted based on the rate of glucose consumption.

Before stimulation of milk secretion, the medium in the ECS is flushed and replaced with PBS. To stimulate milk component secretion, medium supplemented with 100 ng/mL prolactin is added. The lactogenic medium can also be supplemented with an elevated concentration of glucose and the essential dietary precursors for milk fatty acids, linoleic acid and a-linolenic acid. The bioreactor is maintained for 10 days with sampling as described below.

Harvesting and Sample Preparation

Samples, comprised of supernatant from the ECS and an equivalent volume of media from the reservoir, are collected once daily for 10 days after addition of prolactin to the media. The samples are spun in a centrifuge to collect any debris and resuspended in an equivalent volume of PBS. The supernatants from the ECS and media samples are divided into 0.5 mL aliquots in microfuge tubes and frozen at -80°

51

C. The pellet debris is resuspended in a volume of PBS equivalent to the original sample and frozen at -80°C . Samples are processed to determine relative concentrations of milk components produced.

Scaling for Multiliter Production

To scale for multiliter production, the preceding procedure with relative adjustments in reagent volumes for a larger bioreactor (e.g., Fibercell Systems cat. no. C2018) is performed.

Example 2

A cell culture system designed for the collection of milk should support compartmentalized secretion of the product such that the milk is not exposed to the media that provides nutrients to the cells. In the body, milk-producing epithelial cells line the interior surface of the mammary gland as a continuous monolayer. The monolayer is oriented such that the basal surface is attached to an underlying basement membrane, while milk is secreted from the apical surface and stored in the luminal compartment of the gland, or alveolus, until it is removed during milking or feeding. Tight junctions along the lateral surfaces of the cells ensure a barrier between the underlying tissues and the milk located in the alveolar compartment. Therefore, in vivo, the tissue of the mammary gland is arranged such that milk secretion is compartmentalized, with the mammary epithelial cells themselves establishing the interface and maintaining the directional absorption of nutrients and secretion of milk.

The present disclosure describes a cell culture apparatus that recapitulates the compartmentalizing capability of the mammary gland that is used to collect milk from mammary epithelial cells grown outside of the body. Such an apparatus can include a scaffold to support the proliferation of mammary cells at the interface between two compartments, such that the epithelial monolayer provides a physical boundary between the nutrient medium and the secreted milk. In addition to providing a surface for growth, the scaffold provides spatial cues that guide the polarization of the cells and ensures the directionality of absorption and secretion. This invention describes the preparation, cultivation, and stimulation of mammary epithelial cells in a compartmentalizing cell culture apparatus for the production and collection of milk for nutritional use (see e.g., FIG. 1).

Preparation of mammary epithelial cells. Mammary epithelial cells are obtained from surgical explants of dissected mammary tissue (e.g., breast, udder, teat), biopsy sample, or raw breastmilk. Generally, after surgical dissection of the mammary tissue, any fatty or stromal tissue is manually removed under aseptic conditions, and the remaining tissue of the mammary gland is enzymatically digested with collagenase and/or hyaluronidase prepared in a chemically defined nutrient media, which should be composed of ingredients that are “generally recognized as safe” (GRAS). The sample is maintained at 37°C . with gentle agitation. After digestion, a suspension of single cells or organoids is collected, either by centrifugation or by pouring the sample through a sterile nylon cell strainer. The cell suspension is then transferred to a tissue culture plate coated with appropriate extracellular matrix components (e.g., collagen, laminin, fibronectin).

Alternatively, explant specimens can be processed into small pieces, for example by mincing with a sterile scalpel. The tissue pieces are plated onto a suitable surface such as a gelatin sponge or a plastic tissue culture plate coated with appropriate extracellular matrix.

52

The plated cells are maintained at 37°C . in a humidified incubator with an atmosphere of 5% CO_2 . During incubation, the media is exchanged about every 1 to 3 days and the cells are sub-cultured until a sufficient viable cell number is achieved for subsequent processing, which includes preparation for storage in liquid nitrogen; development of immortalized cell lines through the stable transfection of genes such as SV40, TERT, or other genes associated with senescence; isolation of mammary epithelial, myoepithelial, and stem/progenitor cell types by, for example, fluorescence-activated cell sorting; and/or introduction into a compartmentalizing tissue culture apparatus for the production and collection of milk for human consumption.

Cultivation of mammary epithelial cells for the production of milk. Milk for nutritional use is produced by mammary epithelial cells isolated as described above and cultured in a format that supports compartmentalized secretion such that separation between the nutrient medium and the product is maintained. The system relies on the ability of mammary epithelial cells to establish a continuous monolayer with appropriate apical-basal polarity when seeded onto an appropriate scaffold positioned at the interface between the apical compartment, into which milk is secreted, and the basal compartment, through which nutrient media is provided (see, e.g., FIG. 2). Transwell® filters placed in tissue culture plates, as well as bioreactors based on hollow fiber or microstructured scaffolds, for example, are used to support these characteristics.

Following the isolation and expansion of mammary epithelial cells, the cells are suspended in a chemically defined nutrient medium composed of food-grade components and inoculated into a culture apparatus that has been pre-coated with a mixture of extracellular matrix proteins, such as collagen, laminin, and/or fibronectin. The cell culture apparatus is any design that allows for the compartmentalized absorption of nutrients and secretion of product from a polarized, confluent, epithelial monolayer. Examples include hollow fiber and microstructured scaffold bioreactors (see, e.g., FIGS. 3 and 4, respectively). Alternatives include other methods of 3-dimensional tissue culture, such as the preparation of decellularized mammary gland as a scaffold, repopulated with stem cells to produce a functional organ in vitro, or collection of milk from the lumen of mammary epithelial cell organoids or “mammospheres” grown either in a hydrogel matrix or in suspension.

The apparatus includes sealed housing that maintains a temperature of about 37°C . in a humidified atmosphere of about 5% CO_2 . Glucose uptake is monitored to evaluate the growth of the culture as the cells proliferate within the bioreactor. Stabilization of glucose consumption indicates that the cells have reached a confluent, contact-inhibited state. The integrity of the monolayer is ensured using transepithelial electrical resistance. Sensors monitor concentrations of dissolved O_2 and CO_2 in the media at multiple locations. A computerized pump circulates media through the bioreactor at a rate that balances the delivery of nutrients with the removal of metabolic waste such as ammonia and lactate. Media can be recycled through the system after removal of waste using Lactate Supplementation and Adaptation technology (Freund et al. 2018 *Int J Mol Sci.* 19(2)) or by passing through a chamber of packed zeolite.

Stimulation of milk production. In vivo and in cultured mammary epithelial cells, the production and secretion of milk is stimulated by prolactin. In culture, prolactin can be supplied exogenously in the nutrient media at concentrations approximating those observed in the body during lactation, e.g., about 20 ng/mL to about 200 ng/mL. Purified prolactin

53

can be obtained commercially; however, alternative methods of providing prolactin or stimulating lactation are employed, including expression and purification of the recombinant protein from microbial or mammalian cell cultures. Alternatively, conditioned media prepared by culturing cells that express and secrete prolactin can be applied to mammary epithelial cell cultures to stimulate lactation. Bioreactors can be set up in series such that media passing through a culture of cells expressing prolactin or other key media supplements is conditioned prior to exposure to mammary cells grown in a compartmentalizing culture apparatus as described.

Other approaches to upregulate milk production and/or spare the use of exogenous prolactin include molecular manipulation of the signaling pathways that are regulated by binding of prolactin to its receptor on the surface of mammary epithelial cells, such as the following: (a) expression of constructs targeting the posttranslational modification of prolactin; (b) expression of alternative isotypes of the prolactin receptor; (c) expression of a chimeric prolactin receptor in which the extracellular domain is exchanged with the binding site for a different ligand; (d) introduction of a gene encoding a constitutively or conditionally active prolactin receptor or modified versions of its downstream effectors such as STAT5 or Akt; (e) knockout or modification of the PER2 circadian gene; and/or (f) molecular approaches aimed at increasing the rate of nutrient uptake at the basal surface of the mammary epithelial monolayer.

Collection of milk. Secreted milk is collected continuously or at intervals through, for example, a port installed in the apical compartment of the culture apparatus. A vacuum is applied to the port to facilitate collection and also contributes to the stimulation of further production. The collected milk is packaged into sterile containers and sealed for distribution, frozen or lyophilized for storage, or processed for the extraction of specific components.

The present invention provides mammary epithelial cell cultures for the production of milk for nutritional use. In addition to human breast milk, this method may be used to produce milk from other mammalian species, for example, for human consumption or veterinary use. Because it has not been previously possible to produce milk outside the body, this technology may result in novel commercial opportunities, in addition to providing an alternative mode of production for existing products. The social and economic effects of the commercial development of this technology are broad and far reaching. Production of human breast milk from cultured cells may provide a means to address infant malnutrition in food-scarce communities, provide essential nutrients to premature infants who are unable to breastfeed, and offer mothers a new option for feeding their babies that provides optimal nutrition with the convenience of infant formula. Production of cow or goat milk provides an opportunity to reduce the environmental, social, and animal welfare effects of animal agriculture. The process described here addresses an important gap in the emerging field of cellular agriculture and introduces an opportunity to dramatically update the human food supply without compromising our biological and cultural attachment to the most fundamental of our nutrition sources.

Example 3: Manufacturing Milk Compositions Comprising Immunoglobulins

Primary mammary tissue is collected from a healthy donor. CD20+ immune cells and mammary epithelial cell populations are sorted by FACS analysis using known

54

markers for the myoepithelial and luminal epithelial cell populations. The isolated mammary cells and immune cells are immortalized prior to co-culture of the cells. The immortalized immune cells are co-cultured with immortalized isolated mammary epithelial cells in a hollow fiber bioreactor and grown to confluence. The culture is stimulated with prolactin to stimulate milk production. The secretory cell culture product comprising immunoglobulins, including secretory IgA is then harvested by a non-disruptive withdrawal mechanism.

Example 4: Manufacturing of Immunotherapy Compositions from Co-Cultured Mucosal Epithelial Cells and Immune Cells

Primary mammary epithelial cells and CD20+ immune cells are harvested from tissue derived from patients who have recovered from a mucosal infection. Patients are selected that have been tested positive for serum that exhibits viral neutralization activity. The CD20+ immune cells are co-cultured with the mammary epithelial cells in a hollow fiber bioreactor. The immune cells are grown in 2-D culture, followed by inoculation into a hollow fiber bioreactor and grown to maximum density (i.e., superconfluence) within the bioreactor. The co-culture of the mucosal epithelial cells and immune cells are stimulated to produce immunoglobins including secretory IgA. The co-culture is stimulated with one or more of a stimulant selected from, a viral protein, a viral protein fragment, prolactin, TGF- β , and interleukin 10 (IL-10). The products from the co-culture of mucosal epithelial cells and stimulated immune cells are harvested using a standard vacuum method. In certain embodiments, the secreted product is purified. The immunoglobulins are purified from the harvested secreted product. The purified secreted product is formulated as an inhalant for human pulmonary administration and is effective in the treatment of respiratory infection.

Example 5: Treatment Of Respiratory Infection With Immunotherapy Composition From Co-Cultured Mucosal Epithelial Cells And Immune Cells

An immunotherapy composition produced as described above in Example 3 is formulated as an inhalant and is administered to a patient harboring a respiratory infection, in an amount effective to treat the infection. The immunotherapy composition is effective at reducing the severity of symptoms of the infection in the patient.

The foregoing examples are illustrative of the present disclosure and are not to be construed as limiting thereof. Although the disclosure has been described in detail with reference to preferred embodiments, variations and modifications exist within the scope and spirit of the disclosure as described and defined in the following claims.

INFORMAL SEQUENCE LISTING

SEQ ID NO: 1: Human Prolactin Receptor (GenBank
Accession Number AAD32032.1)
MKENVASATVFTLLFLNTCLLNGQLPPGK-
PEIFKCRSPNKETFTCWWRPGTDGGLPTNYSL
TYHREGETLMHECPDYITGGPN-
SCHFGKQYTSMWRTYIMMVNATNQMGSSFSDELY-
VDVT YIVQPDPPLELAVEVKQPE-
DRKPYLWIKWSPPTLIDLKTGWFTLLYEIRLKPEKA
AEWEIHFA

55

GQQTEFKILSLHPGQKYLQVRCKPDHGYW-
SAWSPATFIQIPSDFTMNDTTVWISVAVLSA VICLIIV-
WAVALKGYSMVTCIFPPVPGPKIKGFD AHLLEK GK-
SEELLSALGCQDFPPTS DYED
LLVEYLEVDD SEDQHLMSVHSKEHPSQGMKP-
TYLDPD TDSGRGSCDSPSL LSEKCEEPQAN
PSTFYDPEVIEK PENPETHTHTWDPQCISMEGKIPYF-
HAGGSKCSTWPLPQPSQHNP RSSYHNI
TDVCELAVGPAGAPATLLNEAGKDALKSSQTIKS-
REEGKATQQREVESFHSETDQDTPWLL PQEKT PFG-
SAKPLDYVEIHKVNKD-
GALLSLPKQRENSGKPKKPGTPENNKEYAKVSGVMD
NNILVLVPDPHAKNVACFEESAKE-
APPSLEQNQA EKALANFTATSSK CRLQLG-
GLDYLDPA CFTHSFH
SEQ ID NO: 2: Human Period Circadian Protein Homolog
2 (GenBank Accession Number NM 022817)
MNGYAEFPPSPSNPTKEPVEPQP-
SQVPLQEDVDMSSGSSGHETNENCST-
GRDSQGSDCDDS GKJELGMLVEPPDAR-
QSPDTFSLMMAKSEHNPSTSG
CSSDQSSKVDTHKELIKTLKELKVH LPADK-
KAKGKASTLATLKYALRSVKQVKANEEYYQLL-
MSSEGHPCGADVPSYTV EEMESV TSE-
HIVKNADMF AVAVSLVSGKILYISDQVASIFHCKRD
AFSDAKFVEFLAPHDVG VFHSFT
SPYKLPLWSMCSGADSFTQECMEEKSF FCRVSVRK-
SHENEIRYHPFRMTPYLVKVRDQQGA ESQ LCCLL-
LAERVHSGYEAPRIPPEKRIFTTHTPNCLFQD VDER-
AVPLLGYLPQDLIETPVL
VQLHPSDRPLMLAIHKKILQSGGQPF DYSPIRFRA-
RNGEYITLDTSWSSFINPWSRKISFI GRH KVRVG-
PLNEDVFAAHPCTEEKALHPSIQELTE-
QIHRLLLQPVPHSGSSGYGSLG SNGSHEHL
MSQTSSSDSNGHEDSRRRRAEICKNGNKTKNRSHY-
SHESGEQKKKSVTEMQTNPPAEKKA VPAMEKD-
SLGVSPPEELACKNQPTCSYQQISCLDS-
VIRYLESCNEAATLKRKCEFPANVPAL
RSSDKRKATVSPGPHAGEAEPPSRVNSRTGVGTHLT-
SLALPGKAESVASLTSQCSYSSTIVH
VGDKKPQPELEMVEDAASGPESLD-
CLAGPALACGLS QEKEPFKKLGLTKEVLAAHTQ-
KEEQ SFLQKFKEIRKLSIFQSHCHYYLQERSKGQP-
SERTAPGLRNTSGIDSPWKKTGKNRKLKSKR
VKPRDSSESTGSGGPVSARPPLVGLNA-
TAWSPSDTSQSSCPAVPFPAPVPAAYSLPVFPAPGT
VAAPPAP-
PHASFTVPAVPVDLQH QFAVQPPPPFAPLAPVMA-
FMLPSYSFPGSTPNLPQAFFPS QPQFPSHPTLTSE-
MASASQPEFPEGGTGAMGTTGATETA AVGADCKPG
TSRDQQPKAPLTR DEPSDTQNSDALST-
SSGLLNLLL NEDLCSASGSAASESLGSGSLGC-
DASPSGAGSSDTSHTSK YFGSIDSEN NHKAKMNTG-
MEESEHFIKCVLQDPIWLLMADADSSVMMTYQLP
SRNLEAV LKEDREKLKLLQKLQPRFTESQKQEL-

56

REVHQWMQTGGLPAAIDVAECVYCENKEKGNICIP
YEEDIPSLGLSEVSDTKEDENG SPLNHRIEEQT
SEQ ID NO: 3: Human Isoform 4 of Prolactin Receptor
(GenBank Accession Number AF416619; Trott et al. 2003 *J.*
Mol. Endocrinol 30(1):31-47)
5 MKENVASATVFTLLLFLNTCLLNGQLPPGK-
PEIFKCRSPNKETFTCWWRPGTDGGLPTNYSL
TYHREGETLMHECPDYITGGPN-
SCHFGKQYTSMWRTYIMMVNATNQMGSSFSDELY-
VDVT YIVQPDPPLELAVEVKQPE-
10 DRKPYLWIKWSPPTLIDLKTGWFTLLYEIRLKPEK
AAEWEIHFA GQQTEFKILSLHPGQKYLQVRCK-
PDHGYWSAWSPATFIQIPSDFTMNDTTVWISVAVLSA
VICLIIVWAVALKGYSMVTCIFPPVPGPKIKGFD AHL-
LEK GKSEELLSALGCQDFPPTS DYED LLVEYLEVDD-
15 SEDQHLMSVHSKEHPSQGDPLMLGASHYKNLKS YR-
PRKISSQGR LAVFTKA TLTTVQ
SEQ ID NO: 4. STA5A Human Signal Transducer and
Activator of Transcription 5A Fused at 3' End to Amino
Acids 757-1129 of JAK2 Human Tyrosine-Protein Kinase
20 MAGWIAQQL QGDALRQMQV LYGQHFP IEV
RHYLAQWIES QPWDAIDL DN PQDRAQATQL
LEGLVQELQK KAEHQVGEDG FLLKIKLGHY
ATQLQKTYDR CPLELVR CIR HILYNEQRLV
REANNCSSPA GILVDAMSQK HLQINQTFEE
25 LRLVTQD TEN ELKKLQQTQE YFIIQYQESL
RIQAQFAQLA QLSPQERLSR ETALQQKQVS
LEAWLQREAQ TLQQYRVELA EKHQKTLQLL
RKQQTILDD ELIQWKRRQQ LAGNGGPPEG
SLDVLQSWCE KLA EIWQNR QQIRRAEHL C
30 QQLPIPGPVE EMLAEV NATI TDIISALVTS
TFIIEKQPPQ VLKTQTKFAA TVRLLVGGKL
NVHMNPPQVK ATIISEQQAK SLLKNENTRN
ECSGEILNNC CVMEYHQATG TLSAHFRNMS
LKRIKRADRR GAESVTEEFK TVLFESQFSV
35 GSNELVFQVK TSLPWWIV HGSQDHNATA TVLWD-
NAFAE PGRVPFAVPD KVLWPQLCEA LNMKFKA EVQ
SNRGLTKENL VFLAQKLFNN SSSHLEDYSG
LSVSWSQFNR ENLPGWNYTF WQWFDGVMEV
LKKHHKPHWN DGAILGFVNK QQAHDLLINK PDGT-
40 FLLRFS DSEIGGITIA WKFDSPERNL WNLKPFTTRD
FSIRSLADRL GDLSYLIYVF PDRPKDEVFS KYYTPV-
LAKA VDG YVKPQIK QWPEFVNAS ADAGGSSATY
MDQAPSPAVC PQAPYNMYPQ NPDHVLDQDG EFDL-
DETMDV ARHVEELLRR PMDSLDSRLS PPAGLFT-
45 SAR GSLSLDSQ RKLQFYEDRH QLPAPKWAEL
ANLINNCMDY EPDFRPSFRA IIRDLNSLFT PDYELL-
TEND MLPNM RIGAL GFSGAFEDRD PTQFEERHLK
FLQQLGKGNF GSVEMCRYDP LQDNTGEWA
VKKLQHSTEE HLRDFEREIE ILKSLQHDNI
50 VKYKGV CYSA GRRNLKLIME YLPYGSLRDY LQKH-
KERIDH IKLLQYTSQI CKGMEYLGTK RYIHRDLATR
NILVENENRV KIGDFGLTKV LPQDKEYYKV
KEPGESPIFW YAPESL TESK FSVASDVWSF GWLY-
ELFTY IEKSKSPPAE FMRMIGNDKQ GQMIVFHLIE
55 LLKNNGR LPR PDGCPDEIYM IMTECWNNNV
NQRPSFRDLA LRVDQIRDN.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 4

<210> SEQ ID NO 1
<211> LENGTH: 622
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens

-continued

<400> SEQUENCE: 1																
Met	Lys	Glu	Asn	Val	Ala	Ser	Ala	Thr	Val	Phe	Thr	Leu	Leu	Leu	Phe	
1				5					10					15		
Leu	Asn	Thr	Cys	Leu	Leu	Asn	Gly	Gln	Leu	Pro	Pro	Gly	Lys	Pro	Glu	
			20					25					30			
Ile	Phe	Lys	Cys	Arg	Ser	Pro	Asn	Lys	Glu	Thr	Phe	Thr	Cys	Trp	Trp	
		35					40					45				
Arg	Pro	Gly	Thr	Asp	Gly	Gly	Leu	Pro	Thr	Asn	Tyr	Ser	Leu	Thr	Tyr	
	50					55					60					
His	Arg	Glu	Gly	Glu	Thr	Leu	Met	His	Glu	Cys	Pro	Asp	Tyr	Ile	Thr	
65					70					75					80	
Gly	Gly	Pro	Asn	Ser	Cys	His	Phe	Gly	Lys	Gln	Tyr	Thr	Ser	Met	Trp	
				85					90					95		
Arg	Thr	Tyr	Ile	Met	Met	Val	Asn	Ala	Thr	Asn	Gln	Met	Gly	Ser	Ser	
			100					105					110			
Phe	Ser	Asp	Glu	Leu	Tyr	Val	Asp	Val	Thr	Tyr	Ile	Val	Gln	Pro	Asp	
		115					120					125				
Pro	Pro	Leu	Glu	Leu	Ala	Val	Glu	Val	Lys	Gln	Pro	Glu	Asp	Arg	Lys	
		130				135					140					
Pro	Tyr	Leu	Trp	Ile	Lys	Trp	Ser	Pro	Pro	Thr	Leu	Ile	Asp	Leu	Lys	
145					150					155					160	
Thr	Gly	Trp	Phe	Thr	Leu	Leu	Tyr	Glu	Ile	Arg	Leu	Lys	Pro	Glu	Lys	
				165					170					175		
Ala	Ala	Glu	Trp	Glu	Ile	His	Phe	Ala	Gly	Gln	Gln	Thr	Glu	Phe	Lys	
			180					185					190			
Ile	Leu	Ser	Leu	His	Pro	Gly	Gln	Lys	Tyr	Leu	Val	Gln	Val	Arg	Cys	
		195					200					205				
Lys	Pro	Asp	His	Gly	Tyr	Trp	Ser	Ala	Trp	Ser	Pro	Ala	Thr	Phe	Ile	
	210					215					220					
Gln	Ile	Pro	Ser	Asp	Phe	Thr	Met	Asn	Asp	Thr	Thr	Val	Trp	Ile	Ser	
225					230					235					240	
Val	Ala	Val	Leu	Ser	Ala	Val	Ile	Cys	Leu	Ile	Ile	Val	Trp	Ala	Val	
				245					250					255		
Ala	Leu	Lys	Gly	Tyr	Ser	Met	Val	Thr	Cys	Ile	Phe	Pro	Pro	Val	Pro	
			260					265					270			
Gly	Pro	Lys	Ile	Lys	Gly	Phe	Asp	Ala	His	Leu	Leu	Glu	Lys	Gly	Lys	
		275					280					285				
Ser	Glu	Glu	Leu	Leu	Ser	Ala	Leu	Gly	Cys	Gln	Asp	Phe	Pro	Pro	Thr	
	290					295					300					
Ser	Asp	Tyr	Glu	Asp	Leu	Leu	Val	Glu	Tyr	Leu	Glu	Val	Asp	Asp	Ser	
305					310					315					320	
Glu	Asp	Gln	His	Leu	Met	Ser	Val	His	Ser	Lys	Glu	His	Pro	Ser	Gln	
				325					330					335		
Gly	Met	Lys	Pro	Thr	Tyr	Leu	Asp	Pro	Asp	Thr	Asp	Ser	Gly	Arg	Gly	
			340					345					350			
Ser	Cys	Asp	Ser	Pro	Ser	Leu	Leu	Ser	Glu	Lys	Cys	Glu	Glu	Pro	Gln	
		355					360					365				
Ala	Asn	Pro	Ser	Thr	Phe	Tyr	Asp	Pro	Glu	Val	Ile	Glu	Lys	Pro	Glu	
						375					380					
Asn	Pro	Glu	Thr	Thr	His	Thr	Trp	Asp	Pro	Gln	Cys	Ile	Ser	Met	Glu	
385					390					395					400	
Gly	Lys	Ile	Pro	Tyr	Phe	His	Ala	Gly	Gly	Ser	Lys	Cys	Ser	Thr	Trp	
				405					410						415	

-continued

Pro	Leu	Pro	Gln	Pro	Ser	Gln	His	Asn	Pro	Arg	Ser	Ser	Tyr	His	Asn	
			420					425					430			
Ile	Thr	Asp	Val	Cys	Glu	Leu	Ala	Val	Gly	Pro	Ala	Gly	Ala	Pro	Ala	
		435					440					445				
Thr	Leu	Leu	Asn	Glu	Ala	Gly	Lys	Asp	Ala	Leu	Lys	Ser	Ser	Gln	Thr	
	450					455					460					
Ile	Lys	Ser	Arg	Glu	Glu	Gly	Lys	Ala	Thr	Gln	Gln	Arg	Glu	Val	Glu	
465					470					475					480	
Ser	Phe	His	Ser	Glu	Thr	Asp	Gln	Asp	Thr	Pro	Trp	Leu	Leu	Pro	Gln	
				485					490					495		
Glu	Lys	Thr	Pro	Phe	Gly	Ser	Ala	Lys	Pro	Leu	Asp	Tyr	Val	Glu	Ile	
			500					505					510			
His	Lys	Val	Asn	Lys	Asp	Gly	Ala	Leu	Ser	Leu	Leu	Pro	Lys	Gln	Arg	
		515					520					525				
Glu	Asn	Ser	Gly	Lys	Pro	Lys	Lys	Pro	Gly	Thr	Pro	Glu	Asn	Asn	Lys	
	530					535					540					
Glu	Tyr	Ala	Lys	Val	Ser	Gly	Val	Met	Asp	Asn	Asn	Ile	Leu	Val	Leu	
545					550					555					560	
Val	Pro	Asp	Pro	His	Ala	Lys	Asn	Val	Ala	Cys	Phe	Glu	Glu	Ser	Ala	
				565					570					575		
Lys	Glu	Ala	Pro	Pro	Ser	Leu	Glu	Gln	Asn	Gln	Ala	Glu	Lys	Ala	Leu	
			580					585					590			
Ala	Asn	Phe	Thr	Ala	Thr	Ser	Ser	Lys	Cys	Arg	Leu	Gln	Leu	Gly	Gly	
		595					600					605				
Leu	Asp	Tyr	Leu	Asp	Pro	Ala	Cys	Phe	Thr	His	Ser	Phe	His			
	610						615				620					
<210> SEQ ID NO 2																
<211> LENGTH: 1205																
<212> TYPE: PRT																
<213> ORGANISM: Homo sapiens																
<400> SEQUENCE: 2																
Met	Asn	Gly	Tyr	Ala	Glu	Phe	Pro	Pro	Ser	Pro	Ser	Asn	Pro	Thr	Lys	
1				5					10					15		
Glu	Pro	Val	Glu	Pro	Gln	Pro	Ser	Gln	Val	Pro	Leu	Gln	Glu	Asp	Val	
		20					25						30			
Asp	Met	Ser	Ser	Gly	Ser	Ser	Gly	His	Glu	Thr	Asn	Glu	Asn	Cys	Ser	
	35						40					45				
Thr	Gly	Arg	Asp	Ser	Gln	Gly	Ser	Asp	Cys	Asp	Asp	Ser	Gly	Lys	Glu	
	50					55					60					
Leu	Gly	Met	Leu	Val	Glu	Pro	Pro	Asp	Ala	Arg	Gln	Ser	Pro	Asp	Thr	
65					70					75					80	
Phe	Ser	Leu	Met	Met	Ala	Lys	Ser	Glu	His	Asn	Pro	Ser	Thr	Ser	Gly	
			85						90					95		
Cys	Ser	Ser	Asp	Gln	Ser	Ser	Lys	Val	Asp	Thr	His	Lys	Glu	Leu	Ile	
		100						105					110			
Lys	Thr	Leu	Lys	Glu	Leu	Lys	Val	His	Leu	Pro	Ala	Asp	Lys	Lys	Ala	
		115					120					125				
Lys	Gly	Lys	Ala	Ser	Thr	Leu	Ala	Thr	Leu	Lys	Tyr	Ala	Leu	Arg	Ser	
	130					135					140					
Val	Lys	Gln	Val	Lys	Ala	Asn	Glu	Glu	Tyr	Tyr	Gln	Leu	Leu	Met	Ser	
145					150					155					160	
Ser	Glu	Gly	His	Pro	Cys	Gly	Ala	Asp	Val	Pro	Ser	Tyr	Thr	Val	Glu	

-continued

165																170																175															
Glu	Met	Glu	Ser	Val	Thr	Ser	Glu	His	Ile	Val	Lys	Asn	Ala	Asp	Met																																
			180						185						190																																
Phe	Ala	Val	Ala	Val	Ser	Leu	Val	Ser	Gly	Lys	Ile	Leu	Tyr	Ile	Ser																																
		195						200						205																																	
Asp	Gln	Val	Ala	Ser	Ile	Phe	His	Cys	Lys	Arg	Asp	Ala	Phe	Ser	Asp																																
		210						215						220																																	
Ala	Lys	Phe	Val	Glu	Phe	Leu	Ala	Pro	His	Asp	Val	Gly	Val	Phe	His																																
		225						230						235						240																											
Ser	Phe	Thr	Ser	Pro	Tyr	Lys	Leu	Pro	Leu	Trp	Ser	Met	Cys	Ser	Gly																																
			245						250						255																																
Ala	Asp	Ser	Phe	Thr	Gln	Glu	Cys	Met	Glu	Glu	Lys	Ser	Phe	Phe	Cys																																
			260						265						270																																
Arg	Val	Ser	Val	Arg	Lys	Ser	His	Glu	Asn	Glu	Ile	Arg	Tyr	His	Pro																																
		275						280						285																																	
Phe	Arg	Met	Thr	Pro	Tyr	Leu	Val	Lys	Val	Arg	Asp	Gln	Gln	Gly	Ala																																
		290						295						300																																	
Glu	Ser	Gln	Leu	Cys	Cys	Leu	Leu	Leu	Ala	Glu	Arg	Val	His	Ser	Gly																																
		305						310						315						320																											
Tyr	Glu	Ala	Pro	Arg	Ile	Pro	Pro	Glu	Lys	Arg	Ile	Phe	Thr	Thr	Thr																																
			325						330						335																																
His	Thr	Pro	Asn	Cys	Leu	Phe	Gln	Asp	Val	Asp	Glu	Arg	Ala	Val	Pro																																
			340						345						350																																
Leu	Leu	Gly	Tyr	Leu	Pro	Gln	Asp	Leu	Ile	Glu	Thr	Pro	Val	Leu	Val																																
		355						360						365																																	
Gln	Leu	His	Pro	Ser	Asp	Arg	Pro	Leu	Met	Leu	Ala	Ile	His	Lys	Lys																																
		370						375						380																																	
Ile	Leu	Gln	Ser	Gly	Gly	Gln	Pro	Phe	Asp	Tyr	Ser	Pro	Ile	Arg	Phe																																
		385						390						395						400																											
Arg	Ala	Arg	Asn	Gly	Glu	Tyr	Ile	Thr	Leu	Asp	Thr	Ser	Trp	Ser	Ser																																
			405						410						415																																
Phe	Ile	Asn	Pro	Trp	Ser	Arg	Lys	Ile	Ser	Phe	Ile	Ile	Gly	Arg	His																																
			420						425						430																																
Lys	Val	Arg	Val	Gly	Pro	Leu	Asn	Glu	Asp	Val	Phe	Ala	Ala	His	Pro																																
		435						440						445																																	
Cys	Thr	Glu	Glu	Lys	Ala	Leu	His	Pro	Ser	Ile	Gln	Glu	Leu	Thr	Glu																																
		450						455						460																																	
Gln	Ile	His	Arg	Leu	Leu	Leu	Gln	Pro	Val	Pro	His	Ser	Gly	Ser	Ser																																
		465						470						475						480																											
Gly	Tyr	Gly	Ser	Leu	Gly	Ser	Asn	Gly	Ser	His	Glu	His	Leu	Met	Ser																																
			485						490						495																																
Gln	Thr	Ser	Ser	Ser	Asp	Ser	Asn	Gly	His	Glu	Asp	Ser	Arg	Arg	Arg																																
			500						505						510																																
Arg	Ala	Glu	Ile	Cys	Lys	Asn	Gly	Asn	Lys	Thr	Lys	Asn	Arg	Ser	His																																
		515						520						525																																	
Tyr	Ser	His	Glu	Ser	Gly	Glu	Gln	Lys	Lys	Lys	Ser	Val	Thr	Glu	Met																																
		530						535						540																																	
Gln	Thr	Asn	Pro	Pro	Ala	Glu	Lys	Lys	Ala	Val	Pro	Ala	Met	Glu	Lys																																
		545						550						555						560																											
Asp	Ser	Leu	Gly	Val	Ser	Phe	Pro	Glu	Glu	Leu	Ala	Cys	Lys	Asn	Gln																																
			565						570						575																																
Pro	Thr	Cys	Ser	Tyr	Gln	Gln	Ile	Ser	Cys	Leu	Asp	Ser	Val	Ile	Arg																																
			580						585						590																																

-continued

Tyr	Leu	Glu	Ser	Cys	Asn	Glu	Ala	Ala	Thr	Leu	Lys	Arg	Lys	Cys	Glu	
	595						600					605				
Phe	Pro	Ala	Asn	Val	Pro	Ala	Leu	Arg	Ser	Ser	Asp	Lys	Arg	Lys	Ala	
	610					615					620					
Thr	Val	Ser	Pro	Gly	Pro	His	Ala	Gly	Glu	Ala	Glu	Pro	Pro	Ser	Arg	
625					630					635					640	
Val	Asn	Ser	Arg	Thr	Gly	Val	Gly	Thr	His	Leu	Thr	Ser	Leu	Ala	Leu	
				645					650					655		
Pro	Gly	Lys	Ala	Glu	Ser	Val	Ala	Ser	Leu	Thr	Ser	Gln	Cys	Ser	Tyr	
			660					665					670			
Ser	Ser	Thr	Ile	Val	His	Val	Gly	Asp	Lys	Lys	Pro	Gln	Pro	Glu	Leu	
		675					680					685				
Glu	Met	Val	Glu	Asp	Ala	Ala	Ser	Gly	Pro	Glu	Ser	Leu	Asp	Cys	Leu	
	690					695					700					
Ala	Gly	Pro	Ala	Leu	Ala	Cys	Gly	Leu	Ser	Gln	Glu	Lys	Glu	Pro	Phe	
705					710					715					720	
Lys	Lys	Leu	Gly	Leu	Thr	Lys	Glu	Val	Leu	Ala	Ala	His	Thr	Gln	Lys	
			725						730					735		
Glu	Glu	Gln	Ser	Phe	Leu	Gln	Lys	Phe	Lys	Glu	Ile	Arg	Lys	Leu	Ser	
			740					745					750			
Ile	Phe	Gln	Ser	His	Cys	His	Tyr	Tyr	Leu	Gln	Glu	Arg	Ser	Lys	Gly	
		755					760					765				
Gln	Pro	Ser	Glu	Arg	Thr	Ala	Pro	Gly	Leu	Arg	Asn	Thr	Ser	Gly	Ile	
		770				775					780					
Asp	Ser	Pro	Trp	Lys	Lys	Thr	Gly	Lys	Asn	Arg	Lys	Leu	Lys	Ser	Lys	
785					790					795					800	
Arg	Val	Lys	Pro	Arg	Asp	Ser	Ser	Glu	Ser	Thr	Gly	Ser	Gly	Gly	Pro	
				805					810					815		
Val	Ser	Ala	Arg	Pro	Pro	Leu	Val	Gly	Leu	Asn	Ala	Thr	Ala	Trp	Ser	
			820					825					830			
Pro	Ser	Asp	Thr	Ser	Gln	Ser	Ser	Cys	Pro	Ala	Val	Pro	Phe	Pro	Ala	
		835					840					845				
Pro	Val	Pro	Ala	Ala	Tyr	Ser	Leu	Pro	Val	Phe	Pro	Ala	Pro	Gly	Thr	
		850				855					860					
Val	Ala	Ala	Pro	Pro	Ala	Pro	Pro	His	Ala	Ser	Phe	Thr	Val	Pro	Ala	
865					870					875					880	
Val	Pro	Val	Asp	Leu	Gln	His	Gln	Phe	Ala	Val	Gln	Pro	Pro	Pro	Phe	
			885					890						895		
Pro	Ala	Pro	Leu	Ala	Pro	Val	Met	Ala	Phe	Met	Leu	Pro	Ser	Tyr	Ser	
			900					905					910			
Phe	Pro	Ser	Gly	Thr	Pro	Asn	Leu	Pro	Gln	Ala	Phe	Phe	Pro	Ser	Gln	
		915					920					925				
Pro	Gln	Phe	Pro	Ser	His	Pro	Thr	Leu	Thr	Ser	Glu	Met	Ala	Ser	Ala	
		930				935					940					
Ser	Gln	Pro	Glu	Phe	Pro	Glu	Gly	Gly	Thr	Gly	Ala	Met	Gly	Thr	Thr	
945					950					955					960	
Gly	Ala	Thr	Glu	Thr	Ala	Ala	Val	Gly	Ala	Asp	Cys	Lys	Pro	Gly	Thr	
			965					970						975		
Ser	Arg	Asp	Gln	Gln	Pro	Lys	Ala	Pro	Leu	Thr	Arg	Asp	Glu	Pro	Ser	
			980					985					990			
Asp	Thr	Gln	Asn	Ser	Asp	Ala	Leu	Ser	Thr	Ser	Ser	Gly	Leu	Leu	Asn	
		995					1000					1005				

-continued

Leu	Leu	Leu	Asn	Glu	Asp	Leu	Cys	Ser	Ala	Ser	Gly	Ser	Ala	Ala
1010						1015					1020			
Ser	Glu	Ser	Leu	Gly	Ser	Gly	Ser	Leu	Gly	Cys	Asp	Ala	Ser	Pro
1025						1030					1035			
Ser	Gly	Ala	Gly	Ser	Ser	Asp	Thr	Ser	His	Thr	Ser	Lys	Tyr	Phe
1040						1045					1050			
Gly	Ser	Ile	Asp	Ser	Ser	Glu	Asn	Asn	His	Lys	Ala	Lys	Met	Asn
1055						1060					1065			
Thr	Gly	Met	Glu	Glu	Ser	Glu	His	Phe	Ile	Lys	Cys	Val	Leu	Gln
1070						1075					1080			
Asp	Pro	Ile	Trp	Leu	Leu	Met	Ala	Asp	Ala	Asp	Ser	Ser	Val	Met
1085						1090					1095			
Met	Thr	Tyr	Gln	Leu	Pro	Ser	Arg	Asn	Leu	Glu	Ala	Val	Leu	Lys
1100						1105					1110			
Glu	Asp	Arg	Glu	Lys	Leu	Lys	Leu	Leu	Gln	Lys	Leu	Gln	Pro	Arg
1115						1120					1125			
Phe	Thr	Glu	Ser	Gln	Lys	Gln	Glu	Leu	Arg	Glu	Val	His	Gln	Trp
1130						1135					1140			
Met	Gln	Thr	Gly	Gly	Leu	Pro	Ala	Ala	Ile	Asp	Val	Ala	Glu	Cys
1145						1150					1155			
Val	Tyr	Cys	Glu	Asn	Lys	Glu	Lys	Gly	Asn	Ile	Cys	Ile	Pro	Tyr
1160						1165					1170			
Glu	Glu	Asp	Ile	Pro	Ser	Leu	Gly	Leu	Ser	Glu	Val	Ser	Asp	Thr
1175						1180					1185			
Lys	Glu	Asp	Glu	Asn	Gly	Ser	Pro	Leu	Asn	His	Arg	Ile	Glu	Glu
1190						1195					1200			
Gln	Thr													
1205														
<210> SEQ ID NO 3														
<211> LENGTH: 376														
<212> TYPE: PRT														
<213> ORGANISM: Homo sapiens														
<400> SEQUENCE: 3														
Met	Lys	Glu	Asn	Val	Ala	Ser	Ala	Thr	Val	Phe	Thr	Leu	Leu	Leu
1			5					10					15	
Leu	Asn	Thr	Cys	Leu	Leu	Asn	Gly	Gln	Leu	Pro	Pro	Gly	Lys	Pro
		20						25					30	Glu
Ile	Phe	Lys	Cys	Arg	Ser	Pro	Asn	Lys	Glu	Thr	Phe	Thr	Cys	Trp
		35					40					45		Trp
Arg	Pro	Gly	Thr	Asp	Gly	Gly	Leu	Pro	Thr	Asn	Tyr	Ser	Leu	Thr
	50					55					60			Tyr
His	Arg	Glu	Gly	Glu	Thr	Leu	Met	His	Glu	Cys	Pro	Asp	Tyr	Ile
65					70					75				80
Gly	Gly	Pro	Asn	Ser	Cys	His	Phe	Gly	Lys	Gln	Tyr	Thr	Ser	Met
			85						90					95
Arg	Thr	Tyr	Ile	Met	Met	Val	Asn	Ala	Thr	Asn	Gln	Met	Gly	Ser
		100						105					110	Ser
Phe	Ser	Asp	Glu	Leu	Tyr	Val	Asp	Val	Thr	Tyr	Ile	Val	Gln	Pro
		115					120					125		Asp
Pro	Pro	Leu	Glu	Leu	Ala	Val	Glu	Val	Lys	Gln	Pro	Glu	Asp	Arg
	130					135					140			Lys
Pro	Tyr	Leu	Trp	Ile	Lys	Trp	Ser	Pro	Pro	Thr	Leu	Ile	Asp	Leu
145					150					155				160

-continued

Thr	Gly	Trp	Phe	Thr	Leu	Leu	Tyr	Glu	Ile	Arg	Leu	Lys	Pro	Glu	Lys	
				165					170					175		
Ala	Ala	Glu	Trp	Glu	Ile	His	Phe	Ala	Gly	Gln	Gln	Thr	Glu	Phe	Lys	
			180					185					190			
Ile	Leu	Ser	Leu	His	Pro	Gly	Gln	Lys	Tyr	Leu	Val	Gln	Val	Arg	Cys	
		195					200					205				
Lys	Pro	Asp	His	Gly	Tyr	Trp	Ser	Ala	Trp	Ser	Pro	Ala	Thr	Phe	Ile	
	210					215					220					
Gln	Ile	Pro	Ser	Asp	Phe	Thr	Met	Asn	Asp	Thr	Thr	Val	Trp	Ile	Ser	
225					230					235					240	
Val	Ala	Val	Leu	Ser	Ala	Val	Ile	Cys	Leu	Ile	Ile	Val	Trp	Ala	Val	
				245					250					255		
Ala	Leu	Lys	Gly	Tyr	Ser	Met	Val	Thr	Cys	Ile	Phe	Pro	Pro	Val	Pro	
			260					265					270			
Gly	Pro	Lys	Ile	Lys	Gly	Phe	Asp	Ala	His	Leu	Leu	Glu	Lys	Gly	Lys	
		275					280					285				
Ser	Glu	Glu	Leu	Leu	Ser	Ala	Leu	Gly	Cys	Gln	Asp	Phe	Pro	Pro	Thr	
	290					295					300					
Ser	Asp	Tyr	Glu	Asp	Leu	Leu	Val	Glu	Tyr	Leu	Glu	Val	Asp	Asp	Ser	
305					310					315					320	
Glu	Asp	Gln	His	Leu	Met	Ser	Val	His	Ser	Lys	Glu	His	Pro	Ser	Gln	
			325						330					335		
Gly	Asp	Pro	Leu	Met	Leu	Gly	Ala	Ser	His	Tyr	Lys	Asn	Leu	Lys	Ser	
		340						345					350			
Tyr	Arg	Pro	Arg	Lys	Ile	Ser	Ser	Gln	Gly	Arg	Leu	Ala	Val	Phe	Thr	
		355				360						365				
Lys	Ala	Thr	Leu	Thr	Thr	Val	Gln									
	370					375										
<210> SEQ ID NO 4																
<211> LENGTH: 1163																
<212> TYPE: PRT																
<213> ORGANISM: Artificial Sequence																
<220> FEATURE:																
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic polypeptide																
<400> SEQUENCE: 4																
Met	Ala	Gly	Trp	Ile	Gln	Ala	Gln	Gln	Leu	Gln	Gly	Asp	Ala	Leu	Arg	
1				5					10					15		
Gln	Met	Gln	Val	Leu	Tyr	Gly	Gln	His	Phe	Pro	Ile	Glu	Val	Arg	His	
			20					25					30			
Tyr	Leu	Ala	Gln	Trp	Ile	Glu	Ser	Gln	Pro	Trp	Asp	Ala	Ile	Asp	Leu	
		35					40					45				
Asp	Asn	Pro	Gln	Asp	Arg	Ala	Gln	Ala	Thr	Gln	Leu	Leu	Glu	Gly	Leu	
	50					55					60					
Val	Gln	Glu	Leu	Gln	Lys	Lys	Ala	Glu	His	Gln	Val	Gly	Glu	Asp	Gly	
65					70					75					80	
Phe	Leu	Leu	Lys	Ile	Lys	Leu	Gly	His	Tyr	Ala	Thr	Gln	Leu	Gln	Lys	
			85						90					95		
Thr	Tyr	Asp	Arg	Cys	Pro	Leu	Glu	Leu	Val	Arg	Cys	Ile	Arg	His	Ile	
		100						105					110			
Leu	Tyr	Asn	Glu	Gln	Arg	Leu	Val	Arg	Glu	Ala	Asn	Asn	Cys	Ser	Ser	
		115					120					125				
Pro	Ala	Gly	Ile	Leu	Val	Asp	Ala	Met	Ser	Gln	Lys	His	Leu	Gln	Ile	

-continued

130				135				140							
Asn	Gln	Thr	Phe	Glu	Glu	Leu	Arg	Leu	Val	Thr	Gln	Asp	Thr	Glu	Asn
145				150						155				160	
Glu	Leu	Lys	Lys	Leu	Gln	Gln	Thr	Gln	Glu	Tyr	Phe	Ile	Ile	Gln	Tyr
				165					170					175	
Gln	Glu	Ser	Leu	Arg	Ile	Gln	Ala	Gln	Phe	Ala	Gln	Leu	Ala	Gln	Leu
			180					185					190		
Ser	Pro	Gln	Glu	Arg	Leu	Ser	Arg	Glu	Thr	Ala	Leu	Gln	Gln	Lys	Gln
		195					200					205			
Val	Ser	Leu	Glu	Ala	Trp	Leu	Gln	Arg	Glu	Ala	Gln	Thr	Leu	Gln	Gln
	210					215					220				
Tyr	Arg	Val	Glu	Leu	Ala	Glu	Lys	His	Gln	Lys	Thr	Leu	Gln	Leu	Leu
225					230					235					240
Arg	Lys	Gln	Gln	Thr	Ile	Ile	Leu	Asp	Asp	Glu	Leu	Ile	Gln	Trp	Lys
				245					250					255	
Arg	Arg	Gln	Gln	Leu	Ala	Gly	Asn	Gly	Gly	Pro	Pro	Glu	Gly	Ser	Leu
			260					265					270		
Asp	Val	Leu	Gln	Ser	Trp	Cys	Glu	Lys	Leu	Ala	Glu	Ile	Ile	Trp	Gln
	275						280					285			
Asn	Arg	Gln	Gln	Ile	Arg	Arg	Ala	Glu	His	Leu	Cys	Gln	Gln	Leu	Pro
	290					295					300				
Ile	Pro	Gly	Pro	Val	Glu	Glu	Met	Leu	Ala	Glu	Val	Asn	Ala	Thr	Ile
305					310					315					320
Thr	Asp	Ile	Ile	Ser	Ala	Leu	Val	Thr	Ser	Thr	Phe	Ile	Ile	Glu	Lys
				325					330					335	
Gln	Pro	Pro	Gln	Val	Leu	Lys	Thr	Gln	Thr	Lys	Phe	Ala	Ala	Thr	Val
			340					345					350		
Arg	Leu	Leu	Val	Gly	Gly	Lys	Leu	Asn	Val	His	Met	Asn	Pro	Pro	Gln
		355					360					365			
Val	Lys	Ala	Thr	Ile	Ile	Ser	Glu	Gln	Gln	Ala	Lys	Ser	Leu	Leu	Lys
	370					375					380				
Asn	Glu	Asn	Thr	Arg	Asn	Glu	Cys	Ser	Gly	Glu	Ile	Leu	Asn	Asn	Cys
385					390					395					400
Cys	Val	Met	Glu	Tyr	His	Gln	Ala	Thr	Gly	Thr	Leu	Ser	Ala	His	Phe
			405						410					415	
Arg	Asn	Met	Ser	Leu	Lys	Arg	Ile	Lys	Arg	Ala	Asp	Arg	Arg	Gly	Ala
			420					425					430		
Glu	Ser	Val	Thr	Glu	Glu	Lys	Phe	Thr	Val	Leu	Phe	Glu	Ser	Gln	Phe
		435					440					445			
Ser	Val	Gly	Ser	Asn	Glu	Leu	Val	Phe	Gln	Val	Lys	Thr	Leu	Ser	Leu
	450					455					460				
Pro	Trp	Val	Ile	Val	His	Gly	Ser	Gln	Asp	His	Asn	Ala	Thr	Ala	Thr
465					470					475					480
Val	Leu	Trp	Asp	Asn	Ala	Phe	Ala	Glu	Pro	Gly	Arg	Val	Pro	Phe	Ala
			485						490					495	
Val	Pro	Asp	Lys	Val	Leu	Trp	Pro	Gln	Leu	Cys	Glu	Ala	Leu	Asn	Met
			500					505					510		
Lys	Phe	Lys	Ala	Glu	Val	Gln	Ser	Asn	Arg	Gly	Leu	Thr	Lys	Glu	Asn
		515					520					525			
Leu	Val	Phe	Leu	Ala	Gln	Lys	Leu	Phe	Asn	Asn	Ser	Ser	Ser	His	Leu
	530					535					540				
Glu	Asp	Tyr	Ser	Gly	Leu	Ser	Val	Ser	Trp	Ser	Gln	Phe	Asn	Arg	Glu
545					550					555					560

-continued

Asn	Leu	Pro	Gly	Trp	Asn	Tyr	Thr	Phe	Trp	Gln	Trp	Phe	Asp	Gly	Val	
			565						570					575		
Met	Glu	Val	Leu	Lys	Lys	His	His	Lys	Pro	His	Trp	Asn	Asp	Gly	Ala	
			580					585					590			
Ile	Leu	Gly	Phe	Val	Asn	Lys	Gln	Gln	Ala	His	Asp	Leu	Leu	Ile	Asn	
		595					600					605				
Lys	Pro	Asp	Gly	Thr	Phe	Leu	Leu	Arg	Phe	Ser	Asp	Ser	Glu	Ile	Gly	
	610					615					620					
Gly	Ile	Thr	Ile	Ala	Trp	Lys	Phe	Asp	Ser	Pro	Glu	Arg	Asn	Leu	Trp	
625					630					635					640	
Asn	Leu	Lys	Pro	Phe	Thr	Thr	Arg	Asp	Phe	Ser	Ile	Arg	Ser	Leu	Ala	
				645					650						655	
Asp	Arg	Leu	Gly	Asp	Leu	Ser	Tyr	Leu	Ile	Tyr	Val	Phe	Pro	Asp	Arg	
			660					665						670		
Pro	Lys	Asp	Glu	Val	Phe	Ser	Lys	Tyr	Tyr	Thr	Pro	Val	Leu	Ala	Lys	
		675					680					685				
Ala	Val	Asp	Gly	Tyr	Val	Lys	Pro	Gln	Ile	Lys	Gln	Trp	Pro	Glu	Phe	
		690				695					700					
Val	Asn	Ala	Ser	Ala	Asp	Ala	Gly	Gly	Ser	Ser	Ala	Thr	Tyr	Met	Asp	
705					710					715					720	
Gln	Ala	Pro	Ser	Pro	Ala	Val	Cys	Pro	Gln	Ala	Pro	Tyr	Asn	Met	Tyr	
				725					730					735		
Pro	Gln	Asn	Pro	Asp	His	Val	Leu	Asp	Gln	Asp	Gly	Glu	Phe	Asp	Leu	
			740					745					750			
Asp	Glu	Thr	Met	Asp	Val	Ala	Arg	His	Val	Glu	Glu	Leu	Leu	Arg	Arg	
		755					760					765				
Pro	Met	Asp	Ser	Leu	Asp	Ser	Arg	Leu	Ser	Pro	Pro	Ala	Gly	Leu	Phe	
	770					775					780					
Thr	Ser	Ala	Arg	Gly	Ser	Leu	Ser	Leu	Asp	Ser	Gln	Arg	Lys	Leu	Gln	
785					790					795					800	
Phe	Tyr	Glu	Asp	Arg	His	Gln	Leu	Pro	Ala	Pro	Lys	Trp	Ala	Glu	Leu	
			805						810					815		
Ala	Asn	Leu	Ile	Asn	Asn	Cys	Met	Asp	Tyr	Glu	Pro	Asp	Phe	Arg	Pro	
			820					825					830			
Ser	Phe	Arg	Ala	Ile	Ile	Arg	Asp	Leu	Asn	Ser	Leu	Phe	Thr	Pro	Asp	
		835					840					845				
Tyr	Glu	Leu	Leu	Thr	Glu	Asn	Asp	Met	Leu	Pro	Asn	Met	Arg	Ile	Gly	
	850					855					860					
Ala	Leu	Gly	Phe	Ser	Gly	Ala	Phe	Glu	Asp	Arg	Asp	Pro	Thr	Gln	Phe	
865					870					875					880	
Glu	Glu	Arg	His	Leu	Lys	Phe	Leu	Gln	Gln	Leu	Gly	Lys	Gly	Asn	Phe	
				885					890					895		
Gly	Ser	Val	Glu	Met	Cys	Arg	Tyr	Asp	Pro	Leu	Gln	Asp	Asn	Thr	Gly	
		900						905					910			
Glu	Trp	Ala	Val	Lys	Lys	Leu	Gln	His	Ser	Thr	Glu	Glu	His	Leu	Arg	
		915					920					925				
Asp	Phe	Glu	Arg	Glu	Ile	Glu	Ile	Leu	Lys	Ser	Leu	Gln	His	Asp	Asn	
		930				935						940				
Ile	Val	Lys	Tyr	Lys	Gly	Val	Cys	Tyr	Ser	Ala	Gly	Arg	Arg	Asn	Leu	
945					950					955					960	
Lys	Leu	Ile	Met	Glu	Tyr	Leu	Pro	Tyr	Gly	Ser	Leu	Arg	Asp	Tyr	Leu	
				965					970					975		

-continued

Gln	Lys	His	Lys	Glu	Arg	Ile	Asp	His	Ile	Lys	Leu	Leu	Gln	Tyr	Thr
			980					985					990		
Ser	Gln	Ile	Cys	Lys	Gly	Met	Glu	Tyr	Leu	Gly	Thr	Lys	Arg	Tyr	Ile
			995				1000					1005			
His	Arg	Asp	Leu	Ala	Thr	Arg	Asn	Ile	Leu	Val	Glu	Asn	Glu	Asn	
	1010						1015				1020				
Arg	Val	Lys	Ile	Gly	Asp	Phe	Gly	Leu	Thr	Lys	Val	Leu	Pro	Gln	
	1025					1030					1035				
Asp	Lys	Glu	Tyr	Tyr	Lys	Val	Lys	Glu	Pro	Gly	Glu	Ser	Pro	Ile	
	1040					1045					1050				
Phe	Trp	Tyr	Ala	Pro	Glu	Ser	Leu	Thr	Glu	Ser	Lys	Phe	Ser	Val	
	1055					1060					1065				
Ala	Ser	Asp	Val	Trp	Ser	Phe	Gly	Trp	Leu	Tyr	Glu	Leu	Phe	Thr	
	1070					1075					1080				
Tyr	Ile	Glu	Lys	Ser	Lys	Ser	Pro	Pro	Ala	Glu	Phe	Met	Arg	Met	
	1085					1090					1095				
Ile	Gly	Asn	Asp	Lys	Gln	Gly	Gln	Met	Ile	Val	Phe	His	Leu	Ile	
	1100					1105					1110				
Glu	Leu	Leu	Lys	Asn	Asn	Gly	Arg	Leu	Pro	Arg	Pro	Asp	Gly	Cys	
	1115					1120					1125				
Pro	Asp	Glu	Ile	Tyr	Met	Ile	Met	Thr	Glu	Cys	Trp	Asn	Asn	Asn	
	1130					1135					1140				
Val	Asn	Gln	Arg	Pro	Ser	Phe	Arg	Asp	Leu	Ala	Leu	Arg	Val	Asp	
	1145					1150					1155				
Gln	Ile	Arg	Asp	Asn											
	1160														

- What is claimed is:
1. A method of producing an isolated milk product comprising secretory IgA (sIgA) from cultured mammary cells and plasma cells, the method comprising:
- a. culturing a live cell construct in a bioreactor under conditions which produce the milk product, said cell construct comprising:
 - i. a three-dimensional scaffold having an exterior surface, an interior surface defining an interior cavity, and a plurality of pores extending from the interior surface to the exterior surface;
 - ii. a matrix material disposed on the exterior surface of the three-dimensional scaffold;
 - iii. a culture medium disposed within the interior cavity and in fluidic contact with the interior surface;
 - iv. a plurality of live plasma cells disposed on the matrix material; and
 - v. a confluent monolayer of polarized mammary cells disposed on the plurality of plasma cells, wherein the mammary cells are selected from the group consisting of: live mammary epithelial cells, and live mammary myoepithelial cells, wherein the polarized mammary cells comprise an apical surface from which the milk product is secreted and a basal surface;
- said bioreactor comprising an apical compartment that is in fluidic contact with the apical surface of the live mammary cells, is substantially isolated from the interior cavity of the live cell construct, and is substantially free of cell culture medium; and
- b. isolating the milk product secreted into the apical compartment from the apical surface of the live mammary cells and the plasma cells.
2. The method of claim 1, wherein the monolayer of polarized mammary cells is at least 70% confluent, at least 80% confluent, at least 90% confluent, at least 95% confluent, at least 99% confluent, or 100% confluent.
3. The method of claim 1, wherein at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or 100% of the mammary cells are polarized in the same orientation.
4. The method of claim 1, wherein total cell density of mammary cells within the bioreactor is at least 10¹¹; and alternatively wherein total surface area of mammary cells within the bioreactor is at least 1.5 m².
5. The method of claim 1, wherein total cell density of plasma cells in the bioreactor is about 200 to 500 plasma cells per mm².
6. The method of claim 1, wherein the culturing is carried out at a temperature of about 27° C. to about 39° C.
7. The method of claim 1, wherein the culturing is carried out at an atmospheric concentration of CO₂ of about 4% to about 6%.
8. A live cell construct, comprising:
- a. a three dimensional scaffold having an exterior surface, an interior surface defining an interior cavity/basal chamber, and a plurality of pores extending from the interior surface to the exterior surface;
 - b. a matrix material disposed on the exterior surface of the three-dimensional scaffold;
 - c. a plurality of live plasma cells disposed on the matrix material;

- d. an at least 70% confluent monolayer of live polarized mammary cells disposed on the plurality of live plasma cells and on the matrix material, wherein the live mammary cells are selected from the group consisting of: live mammary epithelial cells, and live mammary myoepithelial cells, and wherein the live polarized mammary cells comprise an apical surface from which the milk product is secreted and a basal surface; and
 - e. a culture medium disposed within the interior cavity/basal chamber and in fluidic contact with the interior surface, wherein the culture medium is substantially isolated from the apical surface of the live mammary cells.
9. The cell construct of claim 8, wherein the basal surface of the mammary cells is in fluidic contact with the culture medium.
10. The cell construct of claim 8, wherein at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or 100% of the mammary cells are polarized in the same orientation.
11. The cell construct of claim 8, wherein the monolayer of polarized mammary cells is at least 70% confluent, at least 80% confluent, at least 90% confluent, at least 95% confluent, at least 99% confluent, or 100% confluent.
12. The cell construct of claim 8, wherein the mammary cells comprise a constitutively active prolactin receptor protein.
13. The cell construct of claim 8, wherein the culture medium comprises prolactin.

* * * * *